

# Hydrogen Sulfide in the Mammalian Cardiovascular System

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## Abstract

For more than a century, hydrogen sulfide (H<sub>2</sub>S) has been regarded as a toxic gas. This review surveys the growing recognition of the role of H<sub>2</sub>S as an endogenous signaling molecule in mammals, with emphasis on its physiological and pathological pathways in the cardiovascular system. In biological fluids, H<sub>2</sub>S gas is a weak acid that exists as about 15% H<sub>2</sub>S, 85% HS<sup>−</sup>, and a trace of S<sup>2−</sup>. Here, we use “H<sub>2</sub>S” to refer to this mixture. H<sub>2</sub>S has been found to influence heart contractile functions and may serve as a cardioprotectant for treating ischemic heart diseases and heart failure. Alterations of the endogenous H<sub>2</sub>S level have been found in animal models with various pathological conditions such as myocardial ischemia, spontaneous hypertension, and hypoxic pulmonary hypertension. In the vascular system, H<sub>2</sub>S exerts biphasic regulation of a vascular tone with varying effects based on its concentration and in the presence of nitric oxide. Over the past decade, several H<sub>2</sub>S-releasing compounds (NaHS, Na<sub>2</sub>S, GYY4137, *etc.*) have been utilized to test the effect of exogenous H<sub>2</sub>S under different physiological and pathological situations *in vivo* and *in vitro*. H<sub>2</sub>S has been found to promote angiogenesis and to protect against atherosclerosis and hypertension, while excess H<sub>2</sub>S may promote inflammation in septic or hemorrhagic shock. H<sub>2</sub>S-releasing compounds and inhibitors of H<sub>2</sub>S synthesis hold promise in alleviating specific disease conditions. This comprehensive review covers in detail the effects of H<sub>2</sub>S on the cardiovascular system, especially in disease situations, and also the various underlying mechanisms. *Antioxid. Redox Signal.* 17, 141–185.

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## I. Introduction

**H**YDROGEN SULFIDE (H<sub>2</sub>S) has long been known as a toxic gas, and many reports on fatal intoxication by H<sub>2</sub>S have been documented (27, 106, 229). Humans can smell <0.1 ppm of H<sub>2</sub>S in the air, and at 3–10 ppm, it has a very unpleasant odor (203). Above 50 ppm, H<sub>2</sub>S irritates the eyes and respiratory tract, and mice inhaling 80 ppm H<sub>2</sub>S at a low environmental temperature go into a reversible hibernation-like state with reduced metabolism and breathing rate (19). This effect is species dependent, as 80 ppm H<sub>2</sub>S has no effect on 6 kg piglets (131), while 100 ppm of it kills canaries and guinea pigs (203). Above 500 ppm, H<sub>2</sub>S may cause unconsciousness and death in humans (203). H<sub>2</sub>S intoxication is often attributed to its potent, reversible inhibition of cytochrome *c* oxidase, thus blocking oxidative phosphorylation (12, 58, 203). The inhibition of other enzymes, such as carbonic anhydrase (178), monoamine oxidase (266), Na<sup>+</sup>/K<sup>+</sup>-ATPase, and cholinesterase (203), also contributes toward its toxicity.

The physiological importance of H<sub>2</sub>S became recognized in the last one and a half decades; starting when Abe and Kimura reported in 1996 that H<sub>2</sub>S acts as a novel neuromodulator (1). Now, H<sub>2</sub>S is accepted as the third “gasotransmitter” after nitric oxide (NO) and carbon monoxide (CO) (257). Similar to NO and CO, H<sub>2</sub>S is endogenously generated by several enzymes and has been demonstrated as influencing a wide range of physiological and pathological processes, including blood vessel relaxation (3, 47, 91, 116, 268, 288, 308, 309), arterial contraction (3, 116, 141, 268), neurotransmission (1), regulation of inflammation (95, 132), cardioprotection (61, 104), neuroprotection, neurotoxicity (93, 112), and insulin release (4). The role of H<sub>2</sub>S has been demonstrated in the arteries of many species, including humans (268).

The molecular mechanisms underlying the biological actions of H<sub>2</sub>S have remained elusive, but a recent article published in *Science Signaling* suggests that one of the key mechanisms may be that H<sub>2</sub>S S-sulfhydrates proteins by converting cysteine-SH groups to –SSH (173). This S-sulfhydration occurs in many different proteins due to the action of endogenously produced H<sub>2</sub>S, and it results in modifying the physiological functions of the proteins. Thus, post-translational modification by H<sub>2</sub>S such as S-sulfhydration, similar to S-nitrosylation induced by NO, may be an important signaling mechanism underlying its diverse effects on the cardiovascular system (173). Several molecules have been proposed as being the potential targets of H<sub>2</sub>S action, such as the adenonsine triphosphate (ATP)-sensitive potassium (K<sub>ATP</sub>) channels (309), adenylyl cyclase (AC) (1, 113), mitogen-activated protein kinases (MAPKs) (95), and nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) (136, 313).

There have been many reviews on the physiological and pathological effects of H<sub>2</sub>S in the past decade (15, 34, 57, 63, 93, 114, 126, 132, 134, 135, 149, 150, 153, 171, 195, 196, 199, 240, 244, 257, 273, 275, 276). In addition, two Forum issues (9 and 10) of volume 12 of ARS (2010) were devoted to many aspects of the biology of H<sub>2</sub>S, including eight experimental reports and seven reviews (16, 34, 115, 142, 183, 253, 258). Our review will focus on recent studies conducted on the role of H<sub>2</sub>S actions in the cardiovascular system, with special emphasis on heart and vascular diseases.

## II. Biochemistry of H<sub>2</sub>S

### A. Physical and chemical properties of H<sub>2</sub>S and its free and stored concentrations in mammals

H<sub>2</sub>S is a colorless, flammable, and water-soluble gas with a strong characteristic rotten egg smell. In water, H<sub>2</sub>S is a weak acid that dissociates to form H<sup>+</sup>, HS<sup>−</sup>, and S<sup>2−</sup> (203). Figure 1 shows the origins and disposal routes for H<sub>2</sub>S and illustrates how free sulfide exists as H<sub>2</sub>S, HS<sup>−</sup>, and S<sup>2−</sup> in body fluids. Earlier papers have often suggested that at pH 7.4, about one third of “H<sub>2</sub>S” exists as the dissolved gas, H<sub>2</sub>S, while the other two thirds are HS<sup>−</sup> plus a trace of S<sup>2−</sup>. This was calculated from the pK<sub>a1</sub> of 7.05 for the reaction H<sub>2</sub>S ↔ H<sup>+</sup> + HS<sup>−</sup> value at 25°C in pure water (140). At a mammalian body temperature of 37°C, the pK<sub>a1</sub> for H<sub>2</sub>S ↔ H<sup>+</sup> + HS<sup>−</sup> is 6.76 (57) in water and 6.6 in 140 mM NaCl (277). For pK<sub>a1</sub> = 6.6, the Henderson–Hasselbach equation predicts that if H<sub>2</sub>S gas, or HS<sup>−</sup> (e.g., NaHS), or S<sup>2−</sup> (e.g., Na<sub>2</sub>S) is dissolved in an aqueous 140 mM NaCl solution at 37°C and pH 7.4, then 14% of the free sulfide will be H<sub>2</sub>S gas and 86% will be HS<sup>−</sup>, plus there will be a trace of S<sup>2−</sup>. There will be only a trace of S<sup>2−</sup>, because pK<sub>a2</sub> is greater than 12 (57, 76, 97). These dissociation reactions for the weak acid H<sub>2</sub>S are shown in Figure 1, along with the sources and disposal routes for sulfides. Since all three species of sulfide are always present in aqueous solutions, it has not been possible to determine which of these species is biologically active. Thus, the terminology of “the H<sub>2</sub>S concentration” usually refers to the sum of H<sub>2</sub>S, HS<sup>−</sup>, and S<sup>2−</sup>, although “sulfide concentration” is more accurate. In this review, we follow the common convention of calling the sum of all free sulfide species “H<sub>2</sub>S concentration.”

One important property of the H<sub>2</sub>S gas is that it is lipophilic: It easily partitions into the hydrophobic core of the cell membrane similar to O<sub>2</sub> and CO<sub>2</sub>, and, thus, rapidly diffuses into or out of cells (161). Since the H<sub>2</sub>S gas is also very volatile, it rapidly diffuses out of the blood into the lungs (170), or out of organ baths or cell culture media into the air. For example, when a 2 mm deep pool of culture medium containing 100 μM NaHS (i.e., ca. 14 μM H<sub>2</sub>S gas and 86 μM HS<sup>−</sup>) was exposed to

the air, the  $\text{H}_2\text{S}$  concentration ( $\text{H}_2\text{S} + \text{HS}^-$ ) decayed exponentially with a half time of about 6 min as the  $\text{H}_2\text{S}$  gas escaped into the air (69). This is an important point to note, especially for *in vitro* experiments. As  $\text{H}_2\text{S}$  escaped, the  $\text{H}^+$  in the buffered medium quickly combined with  $\text{HS}^-$  to keep the  $\text{H}_2\text{S}$  concentration at 14% in accordance with the  $\text{pK}_a$  for  $\text{H}_2\text{S} \leftrightarrow \text{HS}^-$  of 6.6 in 140 mM NaCl at 37°C (277).

The human nose is a very sensitive  $\text{H}_2\text{S}$  gas detector. When sufficient NaHS or  $\text{Na}_2\text{S}$  is added to buffered physiological salt solution to make 10  $\mu\text{M}$  sulfides, our nose detects a very strong  $\text{H}_2\text{S}$  odor. However, fresh blood and tissues are odorless despite being reported as having  $\text{H}_2\text{S}$  concentrations above 35  $\mu\text{M}$  (79, 208, 264, 309). Current evidence has shown that endogenously generated  $\text{H}_2\text{S}$  is rapidly oxidized to sulfate, or can be stored in proteins (Figs. 1 and 2) where it may be released on a physiological stimulus (98). Free  $\text{H}_2\text{S}$  concentration in blood and tissues is generally only 14 or 15 nM, as shown by gas chromatography (GC) (66) or the polarographic sensor (56, 277). The great discrepancies in reported  $\text{H}_2\text{S}$  concentration can be explained by the various  $\text{H}_2\text{S}$  detection methods employed, some of which release bound and stored sulfide (Fig. 3) (43, 56, 107, 130, 169, 247).

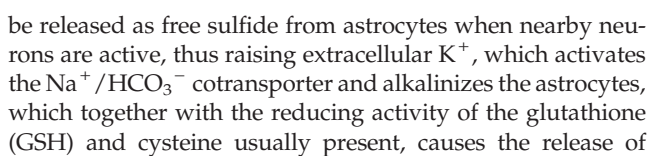
Earlier publications that reported  $\text{H}_2\text{S}$  concentrations above 35  $\mu\text{M}$  in fresh blood or plasma (46, 309, 315) have generally employed a strong acid or a strong base in their  $\text{H}_2\text{S}$  determination methods, both of which can release a sulfide that is bound to proteins (277). For example, the utilization of a strong acid in the methylene blue method (converts  $\text{HS}^-$  to  $\text{H}_2\text{S}$ ) releases sulfides from acid-labile sulfur (98, 277). On the other hand, the strong base (converts  $\text{H}_2\text{S}$  and  $\text{HS}^-$  to  $\text{S}^{2-}$ ) contained in the antioxidant buffer, as a part of the sulfide-sensitive electrode detection method, releases some of the protein-bound sulfide and can cause protein desulfuration (releasing sulfide from the constituent cysteine and methionine) (111, 277). This discrepancy between the methylene blue and sulfide electrode results, and the polarographic sensor and GC results has been reviewed by Olson (180). In this review, we include a discussion of recently reported results with monobromobimane (MBB) and with several fluorescent probes.

The exclusion of a strong acid or a strong base in  $\text{H}_2\text{S}$  measurements has led to the detection of a significantly lowered range of free sulfides. In 1992, Togawa *et al.* reported 1.3  $\mu\text{M}$  bound sulfide in human plasma utilizing the MBB

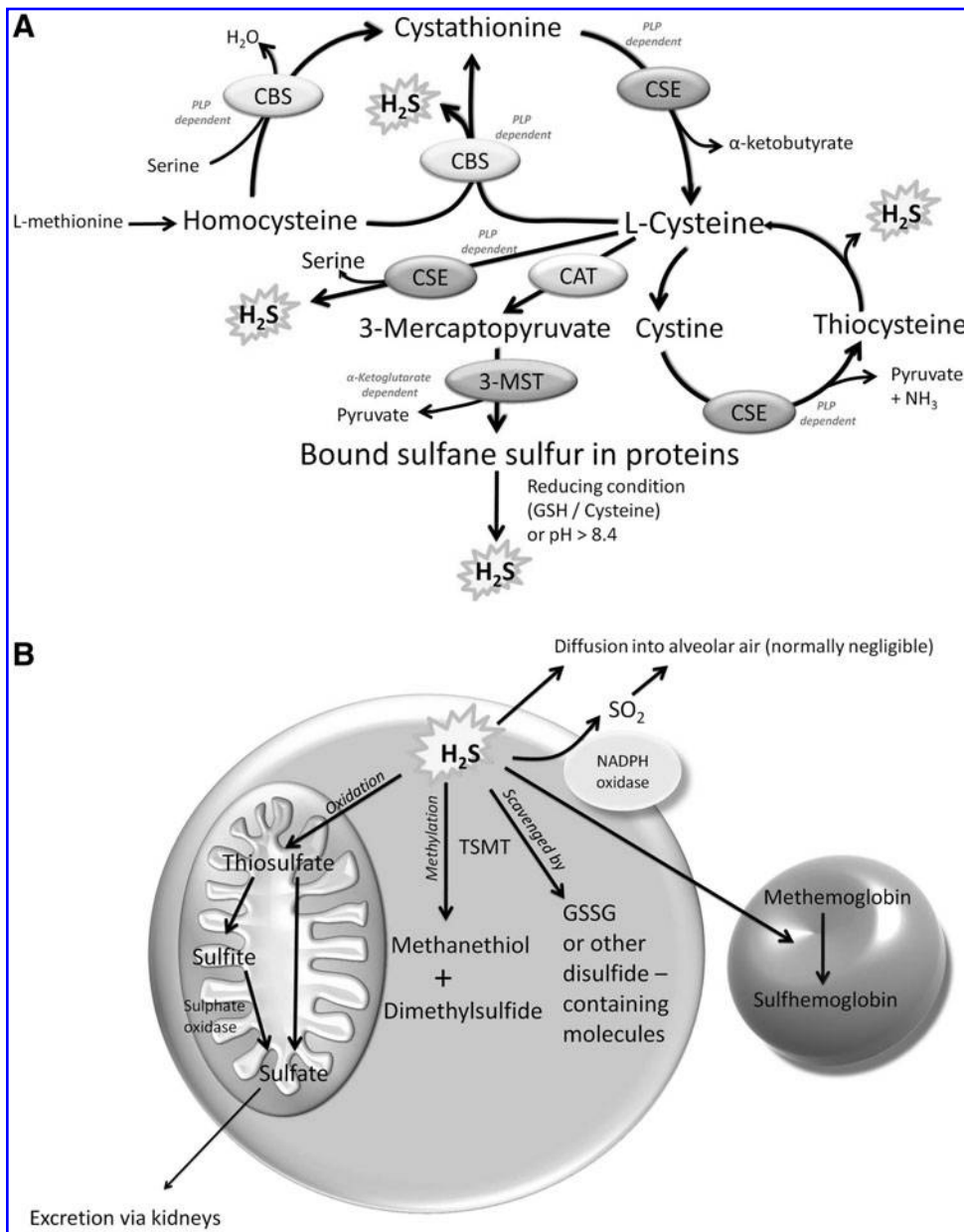
measurement with dithiothreitol (DTT) (pH 8) (247). It was assumed that the DTT together with pH 8 would release bound sulfide. In a mildly basic medium, MBB reacts with  $\text{HS}^-$  to form sulfide dibimane, which can be separated from MBB by HPLC and detected by its fluorescence properties. Recently, the MBB method was employed without using DTT, to measure "available  $\text{H}_2\text{S}$ " in rat blood (278) and in mouse plasma (218). Wintner *et al.* used pH 8 and found 0.7  $\mu\text{M}$   $\text{H}_2\text{S}$  in rat blood (278), while Shen *et al.* used pH 9.5 and found 1.7  $\mu\text{M}$   $\text{H}_2\text{S}$  in mouse plasma (218). Possibly, the pH 9.5 used by Shen *et al.* was alkaline enough to release some bound sulfide. We hypothesize that as MBB removes free  $\text{HS}^-$ , more  $\text{HS}^-$  that is ionically associated with cationic sites on proteins will be released into the free solution, especially in mildly basic solutions that will neutralize some of the protein- $\text{NH}_3^+$  groups, allowing MBB to sequester the  $\text{HS}^-$  released. Thus, the MBB method finds higher  $\text{H}_2\text{S}$  concentrations than the polarographic sensor method (277), which measures free  $\text{H}_2\text{S}$  gas concentration without sequestering it. In addition to the MBB method, Wintner *et al.* also used a polarographic sensor (at physiological pH) and found free  $\text{H}_2\text{S}$  concentrations in the low nanomolar range in rat blood (278), in agreement with earlier measurements by Whitfield *et al.* using the polarographic sensor (277). Levitt *et al.* recently determined that the  $\text{H}_2\text{S}$  concentration in mouse blood is  $\sim 15$  nM, using a new sensitive GC method (130). Olson has reviewed the evidence that  $\text{H}_2\text{S}$  may not be a circulating gasotransmitter, due to the extremely low concentration of free  $\text{H}_2\text{S}$  in blood (180), contrary to previous reports.

Although the concentration of free  $\text{H}_2\text{S}$  in body fluids may be low, in intracellular locations where  $\text{H}_2\text{S}$  synthesizing enzymes are highly concentrated,  $\text{H}_2\text{S}$  might temporarily become highly concentrated in the micro environment before it has time to diffuse away, be bound, or be oxidized. For example, Levitt *et al.* have shown that free  $\text{H}_2\text{S}$  concentration in freshly homogenized mouse aorta is 20 to 200 times more concentrated than in various other tissues they measured with the same method (130), probably due to the higher concentration of cystathionine  $\gamma$ -lyase (CSE) in arteries. Moreover, under the right physiological conditions or on physiological stimuli, free  $\text{H}_2\text{S}$  may be released from sulfur stores to raise the free  $\text{H}_2\text{S}$  concentration in a micro environment (98). In rat brain, for example, it was demonstrated that bound sulfur can

**FIG. 1. Handling of  $\text{H}_2\text{S}$  in the mammalian biological system.** The main source of  $\text{H}_2\text{S}$  in the body starts with the digestion of proteins into amino acids. The cysteine and methionine may be metabolized to  $\text{H}_2\text{S}$  via the action of endogenous enzymes found in many of the body's cells. Bacteria in the lower gut also contribute another source of  $\text{H}_2\text{S}$ . The inhalation of  $\text{H}_2\text{S}$  from air may be an exogenous source of  $\text{H}_2\text{S}$ , but this is minimal under normal conditions. Due to the lipophilic, hydrophilic, and volatile properties of  $\text{H}_2\text{S}$ , it penetrates the cell membranes freely and can easily pass to and from the air, blood, and tissues. In an aqueous environment,  $\text{H}_2\text{S}$  is a weak acid that exists in three free states,  $\text{H}_2\text{S}$  gas,  $\text{HS}^-$ , and a trace of  $\text{S}^{2-}$ . The tissue and circulation level of free  $\text{H}_2\text{S}$  is in the nanomolar range.  $\text{H}_2\text{S}$  is ultimately oxidized to sulfate and excreted by the kidneys in order to maintain the sulfur balance: output (red arrows) should equal input (green arrows). Some  $\text{H}_2\text{S}$  is stored as sulfides by the formation of protein persulfides, or it becomes otherwise bound to proteins and exists as bound sulfane sulfur or acid-labile sulfur. Due to the negative charge on  $\text{HS}^-$ , it is attracted to positively charged amino groups on proteins; we postulate that this pulls some  $\text{HS}^-$  out of the pool of free  $\text{H}_2\text{S}$ , contributing toward the low concentration of free  $\text{H}_2\text{S}$  detected in blood and tissues. While  $\text{H}_2\text{S}$  may be lost from the body via flatus or exhalation, the large majority of  $\text{H}_2\text{S}$  is eliminated from the body via oxidation and excretion as sulfate in the urine. The main  $\text{H}_2\text{S}$  synthesizing enzymes are as follows: cystathionine  $\beta$ -synthase (CBS), cystathionine  $\gamma$ -lyase (CSE), mercaptopyruvate sulfurtransferase (3-MST), and cysteine aminotransferase (CAT) in specific organs are indicated in blue. Some organs express mainly one type of enzyme (e.g., heart), whereas others might express multiple types (e.g., liver). (To see this illustration in color, the reader is referred to the web version of this article at [www.liebertonline.com/ars](http://www.liebertonline.com/ars)).  $\text{H}_2\text{S}$ , hydrogen sulfide.



bound H<sub>2</sub>S (98). The brain has been reported as containing 61  $\mu$ M "bound sulfur" (265). The H<sub>2</sub>S released from stored sulfide in the brain as just described can act as a modulator of synaptic activity (1). Possible mechanisms similar to those described in the brain by Ishigami *et al.* (98) may occur in other



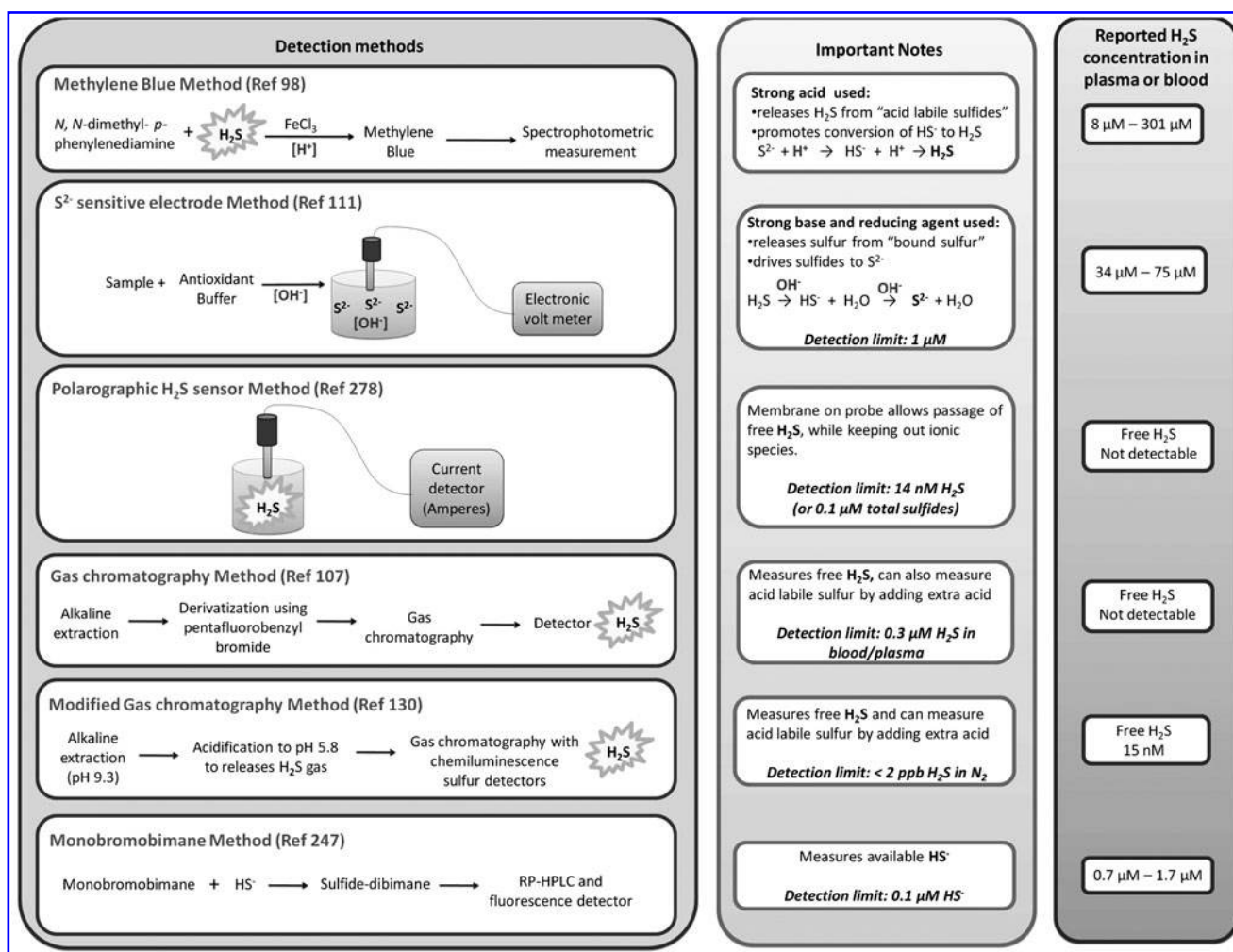
**FIG. 2. Biosynthesis and catabolism of H<sub>2</sub>S in mammals.** A outlines the biosynthesis of H<sub>2</sub>S in mammalian cells. There are four endogenous H<sub>2</sub>S synthesizing enzymes: CBS, CSE, 3-MST, and CAT. The main H<sub>2</sub>S precursor is L-cysteine. The other sulfur-containing amino acid, L-methionine, is a precursor of L-homocysteine, which can be metabolized to cysteines. CSE is expressed in the liver, heart, blood vessels, and other organs. CBS is expressed in the liver, brain, and other organs. CAT and 3-MST have been reported in the brain and aorta. B outlines the breakdown of H<sub>2</sub>S at the cellular level. H<sub>2</sub>S is mainly oxidized in mitochondria to thiosulfate, which is further converted to sulfite and, finally, sulfate. Sulfite can be converted to sulfate by sulfate oxidase. H<sub>2</sub>S may also undergo methylation catalyzed by cytosolic S-methyltransferase (TSMT). H<sub>2</sub>S may be scavenged by either methemoglobin or metallo-/disulfide-containing molecules. Potentially, H<sub>2</sub>S can be exhaled as free H<sub>2</sub>S gas or SO<sub>2</sub> gas after being metabolized by NADPH, but usually, this loss is negligible. NADPH, nicotinamide adenine dinucleotide phosphate.

organs or tissues. Physiological mechanisms, as yet poorly understood, may add to or remove the sulfide carried in plasma proteins. This may explain why the methylene blue and sulfide-sensitive electrode methods have shown that H<sub>2</sub>S in plasma increases or decreases in some human diseases or animal disease models, and that the inhibitors of H<sub>2</sub>S synthesizing enzymes in animal models cause the measured plasma H<sub>2</sub>S (*i.e.*, stored sulfide) to decrease, while also changing physiological parameters such as blood pressure (BP) in parallel.

We postulate that experiments demonstrating the physiological effects of higher concentrations of H<sub>2</sub>S than those that occur in mammalian macro environments may be uncovering the effects of H<sub>2</sub>S concentrations that occur physiologically in micro environments near reservoirs of sulfide bound to proteins or near concentrations of CSE. The development of microelectrodes that are specific for detecting H<sub>2</sub>S or HS<sup>-</sup> may

some day find such H<sub>2</sub>S "hot spots." Another promising method that may be able to find H<sub>2</sub>S "hot spots" in living cells is the use of intracellular fluorescent probes which are selective for H<sub>2</sub>S. Just recently, at least four new fluorescent probes that are selective for H<sub>2</sub>S have been described by different research groups (143, 144, 192, 198, 200).

The ability of H<sub>2</sub>S to act as a reducing agent is often employed as the chemical basis of the probe. Lippert *et al.* utilized the reducing property of H<sub>2</sub>S in the conversion of azides to fluorescent rhodamines to develop two fluorescent probes—Sulfidefluor-1 (SF1) and -2 (SF2) (143). Both probes have similar absorption and emission wavelengths (maximum absorption: ~490 nm; emission: 525 nm). Under *in vitro* conditions, the detection limits were shown to be 5–10 μM. At 100 μM NaHS, these probes showed ~50-fold greater selectivity than thiols and ~3-fold more selectivity than a variety of reactive sulfur, oxygen, and nitrogen species.



**FIG. 3. Methods used to detect H<sub>2</sub>S in biological samples.** In the last 10 years, the two most commonly used methods for measuring the H<sub>2</sub>S concentration have been the methylene blue and the sulfide-sensitive electrode. The reported concentration of H<sub>2</sub>S in the blood or plasma of several species, including rat, human, or mouse, have ranged from 8 to 301 μM using these two methods. These relatively high concentrations would make fresh blood smell similar to rotten eggs if they represented free H<sub>2</sub>S. The polarographic H<sub>2</sub>S sensor and the gas chromatography can accurately measure free H<sub>2</sub>S concentrations down to as low as 14 nM, without releasing stored sulfide. These methods find that the free H<sub>2</sub>S concentration is either not detectable or within the nanomolar range of ~15 nM. The sixth method measures the available HS<sup>-</sup>, which may include ionically bound HS<sup>-</sup> in addition to the free HS<sup>-</sup>. This method finds about 1 μM HS<sup>-</sup>.

Peng *et al.* also utilized H<sub>2</sub>S-mediated reduction to synthesize dansyl azide (DNS-Az), a sulfonamide converted from sulfonyl azide (192). Under *in vitro* conditions, it was reported to have a detection limit of 1 μM sulfide and a signal-to-noise ratio of 3:1. The effects of 17 other anions with some reducing ability were investigated, and the fluorescence intensities were far less than those induced by HS<sup>-</sup>. Interestingly, using this fluorescent chemoprobe, the concentration of H<sub>2</sub>S in the blood of C57BL6/J mice was found to be ~32 μM, a value much higher than the recently reported nanomolar range of free sulfides (130, 277, 278), but similar to the concentrations found with the methylene blue method. A possible explanation of this might be that the high level of GSH (>4 mM) present in the blood plasma could have interfered with the DNS-Az probe assay, as Peng *et al.* did not test the sensitivity of their probe for GSH or cysteine, two potent reducing agents present in the blood (192).

All three probes (SF-1, SF-2, and DNS-Ax) reflect the H<sub>2</sub>S concentration by indirectly measuring redox status. High fluorescence intensity depicts a large extent of reduction, suggesting a high concentration of H<sub>2</sub>S. It should be noted, therefore, that chemicals which strongly affect redox status should not be used along with these probes, as they will severely interfere with the assay.

The fluorescent probe (compound 1) created by Liu *et al.* made use of the ability of H<sub>2</sub>S to undergo nucleophilic reactions twice, whereas most other biological nucleophiles and thiols undergo nucleophilic reactions only once (144). H<sub>2</sub>S, but not cysteine or GSH, leads to increments in fluorescence *in vitro* with maximum intensity reached in 1 h. When this probe was added to bovine plasma, very weak fluorescence was observed, in contrast to the strong fluorescence seen with Peng *et al.*'s DNS-Az. Most importantly, when researchers applied exogenous NaHS (10–600 μM) to bovine plasma,

intensities of the fluorescent probe were weaker than with equimolar concentration of  $\text{H}_2\text{S}$  in buffer solutions, in line with the postulation that  $\text{H}_2\text{S}$  rapidly binds to proteins or is oxidized in plasma (144, 277).

Qian *et al.* synthesized two sulfide-selective fluorescent probes (SFP-1 and SFP-2) that not only detect sulfides under *in vitro* conditions, but also monitor cell-based sulfides using live-cell imaging with minimal toxic effects on cell viability (198). SFP-1 emits blue fluorescence with an emission maximum at 391 nm (excitation: 300 nm), whereas SFP-2 emits green fluorescence with an emission maximum at 510 nm (excitation: 465 nm). Under *in vitro* conditions, SFP-1 produced a fluorescence signal in a concentration-dependent manner, starting from 10  $\mu\text{M}$   $\text{Na}_2\text{S}$  and reaching saturation at 50  $\mu\text{M}$ . SFP-2 responded to  $\text{Na}_2\text{S}$  (5–100  $\mu\text{M}$ ) in a concentration-dependent manner, with a greater selectivity toward sulfides over thiols than SFP-1 (>150-fold *vs.* 50–100-fold). However, under *in vivo* conditions, the specificity was not as good. When tested in HeLa cells, for example, SFP-2 produced a similar extent of fluorescence using equimolar concentrations (100  $\mu\text{M}$ ) of cysteine, GSH, or  $\text{Na}_2\text{S}$  (198).

Other than DNS-Az, all other fluorescence probes were also tested in cultured cells for their potential use to monitor cell-based sulfides. Generally, no signs of toxicity on cell viability were observed when these probes were utilized. It is worthy to note that all these probes showed little or no fluorescence at basal levels and only emit strong fluorescence when high micromolar concentrations of  $\text{H}_2\text{S}$  donors (250  $\mu\text{M}$  NaHS or  $\text{Na}_2\text{S}$ ) are added to the cells (143, 144). In fact, SFP-1 and SFP-2 showed a reduced sensitivity for sulfide detection *in vivo* as compared with *in vitro* experiments. As mentioned earlier, *in vivo* GSH and cysteine may cause major false-positive results. This may limit their use to monitor endogenous  $\text{H}_2\text{S}$  production that is thought to be within the nanomolar range, far below the detection limits of these probes. Furthermore, the high  $\text{H}_2\text{S}$  concentrations applied in these experiments are probably beyond the physiological, and the micromolar detection limit might pose a major limitation of these probes in studying endogenous free sulfides.

It should also be noted that blood emits autofluorescence at 400 nm (160), whereas other endogenous fluorophores such as NaDH, FAD, and tryptophan also commonly display excitation/emission wavelengths in the range of 300–500 nm (312). As such, fluorescence probes with excitation/emission wavelengths falling within this range may face problems during  $\text{H}_2\text{S}$  assay dealing with blood samples. In view of this, Quek *et al.* synthesized a mixed-valence diruthenium complex that they abbreviated as  $[\text{Ru}_2]^+$ , a dye which can detect  $\text{H}_2\text{S}$  with an absorbance in the near infrared range (200).  $[\text{Ru}_2]^+$  absorbs at 789 nm, but  $\text{H}_2\text{S}$  reduces it to  $[\text{Ru}_2]$ , which absorbs at 895 nm. The change over from 789 to 895 nm absorbance correlates linearly with NaHS concentration, with a detection limit of 1.35 to 16  $\mu\text{M}$ . Though this method is highly specific for detecting  $\text{HS}^-$  as compared with many other anions and reducing agents, cysteine and ascorbic acid cause some reduction of  $[\text{Ru}_2]^+$  (200). For now, it is useful in measuring the rate of release of  $\text{H}_2\text{S}$  by slow release donors under *in vitro* conditions.

The most common  $\text{H}_2\text{S}$  donor utilized by most publications is sodium hydrosulfite (NaHS), because it is readily available, inexpensive, and easy to work with. However, the purity of commercially available NaHS hardly exceeds 70%. The 30%

or more uncontrolled impurities may underlie some of the inconsistencies observed among different laboratories. Other  $\text{H}_2\text{S}$  donors such as sodium sulfide ( $\text{Na}_2\text{S}$ ) or its liquid solution form, IK-1001 (Ikaria Holdings, Inc.), should be sought as purer alternatives, while slow  $\text{H}_2\text{S}$ -releasing compounds (*e.g.*, GYY4137) (128, 137, 138, 300) and  $\text{H}_2\text{S}$ -donating hybrids (*e.g.*, ACS14, ACS15, ACS21, ACS 84, S-diclofenac) (151, 206, 304) are under intensive research as promising therapeutics for disease conditions.

Given the factors just mentioned, organ bath or cell culture experiments utilizing <100  $\mu\text{M}$   $\text{H}_2\text{S}$  could be of physiological relevance, in agreement with Furne *et al.*'s proposal (66). The development of new methods that measure  $\text{H}_2\text{S}$  in microenvironments is warranted for a better understanding of the intracellular  $\text{H}_2\text{S}$  levels and actions. According to Henry's Law, breathing in a lethal amount of 500 ppm  $\text{H}_2\text{S}$  gas will produce a blood concentration of 227  $\mu\text{M}$   $\text{H}_2\text{S}$  (277). With our present knowledge, we hypothesize that physiological  $\text{H}_2\text{S}$  concentrations in microenvironments and blood circulation do not exceed 100  $\mu\text{M}$ . It is, therefore, noteworthy that *in vitro* experiments involving  $\text{H}_2\text{S}$  concentrations above 100  $\mu\text{M}$   $\text{H}_2\text{S}$  are not likely to depict physiological effects.

Another important factor affecting the final  $\text{H}_2\text{S}$  concentration in circulation and tissues of interest for *in vivo* experiments is the route of drug administration. While intravenous (*i.v.*) administration of  $\text{H}_2\text{S}$  results in greater bioavailability of  $\text{H}_2\text{S}$ , intraperitoneal (*i.p.*) or subcutaneous administration leads to relatively slower absorption and lower bioavailability. In some studies, researchers administered  $\text{H}_2\text{S}$  *via* oral gavage. This  $\text{H}_2\text{S}$  has to travel from the gastrointestinal tract to the liver, where it is metabolized before it reaches possible target organs. The eventual  $\text{H}_2\text{S}$  concentration in systemic circulation and target organs will be drastically reduced. This may justify the administration of higher than physiological concentrations of  $\text{H}_2\text{S}$  in some *in vivo* experiments. Nevertheless, caution should still be taken when we interpret data generated using high micromolar ranges of  $\text{H}_2\text{S}$  concentration for their physiological relevance.

### B. $\text{H}_2\text{S}$ biosynthesis

The concentration of free sulfides ( $\text{H}_2\text{S}$ ,  $\text{HS}^-$ ) in mammalian blood and most tissues is very low (<100 nM) (277), although it is reported to be higher (1  $\mu\text{M}$ ) in aorta (130). Sulfides are also bound to proteins in blood and tissues; for example, the concentration of acid-labile sulfur in the heart was reported to be about 300  $\mu\text{M}$  (130). Free and bound sulfide originates from the action of enzymes that synthesize  $\text{H}_2\text{S}$ . The four most important mammalian enzymes that synthesize  $\text{H}_2\text{S}$  are cystathionine  $\beta$ -synthase (CBS, EC 4.2.1.22), CSE (cystathionase, CSE, EC 4.4.1.1), and cysteine aminotransferase (CAT, EC 2.6.1.3) in conjunction with mercaptopyruvate sulfurtransferase (3-MST, EC 2.8.1.2). The biochemistry of these four enzymes is summarized in Figure 2A, and the details may be found in the reviews by Kabil and Banerjee (106) and Singh and Banerjee (225).

The expression of CBS and CSE has been detected in a broad variety of cell types, including those from the liver, kidney, heart, vasculature, brain, skin fibroblasts, and lymphocytes. In some tissues, both CBS and CSE contribute to the local generation of  $\text{H}_2\text{S}$  (such as in liver and kidneys) (282),

whereas in others, one enzyme predominates. For example, CSE is the main H<sub>2</sub>S-generating enzyme in the cardiovascular system (17, 309). CSE mRNA expression was detected in the myocardium with 24.4% higher expression than that in the thoracic aorta (75). CSE is present in arteries in relatively large amounts and is expressed in the endothelial cells (ECs) (288) and smooth muscle cells (SMCs) (309). *CSE*<sup>-/-</sup> mice were reported to develop hypertension spontaneously (288), whereas a later study failed to reproduce this finding (99). Nevertheless, the significance of CSE in the cardiovascular system should not be disregarded, as *CSE*<sup>-/-</sup> mice developed lethal myopathy and were susceptible to oxidative injury with a diet deficient in cysteine (99).

It was conventionally regarded that CBS is the predominant H<sub>2</sub>S synthase in the brain and nervous system (1). However, Shibuya *et al.* later discovered that brain homogenates of CBS<sup>-/-</sup> mice produce H<sub>2</sub>S at levels similar to those of wild-type mice (221). They also showed that 3-MST is expressed in neurons in the brain. Along with CAT, 3-MST produces H<sub>2</sub>S using both L-cysteine and  $\alpha$ -ketoglutarate as substrates. Their experiments suggest that 3-MST and CAT contribute to H<sub>2</sub>S formation in both the brain (201) and the vascular endothelium (18, 220, 221). A recent paper has shown that CAT and 3-MST can produce H<sub>2</sub>S only in alkaline conditions and in the presence of DTT, a strong reducing agent (225). Therefore, the physiological relevance of 3-MST as a source of H<sub>2</sub>S formation in the brain remains to be elucidated in the future.

Stearcy and Lee demonstrated the reduction of exogenous S<sub>8</sub> in producing H<sub>2</sub>S by human erythrocytes using reducing equivalents from glucose oxidation (214). In addition, they also found a slower production of H<sub>2</sub>S without adding S<sub>8</sub>, suggesting an endogenous source of sulfur in red blood cells (214). Thus, inorganic synthesis of H<sub>2</sub>S may contribute toward endogenous H<sub>2</sub>S formation *in vivo*, though its implication is yet to be discovered.

### C. H<sub>2</sub>S catabolism

Figure 2B summarizes the cellular mechanisms that dispose of H<sub>2</sub>S. The vast majority of H<sub>2</sub>S is oxidized to sulfate, which leaves the body *via* the kidneys (52, 66, 67, 158). The primary site for this oxidation is in the liver, but all cells in the body can oxidize H<sub>2</sub>S (66, 67, 277). The half time for disappearance of 30  $\mu$ M free sulfides is 51 s for cow blood, and 191 s for 5% bovine serum albumin (277); hence, even plasma can oxidize H<sub>2</sub>S, while blood does it more rapidly. It has been suggested that a major portion of the ability of plasma or blood to rapidly consume sulfide added *in vitro* is due to the binding of the sulfide to proteins (52).

Usually, most or all of the H<sub>2</sub>S produced by CSE and CBS in the liver, from the cysteine and methionine derived from digested proteins, is oxidized (or bound) before it can escape into the hepatic veins that carry the blood to the inferior vena cava (66). Mitochondria are very effective in oxidizing sulfides; however, if the supply of sulfide exceeds the ability of the mitochondrial enzymes to oxidize all of it, then the H<sub>2</sub>S can poison the mitochondrial enzymes (80). No H<sub>2</sub>S is released when the homogenates of liver are incubated with the physiological concentrations of 1 or 0.1 mM cysteine. Anaerobically, the same experiments show a huge release of H<sub>2</sub>S with 10 mM cysteine, but still none with 1 or 0.1 mM cysteine (66). The results suggest that oxidation, especially in the liver,

is an important way by which the body rids itself of endogenously produced H<sub>2</sub>S. Even without oxygen, however, the liver can absorb all the H<sub>2</sub>S produced with physiological levels of cysteine, probably due to the absorption by proteins and to methylation.

Endogenous H<sub>2</sub>S may be metabolized *in vivo* by different routes (Fig. 2B). As a readily diffusible gas, it can be metabolized in mitochondria by oxidation to thiosulfate, which is further converted to sulfite and sulfate by sulfate oxidase (67). Finally, the end products, sulfates, are excreted in the urine as either free or conjugated sulfate (12, 52). Another metabolic pathway involves the methylation of sulfides by cytosolic S-methyltransferase to methanethiol and dimethylsulfide (67). H<sub>2</sub>S can also be scavenged by methemoglobin (12) or metallo- or disulfide-containing molecules such as oxidized GSH (228). Hemoglobin may act as a common sink for vasoactive gases (CO, NO, and H<sub>2</sub>S), and these three gases compete with oxygen for binding, thus contributing to their toxicity on high exposure.

Mammalian lungs may occasionally provide an escape route for H<sub>2</sub>S (Fig. 1), possibly during septic shock, hemorrhagic shock, or pancreatitis when larger than normal amounts of H<sub>2</sub>S may be generated. In healthy individuals, however, very little H<sub>2</sub>S is lost *via* the lungs, because metabolic disposal keeps the free level of H<sub>2</sub>S in blood very low (66). Alveolar air (end expiration) usually contains only 25–50 ppb H<sub>2</sub>S (172, 237) in healthy subjects (not enough to smell); thus, the normal daily loss of H<sub>2</sub>S *via* the lungs is very small compared with the loss of sulfate in the urine.

### D. Subcellular compartmentalization of H<sub>2</sub>S

Subcellular compartmentalization of NO synthase (NOS) is an important regulatory mechanism for NO signaling (8). Subcellular compartmentalization of H<sub>2</sub>S is also suggested in eukaryotic cells, as a substantial amount of H<sub>2</sub>S-oxidizing activity was observed in mitochondria while sulfur-reducing activity was observed in the cytoplasm (213, 214). The localization of H<sub>2</sub>S synthesizing enzymes, sulfur stores, and metabolic routes provides insights to such an understanding. Both CBS and CSE are localized in the cytosol (5), and their activities are absent in mitochondria (5). 3-MST and CAT, on the other hand, are mainly localized in the mitochondria (123, 222). Since different cell types express different types of enzymes, the localization of H<sub>2</sub>S synthesis may account for the differential effects of H<sub>2</sub>S in different tissues. Interestingly, bound sulfane sulfur is localized primarily in the cytoplasm, whereas acid-labile sulfur occurs mainly in iron-sulfur enzymes found in mitochondria (179). Compartmentalization of sulfur stores may facilitate differential H<sub>2</sub>S release for specialized purposes. Furthermore, bound sulfane sulfur is released as H<sub>2</sub>S under reducing conditions or at pH >8.4 (249), while acid-labile sulfur is released at pH <5.4 (98), further contrasting the mechanism of release of H<sub>2</sub>S from these stores. The metabolism of H<sub>2</sub>S, as mentioned earlier, mainly occurs in the mitochondria. Subcellular compartmentalization of H<sub>2</sub>S may have important physiological implications that are worthy of further studies and in-depth research. For example, crosstalk between H<sub>2</sub>S and NO could be highly regulated by their compartmentalization, accounting for the differential effects observed in different systems.

### III. Effects of H<sub>2</sub>S on the Heart

#### A. H<sub>2</sub>S effects on heart function

In the isolated rat heart, perfusion with 100  $\mu$ M or less NaHS had no significant effect on heart rate (165, 314). However, 10–1000  $\mu$ M NaHS decreased the amplitudes of myocyte twitch and electrically induced calcium transients in isolated rat ventricular myocytes treated with isoproterenol (ISO) (298). In the isolated heart, perfusion with NaHS (1–1000  $\mu$ M) inhibited maximal/minimal left ventricular (LV) pressure development ( $\pm$ LVdp/dt<sub>max</sub>) (75). The administration of NaHS (2.8  $\mu$ mol/kg body weight) *in vivo* via the femoral vein produced a similar effect on cardiodynamics and a transient decrease in mean arterial pressure (MAP) in anesthetized rats (75). Since the high concentrations of NaHS used were well above the physiological H<sub>2</sub>S concentrations of <100  $\mu$ M, it is still unclear whether H<sub>2</sub>S plays a physiological role in heart function.

#### B. Heart electrophysiology

The effects of H<sub>2</sub>S on heart electrophysiology and intracellular signaling are illustrated in Figure 4.

1. Intracellular calcium regulation. During a cardiomyocyte action potential, depolarization of the sarcolemma due to an initial brief increase in Na<sup>+</sup> permeability, followed by a longer lasting opening of L-type Ca<sup>2+</sup> channels (LTCC), results in inward Ca<sup>2+</sup> currents. Sun *et al.* found that NaHS (25–400  $\mu$ M) reduced the peak current of LTCC (I<sub>Ca<sub>v</sub>L) in a concentration-dependent manner (239). It should be noted, however, that the effects observed at H<sub>2</sub>S concentrations above 100  $\mu$ M probably do not suggest physiological relevance. Whether or not LTCC is the primary action site of H<sub>2</sub>S still remains to be determined. The possibility that the effect of H<sub>2</sub>S on LTCC is secondary to other signaling pathways may not be fully excluded. For instance, the reduction of the Ca<sup>2+</sup> current through LTCC may also result from hyperpolarization caused by the opening of K<sub>ATP</sub> channels (102, 245) or the suppressed cAMP/protein kinase A [PKA] pathway</sub>

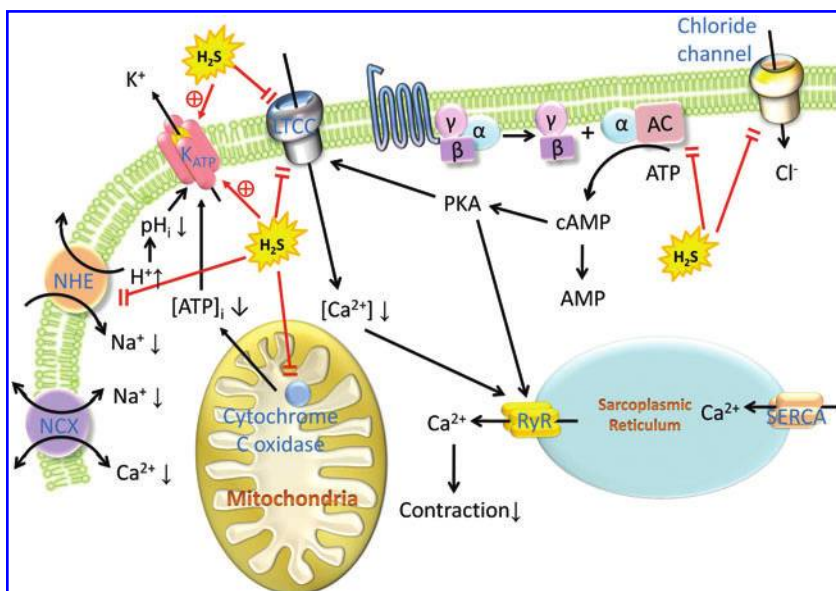
(298) (Fig. 4). More work, such as single-channel recording, is needed to determine conclusively whether or not H<sub>2</sub>S is a direct LTCC blocker using near-physiological H<sub>2</sub>S concentration.

2. K<sub>ATP</sub> channels. K<sub>ATP</sub> channels are widely distributed in the myocardium. The opening of K<sub>ATP</sub> channels generates outward currents, causing hyperpolarization. This reduces calcium influx *via* LTCC and prevents Ca<sup>2+</sup> overload. Thus, the K<sub>ATP</sub> channel opening is an important endogenous cardioprotective mechanism, and it is crucially involved in cardiac ischemia preconditioning.

Evidence that NaHS (100  $\mu$ M) opens the K<sub>ATP</sub> channels was obtained by Wang and coworkers (102, 245). This is discussed in detail in Mechanisms of H<sub>2</sub>S-induced vasorelaxation section. Furthermore, H<sub>2</sub>S may also indirectly activate the K<sub>ATP</sub> channels by inducing intracellular acidosis (14, 51, 119, 127). By activation of the K<sub>ATP</sub> channels, H<sub>2</sub>S shortens action potential duration (APD) (2) and produces cardioprotective effects (17, 104, 188, 226, 306). This is further discussed in Mechanisms for H<sub>2</sub>S-induced cardioprotection and Mechanisms for the cardioprotection of SPreC sections.

3. Chloride channels. Chloride channels play important roles in diverse processes such as BP regulation, cell cycle and apoptosis, muscle tone, volume regulation, synaptic transmission, and cellular excitability (164). Malekova *et al.* investigated the effect of H<sub>2</sub>S on single-channel currents of chloride channels using the patch clamp technique (159). It was found that 20 to 200  $\mu$ M NaHS inhibited the chloride channels by decreasing the channel open probability in a concentration-dependent manner. Therefore, the inhibitory effect of H<sub>2</sub>S on the chloride channels may be involved in the biological actions of H<sub>2</sub>S in the heart (159).

4. Action potential. H<sub>2</sub>S has no significant effect on the amplitude of action potential and resting potential (239). However, NaHS at concentrations of 100–500  $\mu$ M markedly reduced APD and decelerated the sinus rhythm. As just mentioned, the effect of H<sub>2</sub>S on APD is attributed to the



**FIG. 4. Effect of H<sub>2</sub>S on electrophysiology of the heart.** H<sub>2</sub>S inhibits heart contractions mainly by the suppression of adenylyl cyclase (AC), activation of ATP-sensitive potassium (K<sub>ATP</sub>) channels, and blockade of L-type Ca<sup>2+</sup> (LTCC) channels. H<sub>2</sub>S may also inhibit the activity of chloride (Cl<sup>-</sup>) channels, cytochrome *c* oxidase and Na<sup>+</sup>/H<sup>+</sup> exchanger (NHE), and the resultant decrease in intracellular pH and ATP, further implicating the activity of the K<sub>ATP</sub> channels (To see this illustration in color, the reader is referred to the web version of this article at [www.liebertonline.com/ars](http://www.liebertonline.com/ars)).

opening of the K<sub>ATP</sub> channels (2). However, the significance of this finding is yet to be determined, as the concentration used is likely to be more than physiological.

### C. Suppression of the AC/cAMP pathway

Yong *et al.* examined the action site of H<sub>2</sub>S in the  $\beta$ -adrenoceptor system with pharmacological manipulation (298). It was found that the amplitudes of electrically induced [Ca<sup>2+</sup>]<sub>i</sub> and myocyte twitch were significantly increased by the stimulation of AC with forskolin, activation of PKA with 8B-cAMP, or the opening of LTCC with Bay K-8644. NaHS (100  $\mu$ M) treatment significantly attenuated the effects of forskolin, but not those caused by 8B-cAMP and Bay K8644 (298). These data suggest that H<sub>2</sub>S may act on AC or its upstream signaling pathways, but not on PKA and LTCC. To confirm this, Yong *et al.* determined the effect of H<sub>2</sub>S on AC activity and found that NaHS (100  $\mu$ M) significantly decreased AC activity (298). These findings suggest that H<sub>2</sub>S has an inhibitory effect on AC activity, which may account for the decreased cAMP production.

### D. Interaction among gasotransmitters in the heart

Accumulating evidence suggests that interactions of biologically active gases (*i.e.*, H<sub>2</sub>S, NO, and CO) may influence the biological effects of one another (65, 108, 110, 133, 181). Interactions between H<sub>2</sub>S and NO have long been speculated, as several reports have demonstrated that H<sub>2</sub>S and NO may influence each other's production (74, 121, 148, 283, 285, 307). Moreover, others have found evidence that interactions between H<sub>2</sub>S and NO may alter the physiological response to either of these molecules alone (3, 274). Recently, Yong *et al.* found that a mixture of NO and H<sub>2</sub>S (100  $\mu$ M) produces an opposing effect in the heart as compared with either gas alone. This effect could be abolished by thiols, suggesting that the two molecules interact to form a third molecule that is thiol sensitive. Yong *et al.* proposed that

nitroxyl anion (HNO) could be one of the possible candidates (296), based on the important reducing capability of H<sub>2</sub>S (240, 257, 267) and the structural and pharmacological properties of HNO (296). Please refer to Figure 5 for greater details.

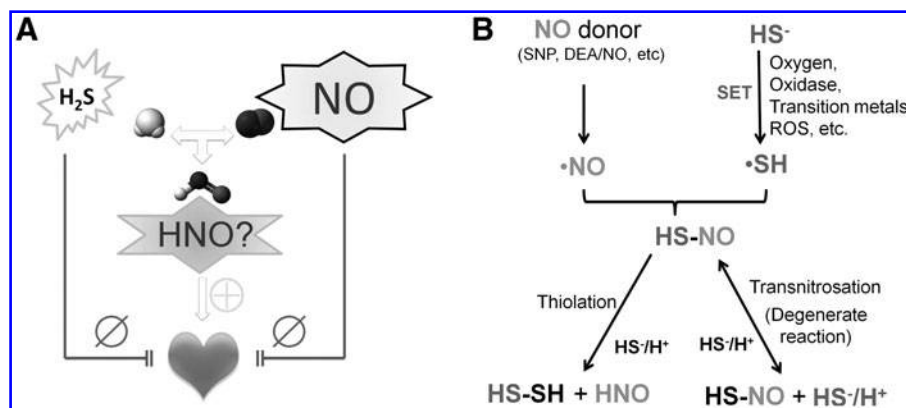
Experiments carried out in the liver suggest that CBS may act as an *in vivo* CO sensor (108, 223). It has also been observed that CBS activity can be directly inhibited by NO and CO (197). Although CBS plays a minor role in H<sub>2</sub>S synthesis in the cardiovascular system, more work has to be done to unveil any possible physiological roles of CBS or of the interactions between H<sub>2</sub>S and CO in the cardiovascular system.

## IV. Potential Therapeutic Effects of H<sub>2</sub>S in Heart Diseases

Cardiovascular diseases (CVD) are collectively a leading cause of death worldwide. The most common manifestations of CVD are coronary artery disease (CAD), chronic heart failure (CHF), and hypertrophy. The protective effects of H<sub>2</sub>S in these pathological conditions are discussed next.

### A. Ischemic heart diseases: Is H<sub>2</sub>S a biomarker for ischemic heart diseases?

Both the endogenous H<sub>2</sub>S level in ventricular myocytes (17) and CSE activity in the isolated heart (297) were found to be suppressed under ischemic conditions. An *in vivo* study showed that an injection of ISO that produces "infarct-like" myocardial necrosis reduces H<sub>2</sub>S levels in the myocardium (73). In the ISO-treated heart model, CSE mRNA gene expression in the myocardium was reported by Geng *et al.* (73) to be upregulated, but by Zhu *et al.* (315), to be decreased. The discrepancy between the two groups may be secondary to the different models used and the different timings for tissue collection. Nevertheless, current evidence collectively indicates that endogenous H<sub>2</sub>S production in the ischemic heart is likely to be reduced.



**FIG. 5. Interaction of H<sub>2</sub>S and NO in the heart.** (A) H<sub>2</sub>S and NO, each of which produces a negative inotropic effect on the heart, produce a positive inotropic effect when added together. Hence, H<sub>2</sub>S may interact with NO to form a novel compound exhibiting distinctive characteristics. Under experimental conditions, one possible candidate for such interactions could be the nitroxyl anion (HNO). (B) Proposed mechanism of HNO formation as a result of H<sub>2</sub>S-NO interaction. In the presence of cellular oxidants (such as molecular oxygen), ROS (such as H<sub>2</sub>O<sub>2</sub>), and oxidases, HS<sup>-</sup> can be oxidized to give thiyl radical (HS<sup>•</sup>). The NO released from the NO donor will then quickly combine with the HS radical to give nitrosylthiol. In the presence of HS<sup>-</sup>, there are two possible reactions with similar activation energy according to a model system studied by computational chemistry. The transnitrosation reaction with another molecule of HS<sup>-</sup> will lead to HS exchange, and the reaction is just a degenerate reaction without the formation of a new product. The other possible reaction between HSNO and HS<sup>-</sup> is the thiolation reaction, which leads to the formation of hydrogen disulfide (H<sub>2</sub>S<sub>2</sub>) and HNO. NO, nitric oxide.

Geng *et al.* also reported that the plasma H<sub>2</sub>S level decreased by 66% (from 60 to 20  $\mu$ M) in an ISO-induced myocardial ischemia rat model (73). Consistent with this, a clinical observational study has shown that the plasma concentration of H<sub>2</sub>S in patients with coronary diseases is significantly lowered in comparison with that in normal control subjects (26 *vs.* 52  $\mu$ M), suggesting that the decreased plasma H<sub>2</sub>S levels may correlate with the severity of coronary diseases (103). These observations lead to the suggestion that the plasma H<sub>2</sub>S level has the potential to be a biomarker for ischemic heart diseases. However, a good diagnostic or prognostic marker should change well before any clinical symptoms or vary according to the severity of the specific disease. It has been reported that the plasma H<sub>2</sub>S concentration also decreases in association with other diseases, for example, hypertension (43), Alzheimer's disease (147), stable asthma (280), and pneumonia (46). As discussed earlier in the introduction, however, the methods of H<sub>2</sub>S detection used in these studies involved harsh conditions that release bound sulfide; hence, caution should be exercised when these data are interpreted.

1. **Cardioprotective effects of H<sub>2</sub>S against ischemia-reperfusion-induced injury.** The reduced endogenous H<sub>2</sub>S production in the ischemic heart implies that the ischemic heart injury might, at least in part, result from the impaired endogenous production of H<sub>2</sub>S. The cardioprotective effects of H<sub>2</sub>S were, therefore, studied by several groups of scientists. Their results are summarized in Table 1.

Myocardial infarction (MI) is the most common presentation of ischemic heart disease; hence, it is also the gold standard in the study of cardioprotection. Johansen *et al.* reported that the perfusion of NaHS at 0.1–1  $\mu$ M reduced MI size in the Langendorff-perfused heart after 30 min of left main coronary artery occlusion and 120 min of reperfusion (104). In *in vivo* studies, NaHS (0.1–14  $\mu$ mol/kg) treatment also reduced MI size in rats (187, 315), mice (61), and pigs (186, 230, 231). Conversely, the administration of propargylglycine (PAG), a CSE inhibitor, significantly increased the infarct size caused by myocardial ischemia (20, 226). Moreover, elevation of endogenously produced H<sub>2</sub>S by cardiac-specific overexpression of CSE (in  $\alpha$ -MHC-CGL-Tg mouse) significantly limited the infarct size caused by left coronary artery (LCA) occlusion for 45 min followed by reperfusion for 72 h (61). This confirms that endogenous H<sub>2</sub>S may play an important role in maintaining cell function in the ischemic heart. Detailed information is provided in Table 1.

In addition to the reduction of infarct size, NaHS (40–100  $\mu$ M) treatment also significantly decreased the severity and duration of ischemia/reperfusion (I/R)-induced arrhythmias (17, 306) (Fig. 6) and improved myocardial contractile function in the ISO-induced ischemic rat heart (73) and in the I/R-induced ischemic porcine heart (230). In the isolated perfused heart, H<sub>2</sub>S-releasing derivative compounds (ACS14 and 21) produced improvement in heart mechanics after I/R injury (206).

Yong *et al.* investigated the effect of rapid intermittent supply of H<sub>2</sub>S in the early phase of reperfusion, termed H<sub>2</sub>S postconditioning (SPostC), on heart contractile function (297). In the SPostC group, hearts received six cycles of 10 s reperfusion and 10 s NaHS (100  $\mu$ M) infusion after ischemia,

while NaHS was given for 2 min continuously after ischemia in the SPostC2 group. Both SPostC and SPostC2 treatments significantly improved the cardiodynamics, including LV end diastolic pressure, LV developed pressure, and  $\pm$ dP/dt during reperfusion after ischemia.

Olson and colleagues reported that oxygen is critical for the catabolism of H<sub>2</sub>S *via* mitochondrial oxidation. During hypoxia when intracellular oxygen is low, the catabolism of H<sub>2</sub>S was found to be greatly reduced (183), resulting in an increased concentration of H<sub>2</sub>S. This seems to contradict the results of most studies which have found that the H<sub>2</sub>S concentration (measured with methylene blue method) was reduced during ischemia/hypoxia in the ischemic heart (see Ischemic heart diseases section). As just discussed, the methylene blue method is not a good assay for the detection of free H<sub>2</sub>S. It is, therefore, not known whether endogenous free-form H<sub>2</sub>S is decreased or increased. For these reasons, whether there is an inverse relationship between oxygen and H<sub>2</sub>S in the heart cannot be concluded. Without a good method to measure endogenous free-form H<sub>2</sub>S intracellularly, it is, therefore, difficult to explain why the inhibition of endogenous H<sub>2</sub>S production with PAG resulted in increased infarct size (20, 226). The harmful effects of PAG may come from a further decrease in the H<sub>2</sub>S level due to suppression of CSE or an attenuated elevation of the H<sub>2</sub>S level if the level is increased due to inhibition of its catabolism in ischemia. Importantly, it should be noted with caution that PAG is a nonselective inhibitor of pyridoxal 5'-phosphate-dependent enzymes, and could have exerted its effects on other enzymes besides CSE (273), contributing toward its detrimental effects. Nonetheless, both ischemic preconditioning (IPreC) (17) and postconditioning (297) have been demonstrated as elevating endogenous H<sub>2</sub>S levels in the heart. This implies that brief episodes of ischemia may increase the endogenous H<sub>2</sub>S level irrespective of whether it is from the stimulation of CSE or the inhibition of H<sub>2</sub>S catabolism.

*a. Mechanisms for H<sub>2</sub>S-induced cardioprotection.* Ischemic injury is a complex process involving the action and interaction of many factors (Fig. 6). On the interruption of oxygen supply, mitochondrial oxidative phosphorylation rapidly stops, resulting in depletion of high-energy phosphate compounds such as ATP and creatine phosphate (89). Anaerobic glycolysis increases to compensate for decreased aerobic ATP production, resulting in the accumulation of H<sup>+</sup> and lactate (25). The resultant intracellular acidosis causes alterations in ion transport in the sarcolemma and organelle membranes (26, 246). Initially, there is increased K<sup>+</sup> efflux related to an increased osmotic load caused by the accumulation of metabolites and inorganic phosphate. With a substantial reduction in [ATP], Na<sup>+</sup>/K<sup>+</sup>-ATPase activity declines, resulting in a decrease in [K<sup>+</sup>]<sub>i</sub> and an increase in [Na<sup>+</sup>]<sub>i</sub>. Intracellular acidosis also activates the sarcolemmal Na<sup>+</sup>/H<sup>+</sup> exchanger (NHE) (109, 292), which facilitates H<sup>+</sup> extrusion in exchange for Na<sup>+</sup>. The accumulated Na<sup>+</sup>, in turn, activates the reverse mode of the Na<sup>+</sup>/Ca<sup>2+</sup> exchanger (NCX), which extrudes Na<sup>+</sup> in exchange for Ca<sup>2+</sup> entry. The resultant cytosolic loading of Ca<sup>2+</sup> brings the progression of ischemic injury to an advanced stage by inducing (a) sustained impairment of contractile function; (b) cell membrane damage by activating Ca<sup>2+</sup>-dependent phospholipase, which generates reactive oxygen species (ROS) from myocytes, ECs, and activated leukocytes;

TABLE 1. COMPARISON OF THE CARDIOPROTECTIVE EFFECTS OF SODIUM HYDROSULFIDE AGAINST ISCHEMIC/REPERFUSION-INDUCED INJURY

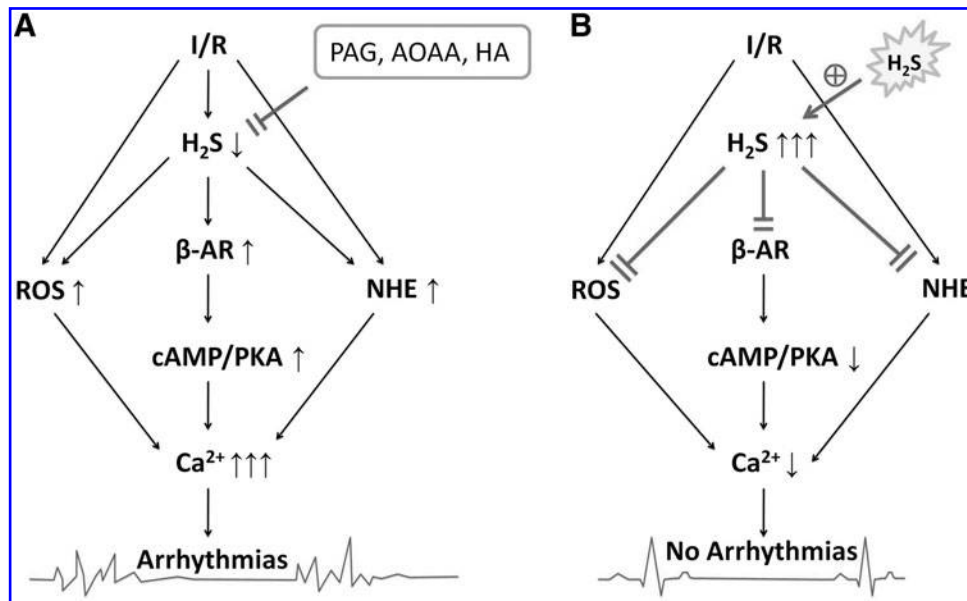
<i>Treatment</i>	<i>I/R protocol</i>	<i>Species/tissue</i>	<i>Effects of NaHS</i>	<i>Mechanism</i>	<i>Ref.</i>
NaHS (0.1 and 1 $\mu$ M perfusion 10 min prior to LAD occlusion till 10 min reperfusion	I (30 min)/R (120 min)	Rats/Langendorff heart	MI ( $\downarrow$ )	K <sub>ATP</sub> channel	(104)
NaHS (40 $\mu$ M) throughout the experiment	I (40 min)/R (120 min)	Rats/Langendorff heart	MI ( $\leftrightarrow$ )	—	(20)
PAG	I (40 min)/R (120 min)	Rats/Langendorff heart	MI ( $\uparrow$ )		
NaHS (40 $\mu$ M) perfusion during reperfusion	I (30 min)/R (30 min)	Rats/Langendorff heart	Antiarrhythmias, improve contractile function	K <sub>ATP</sub> channel	(306)
NaHS (14 $\mu$ mol/kg/day) i.p. from 7 days before to 2 days after MI surgery	Permanent ligation w/o reperfusion	Rats/ <i>in vivo</i>	MI ( $\downarrow$ ), mortality ( $\downarrow$ )	—	(315)
NaHS (0.1, 1, 10 $\mu$ mol/kg/day) i.p. for 3 days after MI surgery	Permanent ligation w/o reperfusion	Rats/ <i>in vivo</i>	MI ( $\downarrow$ ), internal diameter ( $\downarrow$ ), Anterior wall thickness ( $\uparrow$ )	—	(187)
NaHS (10–500 $\mu$ g/kg) administered into LV lumen at the time of reperfusion; CSE overexpression	I (30 min)/R (24 h)	Male C57BL6/J mice or CSE transgenic mice/ <i>in vivo</i>	MI ( $\downarrow$ ), apoptosis ( $\downarrow$ ), inflammation ( $\downarrow$ )	Preserve mitochondrial function, improve recovery of respiration rate, antiapoptosis, Anti-inflammation	(61)
Bolus: NaHS (0.2 mg/kg) over 10 s at the onset of ischemia; Infusion: NaHS (2 mg/kg/h) during I/R period	I(60 min)/R (120 min)	Swine/ <i>in vivo</i>	Bolus: no effect Infusion: MI ( $\downarrow$ ),	Hsp27, $\alpha$ B-crystallin, phosphor-glcogen synthase kinase-3 $\beta$ , antiapoptosis	(186)
Na <sub>2</sub> S: bolus (NaHS, 100 $\mu$ g/kg)+infusin (NaHS, 1 mg/kg)	I(60 min)/R (120 min)	Swine/ <i>in vivo</i>	MI ( $\downarrow$ ), improve contractile function and coronary microvascular reactivity	Anti-inflammation	(231)
NaHS: 100 $\mu$ M perfusion 10 min before and during ischemia in the isolated heart	I (30 min)/R (60 min)	Rat/Langendorff heart	Improve contractile function and increase cell viability	Inhibition of NHE	(92)
Na <sub>2</sub> S: 10 min prior to and through reperfusion	I(60 min)/R (120 min)	Swine/ <i>in vivo</i>	MI ( $\downarrow$ )	Antiapoptosis	(230)
NaHS: 3 mg/kg, i.v.	I(25 min)/R (120 min)	Rat/ <i>in vivo</i>	MI ( $\downarrow$ )	K <sub>ATP</sub>	(226)
PAG: 50 mg/kg, i.v.	I(15 min)/R (120 min)	Rat/ <i>in vivo</i>	MI ( $\uparrow$ )	—	

CSE, cystathionine  $\gamma$ -lyase; i.p., intraperitoneal; i.v., intravenous; I/R, ischemia/reperfusion; K<sub>ATP</sub>, ATP-sensitive potassium; LAD, left anterior descending; LV, left ventricle; MI, myocardial infarction; NaHS, sodium hydrosulfide (H<sub>2</sub>S donor); NHE, Na<sup>+</sup>/H<sup>+</sup> exchanger; PAG, propargylglycine.

and (c) disruption of cellular scaffolds by activating Ca<sup>2+</sup>-dependent proteases that cleave cytoskeletal filaments. Collectively, these Ca<sup>2+</sup>-induced changes lead to a loss of membrane integrity and terminally demolished cellular structure (25).

The re-establishment of coronary blood flow to the infarcted area results in reperfusion injury, mainly mediated by ROS, overloaded Ca<sup>2+</sup>, and neutrophils (38, 81, 191). Single-electron reduction of oxygen in injured myocytes, ECs, and activated neutrophils generates the superoxide anion (O<sub>2</sub><sup>-</sup>) (54), which goes on to generate other ROS. These free radicals exacerbate membrane damage and stimulate vasoconstriction, which, in severe cases, results in a “no flow” phenomenon. The Ca<sup>2+</sup> overload induces maximum contraction of the

myofibrils on reperfusion, resulting in contraction band necrosis (252). An increase in mitochondrial [Ca<sup>2+</sup>] triggers the opening of mitochondrial permeability transition pores (mPTP), causing the release of cytochrome C and other proapoptotic factors that initiate the apoptotic cascade (85). Reperfusion is a potent stimulus for neutrophil activation and accumulation, which, in turn, enhances ROS production (44). In myocardium subjected to I/R, reoxygenated ECs express adhesion proteins, release cytokines, and reduce the production of NO, which promotes adherence, activation, and accumulation of neutrophils (62, 105). These activated neutrophils will release not only ROS but also proteolytic enzymes that can damage myocytes and vascular cells. Proinflammatory



**FIG. 6. Role of H<sub>2</sub>S in ischemia/reperfusion (I/R)-induced arrhythmias.** (A) On subjecting the myocardium to I/R, endogenous H<sub>2</sub>S synthesis was found to be reduced. This leads to augmented  $\beta$ -adrenoceptor signaling, elevated reactive oxygen species (ROS) production, and enhanced action of NHE. Collectively, these events brought about the elevation of intracellular calcium and resulted in arrhythmia. When the inhibitors of endogenous H<sub>2</sub>S synthesis [e.g., DL-propargylglycine (PAG; a CSE inhibitor), amino-oxycacetate (AOAA; a CBS inhibitor) and hydroxylamine (HA; an inhibitor of both CBS and CSE)], were utilized, the extent of arrhythmia worsened. (B) Exogenously applied H<sub>2</sub>S could reverse the effect of I/R on arrhythmias.

lipid metabolites released from neutrophils and other white blood cells enhance the expression and production of a proinflammatory cytokine cascade involving interleukin (IL)-1 and tumor necrosis factor- $\alpha$  (TNF $\alpha$ ) (49). These cytokines then lead to the activation and infiltration of leukocytes. Proinflammatory molecules (such as IL-6, IL-8) and anti-inflammatory factors (including IL-4 and IL-10) may also be produced and act as negative feedbacks on the cascade (105).

(1). **Antioxidative action.** I/R-induced arrhythmias may develop as a result of ROS production and accumulation in the myocardium during reperfusion. Since H<sub>2</sub>S production is markedly decreased during ischemia (17, 73, 103, 297, 298), ROS may, therefore, be increased. Excessive free radicals can react with proteins, lipids, and nucleic acids, thereby disrupting myocardium functions. The application of H<sub>2</sub>S may produce protective effects *via* its antioxidative effects (232). In cultured H9c2 myoblasts, Na<sub>2</sub>S (30–100  $\mu$ M) was shown to protect against free-radical (H<sub>2</sub>O<sub>2</sub>)-induced cell death (242). However, unlike GSH, which is present at a 1–10 mM concentration, H<sub>2</sub>S is present at relatively low concentrations (~15 nM) and it is also a poorer reductant (redox potential of +0.17 V *vs.* -0.25 V for the other two thiols) (106). Hence, the physiological relevance of H<sub>2</sub>S as an endogenous antioxidant is still a question. The effect of H<sub>2</sub>S on GSH activity or level has been previously reviewed (93). Heme oxygenase (HO)-1, the rate-limiting enzyme involved in heme degradation, has been implicated in both pro- and anti oxidative activities. H<sub>2</sub>S infusion in porcine LCA occlusion was found to enhance HO-1 expression (186). The significance of this effect is yet to be understood.

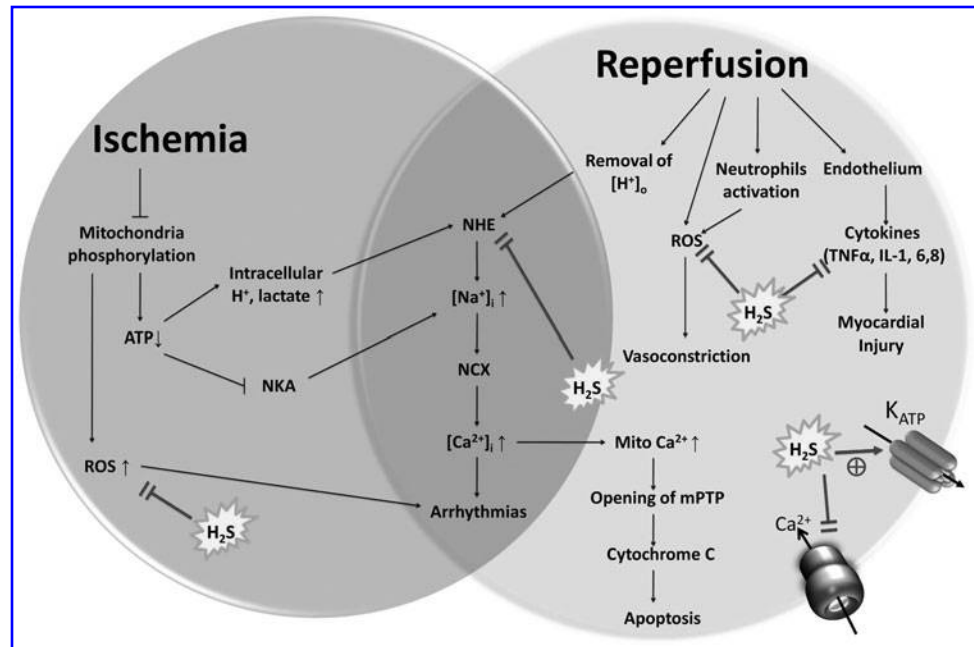
(2). **Suppression of  $\beta$ -adrenergic function.** Overactivation of the sympathetic system is closely linked with the pro-

gression of cell injury and the incidence of ventricular arrhythmias during myocardial ischemia (211, 212). Excessive norepinephrine release and accumulation in the heart may cause the depletion of ATP and the accumulation of intracellular Ca<sup>2+</sup> and Na<sup>+</sup> (39) that can lead to ventricular arrhythmias (185). Yong *et al.* found that NaHS (100  $\mu$ M) negatively modulated  $\beta$ -adrenergic function (298) and the lowered H<sub>2</sub>S production during ischemia may cause overstimulation of the  $\beta$ -adrenergic function. Therefore, exogenous application of H<sub>2</sub>S may antagonize the negative consequences of sympathetic overactivation during ischemia by generating negative feedback to cAMP production. In this regard, H<sub>2</sub>S replacement therapy may be a significant cardioprotective and antiarrhythmic intervention for those patients with chronic ischemic heart disease whose plasma H<sub>2</sub>S level is reduced (103) (Fig. 7).

(3). **Inhibition of NHE.** Intracellular pH (pH<sub>i</sub>) is an important endogenous modulator of cardiac function. The inhibition of NHE-1 protects the heart by preventing Ca<sup>2+</sup> overload during I/R. H<sub>2</sub>S was reported to produce cardioprotection. Hu *et al.* recently reported that NaHS (100  $\mu$ M) may induce intracellular acidosis *via* suppression of NHE-1 and further produce cardioprotection. This effect was mediated by the activation of phosphoinositide 3-kinase (PI3K)/Akt/protein kinase G (PKG) (92). This action further offers cardioprotective effects against I/R-induced injury.

(4). **Opening of K<sub>ATP</sub> channels and/or blockade of Ca<sup>2+</sup> channels.** The opening of sarcolemmal and/or putative mitochondrial K<sub>ATP</sub> channels has been extensively documented as mediating the protection against I/R injury. Johansen *et al.* first tested the involvement of K<sub>ATP</sub> channels in the cardioprotection of NaHS (104). The injury-limiting action

**FIG. 7. H<sub>2</sub>S-induced protection against I/R injuries.** By inhibiting Na<sup>+</sup>/H<sup>+</sup> exchanger (NHE) activity, H<sub>2</sub>S brings about a decrease in intracellular Ca<sup>2+</sup> and the attenuation of arrhythmias induced by both ischemia and reperfusion. Furthermore, H<sub>2</sub>S may further protect against ischemia- and reperfusion-induced injury by attenuating the accumulation of oxidative stress. H<sub>2</sub>S was also found to inhibit the cytokines that are responsible for reperfusion-induced myocardial injury. H<sub>2</sub>S actions on the K<sub>ATP</sub> channels, and the Ca<sup>2+</sup> channels further protect the myocardium against reperfusion-induced injuries.



of NaHS was abolished by glibenclamide, a general nonselective blocker of K<sub>ATP</sub>, or 5-hydroxydecanoate (5-HD, a selective mitochondrial K<sub>ATP</sub> channel blocker), suggesting that the K<sub>ATP</sub> channels play a central role in the cytoprotective action of H<sub>2</sub>S (104). Activation of the K<sub>ATP</sub> channels induces K<sup>+</sup> efflux, which causes hyperpolarization. This may close LTCC. LTCC can also be closed by the H<sub>2</sub>S-suppressed cAMP/PKA pathway (298). Alternatively, H<sub>2</sub>S may directly block LTCC (239). All these may further reduce Ca<sup>2+</sup> influx and shorten APD to produce an antiarrhythmic effect (Fig. 4).

(5). *Anti- and proinflammatory factors.* Cytokines mediate the development of ischemic injury in the heart (194). IL-6 and IL-8 are released on myocardial I/R damage. The former depresses myocardial function (90), whereas the latter increases neutrophil adhesion and inflammatory responses (122). TNF- $\alpha$  may play multiple roles in the pathogenesis of myocardial I/R injury by inducing EC adhesion molecules, allowing for neutrophil infiltration (55), amplifying the inflammatory response, increasing the production of ROS, and having direct myocardial depressant and apoptotic actions.

The effect of H<sub>2</sub>S on inflammation is dubious. Whiteman and Winyard reviewed 14 studies showing an anti-inflammatory effect of H<sub>2</sub>S, and 15 studies showing a proinflammatory effect of H<sub>2</sub>S (276). It has been reported that H<sub>2</sub>S may increase intercellular adhesion molecule-1 (ICAM-1) level in blood vessels (305) and stimulate the production of proinflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$ , and IL-6) *via* activation of the extracellular signal-regulated kinase (ERK)-NF- $\kappa$ B signaling pathway in human monocytes (313). In animal experiments, the biosynthesis of H<sub>2</sub>S was found to be increased along with inflammation-associated septic shock, hemorrhagic shock, and pancreatitis, while the inhibition of H<sub>2</sub>S biosynthesis alleviated such inflammation (132).

On the other hand, anti-inflammatory effects of H<sub>2</sub>S have also been displayed by its ability to interfere with neutrophil activity, suppress granulocyte adherence to the endothelium and infiltration, and suppress the expression of TNF- $\alpha$  (232,

272). In myocardial I/R experiments, Elrod *et al.* have demonstrated that at the time of heart reperfusion, H<sub>2</sub>S decreased the number of leukocytes within the ischemic zone (61). Quantitative myeloperoxidase (MPO) analysis further confirmed a significant decrease in neutrophils within the myocardial tissue after I/R. Intravital microscopy demonstrated that H<sub>2</sub>S was a potent inhibitor of *in vivo* leukocyte-EC interactions. The valuation of inflammatory cytokines revealed that H<sub>2</sub>S decreased myocardial IL-1 $\beta$  (61). Using the ischemic porcine heart, Sodha *et al.* found that NaHS treatment decreased TNF- $\alpha$ , IL-6, and IL-8 levels. Therefore, inhibition of leukocyte transmigration and inhibition of cytokine release are possible mechanisms by which H<sub>2</sub>S restrains the extent of inflammation, thereby limiting the extent of MI (231). Wallace *et al.* recently reviewed the role of endogenous H<sub>2</sub>S in resolving inflammation and injury (254). The authors suggest that H<sub>2</sub>S-releasing agents may be clinically useful.

However, since the effects of H<sub>2</sub>S on inflammation are conflicting, more work has to be done to fully understand its biology and implications.

(6). *Preservation of mitochondrial function.* In dogs subjected to 90 min of cardiopulmonary bypass (CPB) and 60 min of cardiac arrest, Na<sub>2</sub>S (1 mg/kg/h infusion) improved ventricular functions by preserving cardiac ATP pools (242). In addition to being the site of energy production, mitochondria are a central locus in the regulation of cell death. The maintenance of oxidative phosphorylation for preventing myocyte death after ischemic injury has long been recognized as a critical event after MI. Elrod and colleagues found a dose-dependent reduction in mitochondrial oxygen consumption followed by a complete recovery to baseline level after the administration of H<sub>2</sub>S (1–50  $\mu$ M) (61). H<sub>2</sub>S preserves mitochondrial function 24 h after reperfusion as noted by increased complex I and II efficiency (6). Moreover, electron microscopy revealed a striking reduction in mitochondrial swelling and increased matrix density in H<sub>2</sub>S treated mice, further suggesting a prominent role of H<sub>2</sub>S in the preservation

of mitochondrial function in the observed cytoprotection (61). H<sub>2</sub>S may also protect mitochondrial function by inhibiting respiration, thus limiting the generation of ROS and diminishing the degree of mitochondrial uncoupling, leading to decreased infarct size and preserved function (61). Intriguingly, reports demonstrate that a low concentration of H<sub>2</sub>S (<20  $\mu$ M) stimulates oxygen consumption and increases membrane potential (80, 299). The endogenous role of H<sub>2</sub>S *in vivo*, therefore, is yet to be fully understood.

(7). **Antiapoptosis.** The preservation of mitochondrial function and structure is linked to the inhibition of cell apoptosis. Early in apoptotic signaling, the antiapoptotic B-cell lymphoma 2 (Bcl-2) and proapoptotic Bcl-2-associated death promoter (Bad) are in opposition. A shift in favor of the proapoptotic proteins subsequently results in increased mitochondrial pore permeability, releasing cytochrome C and apoptosis-inducing factor (AIF), which can activate caspase-3, and facilitate DNA fragmentation. On cleavage of terminal caspases, cell death is generally thought to be inevitable.

It was found that NaHS (10–500  $\mu$ g/kg) treatment suppressed the activation of caspase-3, poly (ADP-ribose) polymerase and/or terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL)-positive nuclei in both mice (61) and swine (230), suggesting that H<sub>2</sub>S was capable of inhibiting the progression of apoptosis after I/R injury.

However, Osipov *et al.* found that infusion of H<sub>2</sub>S (IK-1001, 2 mg/kg/h), while exerting cardioprotective effects, failed to significantly alter the activation of caspase-3, Bcl-2, and Bad. A single bolus injection of IK-1001 just before infusion led to increased Bcl-2 and Bad, though the elevation did not reach statistical significance among six pigs per group being examined. These results indicate that H<sub>2</sub>S administration by bolus injection may trigger Bcl-2 and Bad, but these do not underlie the protective effects seen in H<sub>2</sub>S infusion alone (186). AIF and Bnip-3 (Bcl-2/adenovirus E1B 19 kDa-interacting protein), on the other hand, were significantly reduced on H<sub>2</sub>S treatments in both infusion alone and bolus injection before infusion groups, suggesting that H<sub>2</sub>S more likely attenuates the expression of proapoptotic proteins *via* caspase-independent cell death (186).

Survivin is an antiapoptotic gene implicated in the initiation of mitochondrial-dependent apoptosis. In an *in vivo* ischemic reperfusion rat model, the administration of NaHS (14  $\mu$ mol/kg/day) for 6 days before surgery significantly upregulated survivin mRNA and protein expressions by 3.4-fold and ~1.7-fold, respectively (317), suggesting another route of action for H<sub>2</sub>S-induced cardioprotection.

Glycogen synthase kinase-3 (GSK-3 $\beta$ ) has been proposed as a viable target in the ischemic injury of the heart. Its activity has been associated with both apoptosis and cell survival. Osipov *et al.* found that H<sub>2</sub>S infusion leads to a higher expression of the phosphorylated form of GSK-3 $\beta$  (186). Similarly, Yao *et al.* also demonstrated that NaHS (30  $\mu$ mol/kg) increased the phosphorylation of GSK-3 $\beta$  (Ser9) and, thus, inhibited the opening of mPTP, preventing apoptosis and protecting the heart against ischemic damage (291).

(8). **Preservation of endothelial function.** In the canine CPB model, Na<sub>2</sub>S (1 mg/kg/h) infusion restored the sensitivity of coronary arteries to acetylcholine (ACh)-induced vasorelaxation, suggesting that H<sub>2</sub>S improves conditions of endothelial dysfunction during cardiac injury (242). Similarly,

in a porcine CPB model, H<sub>2</sub>S treatment (IK-1001 0.2 mg/kg bolus injection and infusion 2 mg/kg/h) improved endothelium-dependent coronary arteriole relaxation (186).

(9). **Attenuation of endoplasmic reticulum stress.** Hyperhomocysteinemia (HHcy) is an independent risk factor for CVD (21, 40, 50, 162, 216, 234). HHcy is a medical condition characterized by an abnormally high level of homocysteine in circulation, and homocysteine at a higher level was reported to inhibit CSE activity (290), causing a reduction in H<sub>2</sub>S production *via* substrate inhibition, as homocysteines contribute to H<sub>2</sub>S synthesis only to a small extent as compared with cysteines (41, 235). Wei *et al.* showed that NaHS (14  $\mu$ mol/kg/day) protected the heart against HHcy-induced cardiomyocyte injury *via* attenuation of cardiomyocyte endoplasmic reticulum stress-associated proteins such as glucose-regulated protein 78, C/EBP homologous protein, and caspase-12 (269).

2. **Cardioprotection caused by H<sub>2</sub>S preconditioning.** IPreC is a powerful natural cardioprotective mechanism, and was later reported to offer similar protective effects in various other systems (*e.g.*, liver, kidney, brain, and ECs). Bian *et al.* first reported that H<sub>2</sub>S preconditioning (SPreC) protects the heart against I/R-induced arrhythmias, cell injuries and death, contractile dysfunction, and MI (17, 94, 96, 187–189). Subsequently, Calvert and colleagues confirmed that SPreC produces cardioprotection against MI-induced injury in mice (36). A more detailed summary is presented in Table 2.

Pan *et al.* compared the cardioprotective effects caused by SPreC and postischemic treatment. The infarct per area at risk in the SPreC group was significantly lower than those in the postischemic H<sub>2</sub>S treatment group (187). In addition, it was found that H<sub>2</sub>S direct treatment had no significant effect on sarcoplasmic/endoplasmic reticulum Ca<sup>2+</sup> ATPase (SERCA) function (298), whereas SPreC significantly stimulated SERCA activity (189). These findings suggest that apart from the mechanisms discussed in Mechanisms for H<sub>2</sub>S-induced cardioprotection section, SPreC may stimulate additional pathways that produce stronger protective effects.

The beneficial effects of postischemic H<sub>2</sub>S treatment may rely mainly on the ability of sulfide to reduce inflammatory responses (302) and to neutralize cytotoxic reactive species such as peroxynitrite (ONOO<sup>−</sup>) (272), which may partly relieve the oxidative stress but is not likely to rescue already infarcted myocardium (See mechanisms discussed in Cardioprotective effects of H<sub>2</sub>S against ischemia-reperfusion-induced injury section). However, SPreC is more likely to protect by switching the heart to a defensive state against ischemic insult. For example, the activation of protein kinases (*e.g.*, PKC, MAPK, and Akt) would lead to the phosphorylation of many cardioprotective proteins (*e.g.*, heat shock proteins [HSPs], cyclooxygenase [COX]-2, and Bcl-2) (Fig. 8).

*a. Mechanisms for the cardioprotection of SPreC.* A series of experiments was conducted to probe the signaling mechanism(s) involved in the late cardioprotection of SPreC. The mechanisms are discussed next.

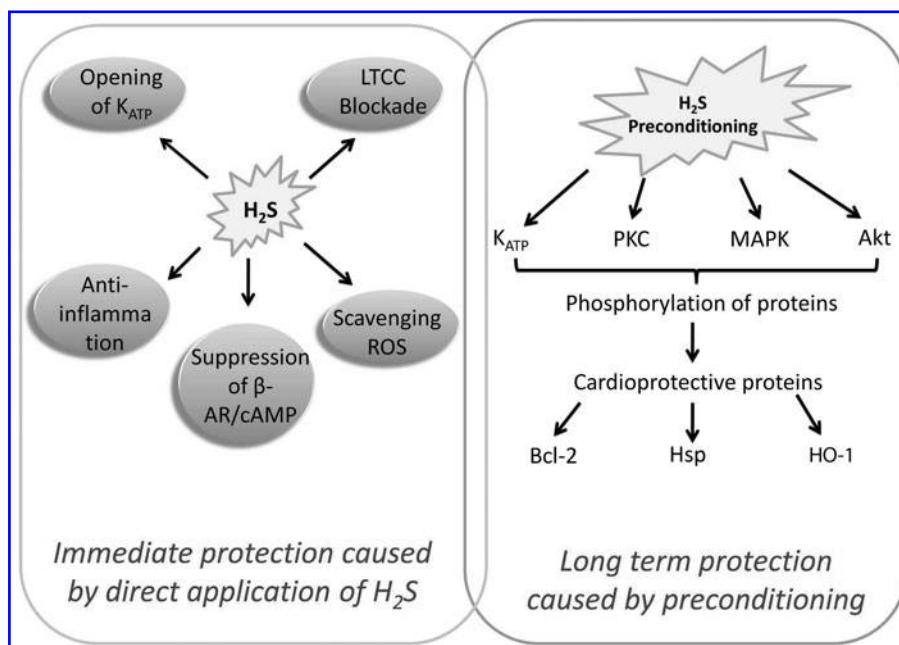
(1). **K<sub>ATP</sub> channels.** The involvement of the K<sub>ATP</sub> channels in the cardioprotection of preischemic H<sub>2</sub>S treatment was first demonstrated by Pan *et al.* (188). Opening of the K<sub>ATP</sub> channels affords protection by shortening APD and by

TABLE 2. CARDIOPROTECTION OF HYDROGEN SULFIDE PRECONDITIONING AGAINST ISCHEMIC INJURY

<i>H<sub>2</sub>S</i> Preconditioning	<i>I/R protocol</i>	<i>Species/tissue</i>	<i>Effects of NaHS</i>	<i>Mechanism</i>	<i>Ref.</i>
Late: After preconditioning with NaHS (100 $\mu$ M) for 30 min, cells were cultured in normal medium for 20 h	I (5 min)/R (10 min)	Rats/ cardiomyocytes	Cell viability ( $\uparrow$ ), LDH ( $\downarrow$ ), improvement of calcium handling	K <sub>ATP</sub> , NO	(188)
Early: Three cycles (NaHS 100 $\mu$ M for 3 min each cycle separated by 5 min of recovery)	I (30 min)/R (10 min)	Rats/ cardiomyocytes	Antiarrhythmias, Cell viability ( $\uparrow$ ), improvement of [Ca <sup>2+</sup> ] <sub>i</sub> handling	K <sub>ATP</sub>	(17)
Late: After preconditioning with NaHS (100 $\mu$ M) for 30 min, cells were cultured in normal medium for 20 h	I (5 min)/R (10 min)	Rats/ cardiomyocytes	Cell viability ( $\uparrow$ ), LDH ( $\downarrow$ ), improvement of contractile function	COX-2/PGE2	(94)
Late: After preconditioning with NaHS (100 $\mu$ M) for 30 min, cells were cultured in normal medium for 20 h	I (5 min)/R (10 min)	Rats/ cardiomyocytes	Cell viability ( $\uparrow$ ), improvement of [Ca <sup>2+</sup> ] <sub>i</sub> handling	PKC	(189)
Early: Three cycles (NaHS 100 $\mu$ M for 3 min each cycle separated by 5 min of recovery)	I (35 min)/R (60 min)	Rats/ Langendorff hearts	Antiarrhythmias, Cell viability ( $\uparrow$ ), improvement of contractile function	ERK, Akt	(96)
Late: NaHS (0.1–1 $\mu$ mol/kg i.p.) 1, 3 or 5 day before MI	Permanent MI	Rats/ <i>in vivo</i>	MI ( $\downarrow$ )	PKC	(187)
Early: Na <sub>2</sub> S (100 $\mu$ g/kg i.v.) 30 min or 2 h before MI	I(45 min)/R (24 h)	mice/ <i>in vivo</i>	MI ( $\downarrow$ )	Early: Antioxidant (Nrf2), PKC $\epsilon$ , STAT-3 Late: Antioxidants (Heme oxygenase-1 and thioredoxin 1), hsp90, 70, antiapoptosis, COX-2	(36)

COX, cyclooxygenase; H<sub>2</sub>S, hydrogen sulphide; LDH, lactate dehydrogenase; NO, nitric oxide; Nrf2, nuclear factor-erythroid-derived 2 related factor 2; PKC, protein kinase C; STAT, signal transducer and activator of transcription.

**FIG. 8. Different mechanisms underlying the cardioprotection produced by direct application of H<sub>2</sub>S and H<sub>2</sub>S preconditioning.** The acute effect of H<sub>2</sub>S-induced cardioprotection involves the actions of the K<sub>ATP</sub> channel opening, blockade of LTCC, reduction of ROS, inhibition of the  $\beta$ -adrenoceptor/cAMP pathway, and anti-inflammatory responses. In addition, H<sub>2</sub>S offers chronic protection *via* numerous signaling mechanisms involving the protein kinase C (PKC) pathway, mitogen-activated protein kinase (MAPK) pathway, serine/threonine protein kinase Akt pathway, and the opening of K<sub>ATP</sub> channels. A cascade of proteins would then be phosphorylated, and cardioprotective proteins, including B-cell lymphoma 2 (Bcl-2), heat shock proteins (Hsp), and heme oxygenase-1 (HO-1), would be regulated.



preventing calcium overload (83) as just discussed in the  $K_{ATP}$  channels section. However, the  $K_{ATP}$  channel opening also exerts a cardioprotective effect *via* more complicated crosstalk with other rescuing pathways. The opening of the  $K_{ATP}$  channels and activation of protein kinase C (PKC) have been shown to be co-dependent during IPreC (72, 139). The crosstalk among the  $K_{ATP}$  channels and protein kinases (see next section) suggests that, similar to other preconditioning events, SPreC may orchestrate a vast network of rescuing pathways that execute the prosurvival signals.

(2). *Protein kinase C.* The PKC family consists of at least 10 isoforms, of which PKC- $\alpha$ ,  $\epsilon$ , and  $\delta$  are the prominent isoforms expressed in the heart (157). Pan *et al.* found that the SPreC induced the translocation of all three prominent isoforms of PKC from the cytosol to the membrane (189). Such translocation before an ischemic attack may offer the cells tolerance to the insults.

H<sub>2</sub>S was reported as exerting a direct effect on the  $K_{ATP}$  channels (245, 309); hence, the effect of SPreC on PKC might only be secondary to the opening of the  $K_{ATP}$  channels. However, Pan and colleagues observed that blockade of the  $K_{ATP}$  channel by glibenclamide not only blocked SPreC-induced translocation of PKC $\epsilon$ , but also failed to affect the translocation of PKC $\alpha$  and  $\delta$  (189). Thus, the  $K_{ATP}$  channel opening may only activate PKC $\epsilon$  translocation in the SPreC signaling pathway. PKC can also be activated by other signaling molecules such as NO or Ca<sup>2+</sup> (168, 193). More studies are warranted to test whether SPreC induces activation of PKC $\alpha$  and PKC $\delta$  through provoking the release of these signaling molecules.

(3). *Prevention of intracellular calcium overload and hypercontracture.* A timely reduction of the elevated [Ca<sup>2+</sup>]<sub>i</sub> during an ischemic challenge could be of therapeutic importance, as Ca<sup>2+</sup> overload could induce irreversible injuries such as mitochondria dysfunction (166), membrane degradation, and contractile derangement (82). Pan *et al.* found that SPreC lowered the elevation of [Ca<sup>2+</sup>]<sub>i</sub> during ischemia. This is mediated by the stimulatory effect of H<sub>2</sub>S on SERCA2 and NCX (189).

Elevated [Ca<sup>2+</sup>]<sub>i</sub> may also stimulate myocytes that cause hypercontracture as determined by cell length shortening a few minutes after the onset of reperfusion (224). Pan *et al.* found that SPreC significantly attenuated I/R-induced myocyte detrimental shortening (189). This protective effect of H<sub>2</sub>S is PKC dependent (189).

(4). *NO.* In the past decade, many studies have revealed a critical role of NO in IPreC-induced cardioprotection (22). Importantly, NO alone is also sufficient to induce late cardioprotection against myocardial ischemia (243). Pan *et al.* found that the inhibition of NO production with N (G)-nitro-L-arginine methyl ester (L-NAME), a nonselective inhibitor of all NOSs, significantly attenuated the cardioprotective effects of SPreC, but not those of IPreC. Therefore, NO may well be the main reason that NaHS produced a stronger protective effect on cell viability in SPreC than that in IPreC (188).

The administration of NaHS (50  $\mu$ mol/kg/day) 1 h before ISO-induced toxic cardiomyopathy produced cardioprotection along with significantly increased myocardial and serum NO levels (232). More evidently, the coadministration of NOS inhibitor (L-NAME) reversed the protective effects of H<sub>2</sub>S,

indicating that H<sub>2</sub>S-induced cardioprotection may be mediated by NO.

(5). *ERK1/2-MAPK.* ERK 1 and 2 belong to the MAPKs family, which includes ERK1/2 and stress-activated protein kinases containing Jun N-terminal Kinase 1/2 and p38-MAPK. It is commonly accepted that phosphorylation of ERK1/2 in cardiomyocytes during early reperfusion serves as a defense mechanism against ischemic stress stimuli (301). This defense mechanism can be magnified by IPreC and several forms of pharmacologically induced preconditioning (86, 248). Hu *et al.* observed that SPreC protected the heart against I/R injury by reducing myocardial injury (96). The blockade of ERK1/2 with PD98059 during either preconditioning or ischemia periods reversed these cardioprotective effects, suggesting that ERK1/2 may at least partly mediate the cardioprotection afforded by SPreC (96). This was further confirmed by the demonstrable phosphorylation of ERK1/2 (96).

(6). *PI3K/Akt.* The activation of the PI3K/Akt pathway has been demonstrated as playing a key role in both early and delayed myocardial preconditioning (86–88). Hu *et al.* found that SPreC induced a significant phosphorylation of Akt in the isolated rat hearts, and, more importantly, the cardioprotective effects of SPreC were greatly attenuated by inhibition of PI3K and Akt during preconditioning and ischemia (96). These data suggest that, similar to ERK1/2, PI3K/Akt may serve as a trigger and mediator in SPreC-induced cardioprotection.

(7). *COX-2/PGE2 pathway.* COX, the rate-limiting enzyme in prostaglandin synthesis, catalyzes the conversion of arachidonic acid (AA) to prostaglandin H<sub>2</sub>. Two distinct COX isoforms have been characterized. COX-1 is constitutive and COX-2 is usually induced in response to stress and inflammation. There is accumulating evidence that COX-2 plays an essential role in mediating the late phase, but not the early phase, of IPreC-induced cardioprotection (261, 293).

Hu *et al.* found that the blockade of COX-2 with two selective inhibitors, NS-398 and celecoxib, attenuated SPreC-induced cardioprotection (94). Moreover, SPreC significantly increased PGE<sub>2</sub> formation at the end of lethal ischemia, indicating that PGE<sub>2</sub> is likely to be one of the mediators of COX-2-dependent protection produced by SPreC (94). The administration of another selective COX-2 inhibitor, Celecoxib, abolished the protective effects of NaHS (50  $\mu$ mol/kg/day) pretreatment in ISO-induced MI (232). Collectively, the current evidence implies that COX-2 may mediate the cardioprotective effects of H<sub>2</sub>S.

(8). *Nuclear factor-erythroid-derived 2 related factor 2/antioxidant.* Nuclear factor-erythroid-derived 2 (NF-E2) related factor 2 (Nrf2), a member of the NF-E2 family of nuclear basic leucine zipper transcription factors, regulates the gene expression of a number of enzymes that serve to detoxify pro-oxidative stressors. This regulation is mediated by Nrf2 binding to the antioxidant responsive element, a *cis*-acting regulatory element, or enhancer sequence, found in the promoter region of certain genes, including HO-1 and thioredoxin-1 (Trx1). During I/R, the activities of many endogenous antioxidant enzyme systems are compromised or even abolished. Calvert *et al.* found that H<sub>2</sub>S (100  $\mu$ g/kg) induced

the nuclear accumulation of Nrf2 very rapidly after its administration and subsequently increased the protein expression of HO-1 and Trx1 (36). These results suggest that H<sub>2</sub>S therapy may enhance the endogenous antioxidant defenses of myocytes and create an environment resistant to the oxidative stress associated with myocardial I/R injury, as evidenced by the preservation of redox state and a reduction in lipid peroxidation.

The ability of a cell to counteract stressful conditions, known as cellular stress response (30, 33), requires the activation of prosurvival pathways. These pathways, under the control of protective genes called vitagenes (31), result in the production of various molecules (*e.g.*, HSPs, GSH, and bilirubin) endowed with antioxidant and antiapoptotic activities (32). The vitagene network relevant to cytoprotection could be a potential target for the cardioprotective effects of H<sub>2</sub>S.

(9). **Antiapoptosis.** Myocardial I/R induces apoptosis and necrosis. Calvert *et al.* found that SPreC preserved un-cleaved caspase-3, decreased cleaved caspase-3, inhibited the translocation of cytochrome C, and decreased the number of TUNEL-positive nuclei (36). The antiapoptotic actions were mediated in part by the phosphorylation and inhibition of the proapoptotic factor Bad, an upregulation of the pro-survival factors Bcl-2 and Bcl-xL, and an upregulation of HSPs. During the early preconditioning period (30 min and 2 h), H<sub>2</sub>S activated PKC $\epsilon$ , ERK1/2-MAPK, and signal transducer and activator of transcription (STAT)-3. The STAT pathway has been shown to be an integral part of the response of the myocardium to various cardiac insults, including MI (9).

(10). **Heat shock proteins.** HSPs have also been demonstrated as providing cardioprotection in the setting of I/R. In particular, HSP70 suppresses apoptosis in a caspase-dependent (207) and caspase-independent manner (202). During the late preconditioning period (24 h), SPreC increased the expression of HSP90 and HSP70 (36). These data suggest that SPreC not only reduces apoptosis through a reduction in oxidative stress, but also promotes direct antiapoptotic signaling.

In contrast, a bolus injection of Na<sub>2</sub>S (IK-1001, Ikaria Inc) before CPB together with continuous IK-1001 infusion resulted in reduced expressions of HSP70, enhanced expression of HO-1, and no significant difference in HSP90 expression (186). The controversies in findings, therefore, warrant further investigation and in-depth studies.

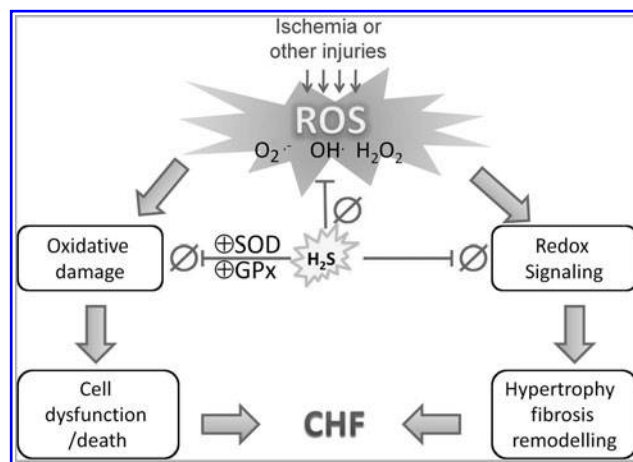
Most concentrations used in SPreC in the *in vitro* studies just mentioned were within 10–100  $\mu$ M. Interestingly, it was found that with concentrations higher than 100  $\mu$ M, the cardioprotection induced by SPreC may be weaker. The possible explanation for the phenomenon just mentioned might be the involvement of hormesis (30), which refers to generally favorable biological responses to low exposures to many stressors (29). H<sub>2</sub>S showing hormesis, thus, has the opposite effect in small doses as in large doses. The biochemical mechanisms for hormesis are still not well understood. It is conjectured that low doses of H<sub>2</sub>S or other stressors might activate the repair mechanisms of the body. The repair process fixes not only the damage caused by the toxin, but also other low-level damage that might have accumulated before the repair mechanism was triggered.

## B. Heart failure and other related heart diseases

CHF is a growing health concern. In the United States, for example, it affects 5.2 million Americans with more than 400,000 new cases being diagnosed each year (204). In 2007, it has been associated with one in every nine deaths (204). However, existing treatments are far from adequate, and the search for new treatment strategies is crucial and urgently needed.

The main pathological mechanism for CHF appears to be oxidative stress (Fig. 9). When the local levels of ROS are high, they tend to react with numerous protein centers, DNA, cell membranes, and other molecules, causing considerable cellular damage as well as generating other more reactive radicals (215). At lower concentrations, however, local targeted ROS serves as a second messenger system that transmits biological information through the highly specific modulation of small intracellular molecules and proteins (215). Redox signaling processes are involved in the activation of many signal transduction protein kinases and transcription factors, the stimulation of DNA synthesis, and the expression of growth-related genes. These effects are relevant not only to CHF but also to its predisposing conditions, such as LV hypertrophy and adverse remodeling after MI (215) (Fig. 9). H<sub>2</sub>S is a ROS scavenger and an antioxidant activator (167). The application of H<sub>2</sub>S donors may, therefore, be potentially useful for treating heart failure (HF) Table 3.

1. **Endogenous H<sub>2</sub>S production in CHF.** MI is one of the leading causes for HF. Wang *et al.* found that the plasma H<sub>2</sub>S level decreased from 65 to 53  $\mu$ M in an MI-induced CHF model (259). In addition, endogenous H<sub>2</sub>S synthesis in the heart was also found to be lowered in hearts of arteriovenous fistula (AVF)-induced CHF model (167, 217) and adriamycin (ADR)-induced cardiomyopathy model (236). This was further supported by transgenic mice overexpressing CSE, as excessive H<sub>2</sub>S production protects against HF injuries in both the permanent LCA ligation model and the LCA I/R model (35).



**FIG. 9. Effect of H<sub>2</sub>S on the pathogenesis of heart failure.** H<sub>2</sub>S was shown to protect against chronic heart failure (CHF) by scavenging ROS, inhibiting redox signaling, and attenuating oxidative damages. The action of H<sub>2</sub>S in reducing oxidative damage is partially mediated by the activation of superoxide dismutase (SOD) and glutathione peroxidase (GPx).

TABLE 3. PROTECTIVE EFFECTS OF HYDROGEN SULFIDE AGAINST HEART FAILURE

Experimental model	Species	H <sub>2</sub> S treatment	Results	Conclusions	Proposed mechanism(s)	Ref.
Permanent ligation of the LCA	CSE overexpression transgenic mice (MHC-CGL-Tg <sup>+</sup> ) vs. C57BL/6J mice	NA	Transgenic mice displayed: 68% ↑ in survival rate smaller ↑ in LVEDD, LVESD and heart to body weight ratio Transgenic mice displayed: 38% ↓ in infarct area smaller ↑ in LVEDD, LVESD, and heart to body weight ratio better LV ejection fraction 24 h reperfusion: 14% ↓ in infarct area/area at risk 20% ↓ in infarct area/LV 4 weeks reperfusion: 25% ↓ in infarct area/LV No change in LVEDD, LVESD, heart/body weight ratio, LV ejection fraction, or heart rate	CSE overexpression reduced LV dilatation and cardiac hypertrophy ↑ production of H <sub>2</sub> S during reperfusion has positive impact on LV structure and function Single administration of H <sub>2</sub> S at infarct size, but not sufficient to improve LV function at 4 weeks	Transgenic mice hearts expressed: ↑Nrf2 and NRF-1 ↑Akt ↑ nuclear localization of Nrf2 and NRF-1 ↑Akt phosphorylation in heart at serine residue 473 Attenuation of oxidative stress ↑ mitochondrial respiration and ATP synthesis, but no effect on mitochondrial biogenesis	(35)
60 min of LCA occlusion followed by 4 weeks of reperfusion	C57BL/6J mice	Single bolus of Na <sub>2</sub> S at reperfusion (100 µg/kg, i.c)	Na <sub>2</sub> S treatment: 25% ↓ in infarct area, ↓ in LV dilatation and cardiac hypertrophy improved cardiac function H <sub>2</sub> S treatment: ↓ heart weight ↓ collagen, ↓ fibrosis ↓ caspase-3 and apoptosis ↓ nitrotyrosine formation ↓ MMP-9 and MMP-2 activation ↑ TIMP-4, ↓ TIMP-1 and TIMP-3 ↑ β1-integrin, ↓ ADAM-12	H <sub>2</sub> S during first 7 days of reperfusion is critical for sustained improvements in LV structure and function H <sub>2</sub> S ↓ oxidative and proteolytic stresses improved cardiac histology by ↓ fibrosis and apoptosis	↓ oxidative and nitrosative stresses Reversed altered expression of MMPs, TIMPs, β1 and ADAM-12	(167)
Arteriovenous fistula (AVF)—volume overload	C57BL/6J mice	NaHS; 30 µM in drinking water	NaHS treatment: ↓ in LV chamber diameters restored hemodynamics parameters of heart—EF, EDP, ESP, dP/dt max and SV ↑ expression of MMP-2, CD31 and VEGF ↓ expression of MMP-9, endostatin, angiotensin, TIMP-3	H <sub>2</sub> S ↓ dilatation of heart ↑ LV functional status promote angiogenic inhibit antiangiogenic factors	↑ MMP-2 activation to promote VEGF synthesis and angiogenesis ↓ MMP-9, TIMP-3 levels and antiangiogenic factors	(77)
Aortic banding (AB)—pressure overload	C57BL/6J mice	NaHS; 30 µM in drinking water	H <sub>2</sub> S treatment: ↑ survival rate by 15% ↑ LVSP ↓ LVEDP ↑ LV ± dp/dt ↓ lung/body weight ratio ↓ fibrosis area/total LV area ↑ CSE, Bcl-2 expression ↓ Bax expression ↓ mitochondrial:cytoplasm cytochrome C and caspase-3 activation	H <sub>2</sub> S improve cardiac functions ↓ pulmonary oedema ↓ fibrosis ↓ cardiac apoptosis	↓ leakage of cytochrome c protein from mitochondrial to cytoplasm to improve mitochondrial derangements ↑ Bcl-2 protein and mRNA expression ↓ Bax and caspase-3 protein and mRNA expression	(259)
Ligation of left anterior descending coronary artery	Sprague-Dawley rats, male	NaHS (3.136 mg/kg/day)				

ADAM-12, disintegrin and metalloproteinase domain-containing protein 12; Bcl-2, B-cell lymphoma 2; LCA, left coronary artery; LVEDP, LV end diastolic pressure; NRF-1, nuclear respiratory factor 1; TIMP, tissue inhibitor of matrix metalloproteinases; VEGF, vascular endothelial growth factor; MMP, matrix metalloproteinases. NA, not applicable.

2. Effect of H<sub>2</sub>S on HF, hypertrophy, and cardiomyopathy. Cardiac hypertrophy as a result of sustained overload can lead to progression of HF. H<sub>2</sub>S has been reported as inhibiting hypertrophy (145) as well as its transition to HF (77). In rat primary cultures, NaHS (100  $\mu$ M) pretreatment prevented cardiomyocyte hypertrophy by lowering intracellular ROS, upregulating microRNA-133a, and suppressing microRNA-21 (145). Transgenic mice with cardiac-restricted overexpression of CSE displayed a clear protection against LV structural and functional impairment as assessed by echocardiography in response to ischemia-induced HF (35). The endogenous overexpression of CSE reduced LV dilation and cardiac hypertrophy. The exogenous application of NaHS (100  $\mu$ g/kg or 30  $\mu$ M in drinking water) also produced similar cardioprotective effects in different HF models (35, 167, 219, 259) and attenuated the development of ADR-induced cardiomyopathy (236) and hypertrophy in spontaneously hypertensive rats (SHR) (219).

The induction of antioxidative responses is an important mechanism for the protective effect of H<sub>2</sub>S. Na<sub>2</sub>S (100  $\mu$ g/kg) treatment significantly protected the heart against ischemia-induced lipid hydroperoxidation (LPO) in HF (35). Nrf2 is a key transcription factor involved in the regulation of antioxidant genes as well as mitochondrial biogenesis through the upregulation of nuclear respiratory factor 1 (NRF-1). Calvert *et al.* found that treatment with H<sub>2</sub>S for 7 days stimulated Akt and nuclear localization of NRF-1 and Nrf2. However, H<sub>2</sub>S therapy failed to increase mitochondrial biogenesis and did not significantly improve mitochondrial function 4 weeks after MI, although ATP synthesis was slightly improved. The slight improvements are more likely attributed to the ability of H<sub>2</sub>S to reduce oxidative stress, suggesting that in this model of HF, the antioxidant effects of H<sub>2</sub>S may play a more prominent role in mediating its cardioprotective actions (35). The protective effects of H<sub>2</sub>S in the cardiomyopathy model were also attributed to its antioxidant effects (236). NaHS (100  $\mu$ g/kg) treatment inhibited LPO and increased superoxide dismutase (SOD) and GSH peroxidase activities. Therefore, treatment with NaHS stimulates the activity of antioxidant enzymes. The protective effects against ischemia-induced injury are also mediated by the antiapoptotic effects of H<sub>2</sub>S (259). In an ischemia-induced HF model, NaHS reduced the number of apoptotic cells through promoting the expression of antiapoptotic factor Bcl-2 while suppressing the expressions of proapoptotic factors Bax and caspase-3. NaHS (3.136 mg/kg/day) treatment also significantly reduced the release of cytochrome c from the mitochondria and, therefore, protected the heart against ischemic injury during HF (259).

H<sub>2</sub>S may also protect against HF *via* promoting angiogenesis. One group of researchers reported that NaHS (30  $\mu$ M in drinking water) reversed the alteration of various matrix metalloproteinases and tissue inhibitor of matrix metalloproteinases (TIMP) in response to cardiac insults, and these factors resulted in enhanced angiogenesis in H<sub>2</sub>S-treated animals (77, 167). However, it should be taken into consideration that H<sub>2</sub>S is known to be very volatile, has a very short half life in a solution when exposed to air, and may escape readily within minutes. Furthermore, NaHS at 30  $\mu$ M produces strong irritating odor, and its dissociation ions (HS<sup>-</sup> or S<sup>2-</sup>) may change the taste of water. The amount of water intake by H<sub>2</sub>S-treated animals may differ from those in other groups.

Gap junction channels are specialized intercellular connections between the cytoplasm of two cells, controlling the passage of molecules and ions. A gap junction channel is made up of two connexons, each composed of six connexin proteins. The administration of NaHS (50  $\mu$ mol/kg/day) and H<sub>2</sub>S-donating S-diclofenac (25 and 50  $\mu$ mol/kg/day) enhanced mRNA and protein expressions of connexin 43 and 45 in doxorubicin-induced cardiomyopathy (304), indicating that H<sub>2</sub>S may also protect the heart against HF *via* inhibiting pathogenic gap junction remodeling in the heart.

## V. Physiological Function of H<sub>2</sub>S in Vascular Tissue

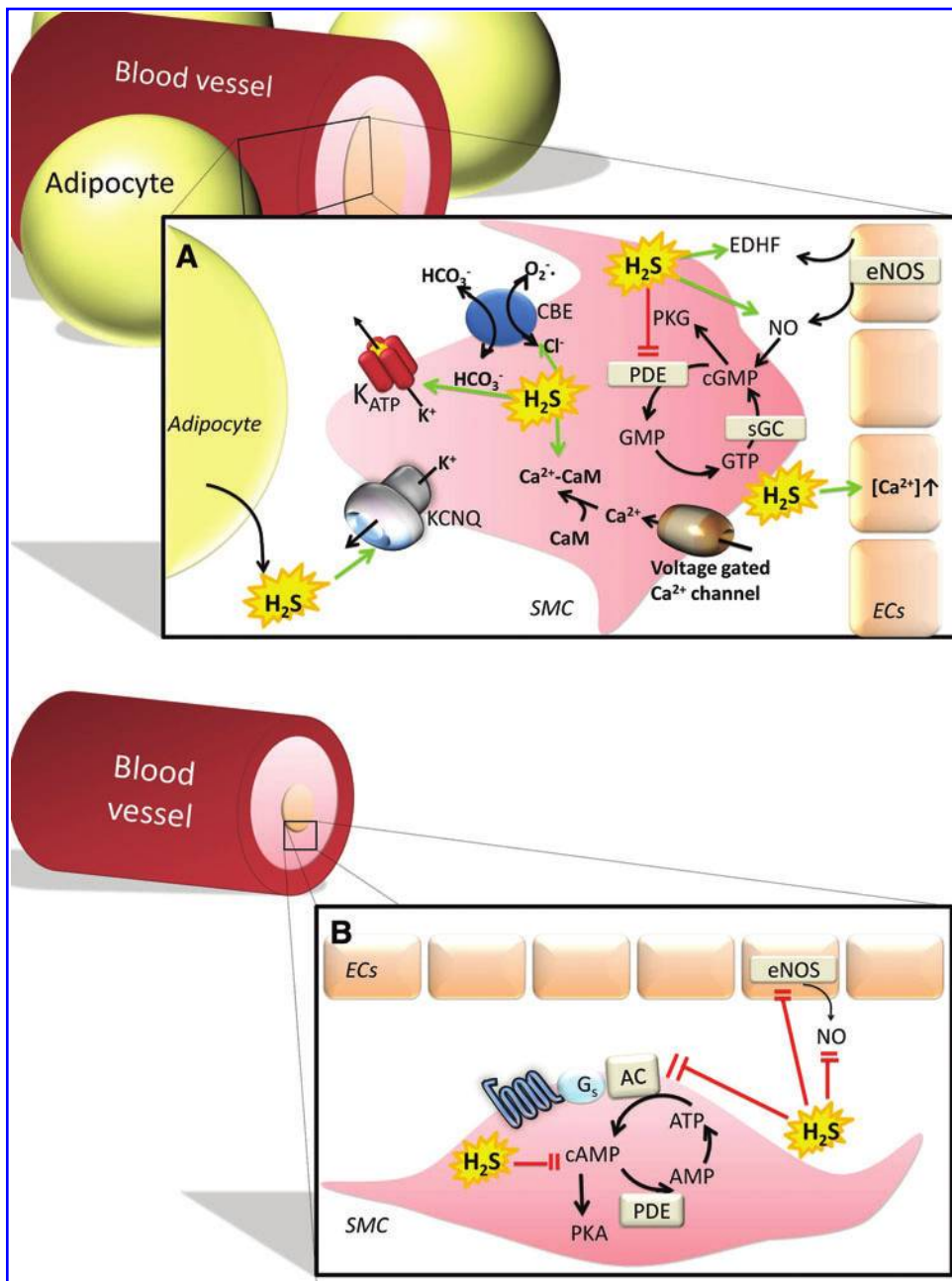
It was first demonstrated by Hosoki *et al.* in 1997 that arteries and veins express CSE and generate H<sub>2</sub>S (91). NaHS at concentrations above 100  $\mu$ M may induce the relaxation of precontracted isolated rat artery (3, 91, 309). Furthermore, perfusion of the rat mesenteric arterial bed with the H<sub>2</sub>S precursor, 1 mM cysteine, increased endogenous release of H<sub>2</sub>S and relaxed the arterial bed (47). In contrast, NaHS at concentrations below 100  $\mu$ M may induce further contraction of precontracted isolated vessels (3, 141, 148). Figure 10A and B show the mechanisms accounting for the relaxant and constrictive effects seen with H<sub>2</sub>S, respectively. The response of blood vessels to H<sub>2</sub>S varies according to the type of vessel: large conductance vessels *versus* small resistance vessels; systemic *versus* pulmonary; the condition of endothelium (intact *vs.* denuded); the precontraction agonist used (*e.g.*, potassium chloride *vs.* phenylephrine); the method of H<sub>2</sub>S administration (single *vs.* cumulative application); and the duration, concentration, and rate of change in the concentration of the H<sub>2</sub>S administered. The administration of slow-release H<sub>2</sub>S donors probably more closely resembles endogenous H<sub>2</sub>S release. The concentration of H<sub>2</sub>S in blood has been reported to be altered in several pathological states, including patients suffering from CAD (103), hypertension (43), and diabetes (100). Although these changes in H<sub>2</sub>S levels reflect changes in the amounts of stored sulfide (due to the methods used to measure blood concentrations), the H<sub>2</sub>S concentrations of stored sulfide probably reflect the status of H<sub>2</sub>S activity. Whether such changes in the H<sub>2</sub>S level are the causes or consequences of these diseases warrants further investigations.

### A. H<sub>2</sub>S-induced vasorelaxation

As just mentioned, Hosoki *et al.* first found that H<sub>2</sub>S can relax portal vein and thoracic aorta (91). The vasodilatory effect of H<sub>2</sub>S was later observed in several types of arteries, including mesenteric (47), pulmonary (263), hepatic (63), and tail arteries (309). H<sub>2</sub>S-induced vasorelaxation is mainly brought about by the opening of the K<sub>ATP</sub> channels (47, 121, 309) and partially mediated by endothelium-dependent mechanism(s) (309). Other signaling mechanisms may involve intracellular acidosis (127) and depletion of intracellular ATP levels in aortic rings (116, 240, 268) (Fig. 10A). Since vasorelaxant responses are only observed in *in vitro* organ bath studies at NaHS concentrations above 100  $\mu$ M, the significance of this effect and its mechanisms are yet to be fully understood *in vivo*.

#### 1. Mechanisms of H<sub>2</sub>S-induced vasorelaxation

*a. Opening of K<sub>ATP</sub> channels.* The K<sub>ATP</sub> channel is an inward rectifying channel composed of two types of subunits.



**FIG. 10. Mechanisms for H<sub>2</sub>S-induced vascular responses.** (A) H<sub>2</sub>S may induce vasorelaxant responses by the activation of the K<sub>ATP</sub> channels and Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> (CBE) channels. H<sub>2</sub>S has been implicated as an adipocyte-derived relaxing factor that causes the relaxation of smooth muscle cells (SMCs) by acting on K<sub>v</sub>7.x (KCNQ) channels. Other mechanisms of H<sub>2</sub>S-induced relaxation include binding of the Ca<sup>2+</sup>-calmodulin complex; interaction with NO and/or endothelium-derived hyperpolarizing factor (EDHF); inhibition of phosphodiesterase (PDE); and elevation of intracellular Ca<sup>2+</sup> in endothelial cells (ECs). (B) H<sub>2</sub>S may induce vasoconstrictive responses by the inhibition of AC, eNOS, and scavenging of NO (To see this illustration in color, the reader is referred to the web version of this article at [www.liebertonline.com/ars](http://www.liebertonline.com/ars)). eNOS, endothelial NOS.

The smaller pore-forming subunits are the inwardly rectifying potassium channel subunits (Kir6.1 or Kir6.2), while the larger regulatory subunits are the sulfonylurea receptors (SUR1, SUR2A, and SUR2B). The K<sub>ATP</sub> channel isoforms come from differing combinations of subunits. For example, Kir6.1/SUR2B is found in vascular smooth muscle cells (VSMCs), whereas Kir6.2/SUR2B is found in nonvascular SMCs (64). The K<sub>ATP</sub> channels exist in the sarcolemmal, mitochondrial, and nuclear membranes of various tissues (316), and are characterized by their inhibition by micromolar concentrations of intracellular ATP (7). Hence, an increase in intracellular ATP closes these channels, leading to cell depolarization. The K<sub>ATP</sub> channels participate in the control of membrane potentials and the regulation of cellular activities such as energy metabolism, apoptosis, and gene expression.

Zhao *et al.* first reported that glibenclamide, a K<sub>ATP</sub> channel antagonist, significantly attenuated the vasodilatory effect of H<sub>2</sub>S (600  $\mu$ M), whereas pinacidil, a K<sub>ATP</sub> channel opener, mimicked the effect of H<sub>2</sub>S (300  $\mu$ M) in a concentration-dependent manner (309). Using the patch clamp technique in single VSMC isolated from the aorta, H<sub>2</sub>S induced K<sup>+</sup> currents and led to membrane hyperpolarization, inhibitable by glibenclamide (309). In another study, H<sub>2</sub>S elevated whole-cell K<sub>ATP</sub> currents and membrane hyperpolarization in the VSMCs of rat mesenteric arteries with an EC<sub>50</sub> value of  $116 \pm 8.3 \mu$ M (245). Moreover, whole-cell K<sub>ATP</sub> currents could be attenuated by the inhibition of endogenous H<sub>2</sub>S production, indicative of basal K<sub>ATP</sub> stimulation by endogenous H<sub>2</sub>S (245). In addition, H<sub>2</sub>S (200  $\mu$ M) enhanced single-channel activity of K<sub>ATP</sub> channels by increasing the open probability without any effect on single-channel conductance (245).

When the K<sub>ATP</sub> subunits (*i.e.*, rvKir6.1 and rvSUR1) were heterologously expressed in HEK-293 cells and studied using whole-cell patch-clamp technique (102), it was observed that although both rvKir6.1 alone and rvKir6.1/rvSUR1 co-expressed channels conduct K<sub>ATP</sub> channel currents, H<sub>2</sub>S only elevated K<sub>ATP</sub> currents generated by the latter (102). This strongly indicates that H<sub>2</sub>S acts on SUR1 but not Kir6.1 subunit, of the K<sub>ATP</sub> channel. It was also reported that expressions of SUR2B were higher in the aorta than in the pulmonary artery, suggesting that the greater K<sub>ATP</sub> channel density in the aorta may underlie its greater extent of relaxation than that in pulmonary arteries (238).

To localize the target of H<sub>2</sub>S action on the K<sub>ATP</sub> channel, the channel was chemically modified by N-ethylmaleimide (NEM, a cysteine-specific sulfhydryl alkylating agent) or chloramine T (CLT, an agent for oxidation of sulfhydryl groups cysteine and methionine) (102). Both drugs could abolish the stimulatory effect of H<sub>2</sub>S on K<sub>ATP</sub> currents completely. While NEM did not alter rvKir6.1/rvSUR1 currents, CLT inhibited rvKir6.1/rvSUR1 currents by ~45%, suggesting that its high concentration could have oxidized methionine residues besides the intended cysteine sulfhydryl groups. Using site-directed mutagenesis, extracellular cysteine residues on rvSUR1 subunit were replaced with structurally similar serine residues. Single-site mutants on cysteine-6 and cysteine-26, but not cysteine-1051 and cysteine-1057, completely abolished the H<sub>2</sub>S effect on rvKir6.1/rvSUR1. Hence, extracellular cysteine residues cysteine-6 and cysteine-26 of rvSUR1 subunit are most likely the targets of H<sub>2</sub>S actions (102). In the most recent report published by the same group, the researchers found evidence that H<sub>2</sub>S covalently sulphydrates cysteine-43 on Kir 6.1 to bring about the hyperpolarization which underlies H<sub>2</sub>S-induced vasorelaxation (174). Furthermore, H<sub>2</sub>S has been shown as enhancing the binding of phosphatidylinositol-4,5-bisphosphate to Kir 6.1 to elicit K<sub>ATP</sub> activation. Conversely, H<sub>2</sub>S attenuated the binding of the inhibitor, ATP, to Kir6.1 (174).

Despite numerous reports and the widely accepted notion that the K<sub>ATP</sub> channel is the main underlying mediator of most H<sub>2</sub>S-induced vascular responses, H<sub>2</sub>S-induced vascular relaxation is often only partially inhibited by glibenclamide, and, in some cases, no inhibition can be achieved (42, 116, 268). Cheang *et al.* recently reported that the K<sub>ATP</sub> channels may not be the underlying mechanism of H<sub>2</sub>S effects in rat coronary arteries, suggesting that the effect of H<sub>2</sub>S on K<sub>ATP</sub> channels could be tissue specific (42). There is also no evidence that supports a role for the K<sub>ATP</sub> channels in the vasoconstrictive action of H<sub>2</sub>S (3, 141, 268, 274).

*b. Endothelium-dependent mechanism.* The endothelium is the production site of NO and perhaps other unidentified endothelium-derived relaxing factors in blood vessels. NO is a potent vasorelaxant, acting in the nanomolar range. Hosoki *et al.* first reported the synergistic effect between NO and H<sub>2</sub>S where NaHS was observed as inducing a significantly greater concentration-dependent relaxant response in the presence of an NO donor (sodium nitroprusside [SNP] or morpholin-sydnonimine). Interestingly, NaHS at a low concentration (30  $\mu$ M) that induces vasoconstriction by itself enhanced the NO-induced relaxant effect by up to 13-fold (91). Conversely, blocking endogenous NO production with L-NAME,

or physically removing the endothelium, attenuated H<sub>2</sub>S-induced relaxation (309). The co-administration of charybdotoxin and apamin to inhibit the classical pathway of endothelium-derived hyperpolarizing factor (EDHF) in endothelium intact vessels also attenuated H<sub>2</sub>S-induced relaxation (309). In line with these synergistic effects, the incubation of homogenized rat vascular tissues with SNP increased H<sub>2</sub>S production in a concentration-dependent manner (307, 309), an effect that could be abolished by a soluble guanylate cyclase inhibitor (1H-[1,2,4]oxadiazolo[4,3-a]quinoxalin-1-one). Similarly, the NO donor (diethylamine NONOate) also increased H<sub>2</sub>S production rate (307). Moreover, incubation of cultured VSMCs with another NO donor (S-nitroso-N-acetylpenicillamine [SNAP]) increased the transcription level of CSE (309). However, the relationship between NO and H<sub>2</sub>S is complicated. Mechanisms of H<sub>2</sub>S-induced vasoconstrictions section describes the antagonizing effects of these gasotransmitters.

*c. Acidification.* The level of pH<sub>i</sub> has been shown as regulating vascular tone. A change in pH<sub>i</sub> alters [Ca<sup>2+</sup>]<sub>i</sub> in VSMC, which may subsequently lead to changes in force generation *via* modulation of various ion channels and actin/myosin crossbridge activity in a concerted manner (279). In most vascular tissues, it is widely regarded that alkalization causes contraction, whereas acidification causes relaxation.

Lee *et al.* found that H<sub>2</sub>S may induce intracellular acidosis by stimulating Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchanger (CBE) activity (127). Intracellular acidosis can stimulate K<sub>ATP</sub> channels and, thus, cause vasodilation (227, 260); H<sub>2</sub>S may, therefore, induce vasodilation *via* the intracellular acidosis/K<sub>ATP</sub> pathway. In addition, angiotensin II (Ang II) has been reported as exhibiting a similar pH<sub>i</sub>-lowering effect in cat papillary muscles *via* the activation of cardiac CBE in a PKC-dependent manner (37). Thus, more studies are warranted to examine the involvement of PKC in the pH regulatory role of H<sub>2</sub>S.

*d. Metabolic inhibition.* H<sub>2</sub>S (320  $\mu$ M) significantly decreased ATP levels in vascular tissues 10 s after its administration, with an even greater reduction after 3 min. The extent of H<sub>2</sub>S-induced relaxation and the time course of H<sub>2</sub>S effects on ATP were similar to those induced by hydrogen cyanide (complex IV inhibitor) and 2,4-dinitrophenol (mitochondrial uncoupler) (116). The physiological significance of this finding warrants further investigations using lower H<sub>2</sub>S concentration. It was also reported that H<sub>2</sub>S-induced relaxation was more pronounced at a low ambient oxygen level, whereas vasoconstriction was enhanced by a high oxygen level (116, 117), probably due to the competition between sulfide and oxygen at the level of cytochrome c (116). Since sustained contraction of blood vessels requires ATP, it was thought that H<sub>2</sub>S may induce relaxation *via* metabolic inhibition in vascular tissues, thus reducing ATP and oxygen consumption (116), in line with its cardioprotective effect in the heart (61, 188). Intriguingly, it should be noted that NaHS below 20  $\mu$ M has shown a stimulatory effect on oxygen consumption (80, 299). As such, the role of H<sub>2</sub>S under physiological conditions is still unclear and worthy of in-depth research.

*e. Opening of voltage-dependent K<sup>+</sup> channels.* The involvement of voltage-dependent K<sup>+</sup> (K<sub>v</sub>) channels in the vascular function of H<sub>2</sub>S is somewhat controversial. Zhao *et al.* first reported that 4-aminopyridine (4-AP, a specific K<sub>v</sub> inhibitor)

failed to affect H<sub>2</sub>S-induced vasorelaxation in rat aorta (309). A more recent study using denuded rat coronary artery, however, reported that 4-AP reduced NaHS-induced vasorelaxation when the tissue was precontracted with U46619 (42). Results obtained from these studies suggest that the action of H<sub>2</sub>S varies among the type of vascular tissue being examined. While H<sub>2</sub>S activates K<sub>ATP</sub> channels in rat mesenteric arteries (47) and the aorta (309), it appears to activate the K<sub>v</sub> channels that induce hyperpolarization in the rat coronary artery (42).

In another study, H<sub>2</sub>S was implicated as an adipocyte-derived relaxing factor (ADRF) *via* its action on the KCNQ channels (210). The KCNQ channels are a type of K<sub>v</sub> channel residing in many types of vascular tissues. Perivascular adipose tissue has been reported to secrete ADRF in a paracrine fashion that reduces vascular tone (60, 78, 152, 251). The presence of perivascular fat in mesenteric arteries [(+)fat] significantly attenuated 5-HT-induced contraction when compared with fat-free [(-)fat] vessels. This anticontractile effect of perivascular fat can be reversed by 4-AP (K<sub>v</sub> channel blocker) or XE991 (KCNQ channel blocker), but not glibenclamide, suggesting that the K<sub>v</sub> channels, not the K<sub>ATP</sub> channels, underlie the differential observations in fat-mediated contractile responses. The inhibition of endogenous H<sub>2</sub>S production by incubating aortic rings with CSE inhibitors,  $\beta$ -cyano-L-Alanine (BCA) or PAG, has no effect on (-)fat vessels, but significantly reverses the anticontractile effect of (+)fat vessels. Hence, H<sub>2</sub>S could be an endogenous ADRF released by perivascular adipose tissues acting on the vascular tissues to exert its relaxant effects. More direct evidence was provided when (-)fat aortic tissue was preincubated with XE991, which prevented NaHS evoked concentration-dependent vasorelaxations, indicating that KCNQ channels play a role in mediating control of vascular tone by H<sub>2</sub>S (210).

*f. Increase in cyclic guanosine monophosphate levels.* Cyclic guanosine monophosphate (cGMP)/PKG facilitates the dephosphorylation of myosin light chain (MLC), preventing the association of myosin with actin. Bucci *et al.* recently reported that H<sub>2</sub>S elevates cGMP levels by inhibiting phosphodiesterase (PDE) activity (24). The incubation of cultured rat aortic SMC with exogenous NaHS (10–200  $\mu$ M) increased cGMP levels in a concentration-dependent manner. More importantly, CSE overexpression (by CSE cDNA transfection) and CSE gene silencing (using small interfering RNA [siRNA] approach) greatly enhanced and attenuated cGMP levels, respectively, suggesting an endogenous role of H<sub>2</sub>S in regulating cGMP levels (24). There is a need, therefore, to re-evaluate the conventional view that H<sub>2</sub>S has no effect on cGMP levels (245, 308, 309), and to clarify the inconsistency in observations.

*g. Modulation of Ca<sup>2+</sup> signaling.* Both calcium-free bath solution and nifedipine (a Ca<sup>2+</sup> channel blocker) attenuated H<sub>2</sub>S-induced vasorelaxation of rat aortic tissue, implying that it may partly be dependent on Ca<sup>2+</sup> influx (308). However, in human saphenous vein ECs, NaHS significantly increases [Ca<sup>2+</sup>]<sub>i</sub> with maximal responses observed at 200–500  $\mu$ M (11). This concentration range, however, is probably beyond a physiological level. Nevertheless, the chelation of extracellular Ca<sup>2+</sup> with EDTA failed to block this [Ca<sup>2+</sup>]<sub>i</sub> elevation, revealing that the mobilization of Ca<sup>2+</sup> came from intracellular sources. In fact, in the absence of extracellular Ca<sup>2+</sup>, depletion

of Ca<sup>2+</sup> stores by exposing cells to either ATP (to activate P2Y receptors and generate IP<sub>3</sub>) or 4-chloro-3-ethylphenol (4-CEP, to activate ryanodine receptors [RyRs]) diminished H<sub>2</sub>S-induced [Ca<sup>2+</sup>]<sub>i</sub> rise. Store depletion with SERCA inhibitors, thapsigargin, or cyclopiazonic acid produced similar effects. H<sub>2</sub>S probably mobilizes the same pool of intracellular Ca<sup>2+</sup> store as IP<sub>3</sub> receptors (IP<sub>3</sub>Rs) or RyRs (11). The regulatory effect of H<sub>2</sub>S on [Ca<sup>2+</sup>]<sub>i</sub> is not mediated by NCX, Ca<sup>2+</sup>-ATPase, or cyclic ADP ribose (cADPR), as the blockade of NCX with its two structurally distinct inhibitors, SEA0400 or bopindril, Ca<sup>2+</sup> ATPase with carboxyeosin or cADPR with 8-Br-cADPR (a cADPR antagonist) or nicotinamide (an ADP ribosyl cyclase inhibitor), failed to attenuate the effect of NaHS (11). In some but not all the cells, re-exposure to extracellular Ca<sup>2+</sup> after the addition and removal of H<sub>2</sub>S activated capacitative Ca<sup>2+</sup> entry (11). Taken together, these data suggest that H<sub>2</sub>S modulates endothelial [Ca<sup>2+</sup>]<sub>i</sub> *via* multiple mechanisms.

*h. AA cascade.* AA is the precursor of eicosanoids such as prostaglandins and leukotrienes, present in abundance in muscles. AA implicated in cell signaling is derived by the action of a phosphatidylcholine-specific cytosolic PLA<sub>2</sub> (cPLA<sub>2</sub>). NaHS (100  $\mu$ M–1 mM)-induced relaxation in mesenteric artery beds could be greatly attenuated by 4-(4-Octadecylphenyl)-4-oxobutenoic acid (OBAA, PLA<sub>2</sub> inhibitor) (53). Moreover, NaHS (1 mM) induced a greater extent of cPLA<sub>2</sub> translocation from the cytoplasm to the nucleus, indicative of increased activation of AA synthesis in response to H<sub>2</sub>S (53). These authors further demonstrated that H<sub>2</sub>S-induced relaxation could also be significantly reduced in the presence of proadifen (PRD, CYP inhibitor), suggestive of the involvement of AA metabolites in mediating H<sub>2</sub>S-induced vasorelaxation (53). It should be noted, however, that the concentration of NaHS used in this study was way above the physiological range and, hence, probably lacks relevance to *in vivo* situations. Furthermore, several other studies have reported that prostaglandins and COX have little role in the vascular effects of H<sub>2</sub>S (42, 116, 309), hence suggesting probable species or cell specificity for such effects. More studies are needed to further understand the interaction of H<sub>2</sub>S with AA and its metabolites.

*i. Release of EDHF.* EDHF causes vasodilatation by hyperpolarizing VSMCs, particularly in small-resistance arteries. EDHF-induced vasorelaxation is mainly generated by the activation of small and intermediate conductance calcium-activated potassium channels (SK<sub>Ca</sub>/K<sub>Ca</sub>2.3 and IK<sub>Ca</sub>/K<sub>Ca</sub>3.1). H<sub>2</sub>S has been proposed as exhibiting properties that resemble EDHF in several reports (53, 150, 174, 288). As discussed earlier, H<sub>2</sub>S-induced vascular responses are endothelium dependent, and it was suggested that its relaxant responses in small-resistance arteries such as mesenteric arteries or arterioles may be greater than in large conduit arteries such as the aorta (47, 174). Furthermore, both Zhao *et al.* and Bianca *et al.* reported that coapplication of charybdotoxin (IK<sub>Ca</sub> blocker) and apamin (SK<sub>Ca</sub> blocker) reduced the extent of the H<sub>2</sub>S-induced vasorelaxation (53, 308), while charybdotoxin and iberiotoxin (BK<sub>Ca</sub> blocker) alone had no such effect (309). In addition, coapplication of charybdotoxin and apamin significantly attenuated H<sub>2</sub>S-induced vasorelaxation in the endothelium intact mesenteric artery bed, but not that in the endothelium-denuded counterpart (47). These findings suggest that H<sub>2</sub>S may act by releasing EDHF from the endothelium.

The strongest evidence that supports the role of H<sub>2</sub>S as an EDHF probably comes from the recent findings of Mustafa *et al.* (174). They found that coapplication of the K<sub>ATP</sub> blocker (glibenclamide) with IK<sub>Ca</sub>/SK<sub>Ca</sub> blockers (charybdotoxin/apamin) abolished all H<sub>2</sub>S-mediated vasorelaxation in rat mesenteric arteries. By measuring membrane potentials of primary mouse aortic ECs, they found that CSE-knockout mice failed to respond to Ach-induced hyperpolarization, unlike wild-type mice. The coapplication of charybdotoxin/apamin completely attenuated hyperpolarization in wild-type mice, suggesting that IK<sub>Ca</sub>/SK<sub>Ca</sub> channels underlie H<sub>2</sub>S effects (174). In separate experiments, the researchers further proved that IK<sub>Ca</sub>, but not K<sub>ATP</sub> and BK<sub>Ca</sub> channels, mediate H<sub>2</sub>S-induced hyperpolarization in cultured human aortic ECs (174). Collectively, these data suggest that H<sub>2</sub>S may play an important role in mediating the vascular responses of small and intermediate resistance vessels.

### B. H<sub>2</sub>S-induced vasoconstriction

NaHS induces concentration-dependent vasoconstriction at concentrations of 10–100  $\mu$ M, lower than those that cause vasorelaxation (3, 121, 141) (Fig. 10B). Unlike H<sub>2</sub>S-induced vasorelaxation, the K<sub>ATP</sub> channels are unlikely to be involved in H<sub>2</sub>S-induced vasoconstriction (121, 141).

#### 1. Mechanisms of H<sub>2</sub>S-induced vasoconstrictions

*a. NO-H<sub>2</sub>S interaction.* While NO and H<sub>2</sub>S act synergistically in vasorelaxation (previous section on Endothelium-Dependent Mechanism), H<sub>2</sub>S-induced vasoconstriction is more likely a result of NO depletion. Zhao *et al.* observed that H<sub>2</sub>S (60  $\mu$ M) pretreatment inhibited SNP-induced vasorelaxation with a rightward shift of the concentration-response curve and elevated the IC<sub>50</sub> value (308). Ali *et al.* showed that H<sub>2</sub>S, at relatively low concentrations of 10–100  $\mu$ M, had a vasoconstrictive effect in endothelium-intact vessels but not in endothelium-denuded vessels (3), suggesting probable quenching of endogenous NO by H<sub>2</sub>S. Moreover, NaHS (50–200  $\mu$ M) reversed Ach- or histamine-induced relaxations (both are endothelium dependent) but not ISO-induced relaxation (endothelium independent), showing that H<sub>2</sub>S-induced constriction is endothelium dependent (3). Furthermore, mixing NO donors (SNP, SIN-1, or SNAP) with NaHS (100  $\mu$ M) reduced the extent of vasorelaxation compared with the relaxation with NO donors alone, further indicating the inactivation of NO by H<sub>2</sub>S (3). The authors ascribed these observations to the formation of a nitrosothiol compound, as the administration of copper sulfate, which converts nitrosothiol to nitrite and nitrates, abolished NaHS (50–200  $\mu$ M)-induced vasoconstriction (3). This is in line with their earlier report that utilized a combination of biochemical assays for NO<sub>2</sub><sup>−</sup>, NO amperometric detection and electron paramagnetic resonance that determines nitrosothiol formation between NO and H<sub>2</sub>S (274). Since the identity and physiological significance of the nitrosothiol compound is unknown, and the methods employed are indirect, more studies such as mass spectrometry or high-performance liquid chromatography are warranted to confirm this interaction.

*In vivo*, a bolus injection of SNP (16.5 nmol/kg) or NaHS (5  $\mu$ M/kg) alone resulted in a 35 mm Hg or a <10 mm Hg drop in the MAP of anesthetized rats, respectively, but an injection of their mixture resulted in a loss of MAP reducing

ability (3). Interestingly, a slow infusion of NaHS (10  $\mu$ M/kg/min) significantly increased MAP, an effect that could be reversed by the infusion of L-NAME (3). This implies that a low concentration of H<sub>2</sub>S may scavenge endogenously produced NO. The sustained effect of NaHS infusion on MAP is inconsistent with the transience or absence of vasodepression reported by others (refer to Effect of H<sub>2</sub>S on BP of normotensive animals section), probably due to the different route of H<sub>2</sub>S administration.

*b. Downregulation of endothelial NOS.* H<sub>2</sub>S can exert inhibitory effects on endothelial NOS (eNOS) activity and, thus, reduce NO synthesis in the endothelium (121). Kubo *et al.* first observed that NaHS (30–3000  $\mu$ M) concentration dependently inhibited the activity of recombinant eNOS (121). Geng *et al.* further demonstrated that H<sub>2</sub>S could inhibit NO production, downregulate eNOS activity, reduce eNOS transcript abundance, and decrease L-[<sup>3</sup>H] Arg uptake in aortic tissues (74). Using human umbilical vein ECs (HUVECs), NaHS (50  $\mu$ M) decreased the catalytic efficiency of eNOS but not its affinity for L-arginine (eNOS substrate). NaHS (14  $\mu$ M/kg, i.p.) administration into rats resulted in a decreased plasma level of NO metabolites, aortic eNOS activity, and L-arginine uptake (74), suggesting the downregulation of L-Arginine/eNOS/NO pathway *in vivo*. iNOS and nNOS, on the other hand, are not targets of H<sub>2</sub>S action (74). However, the significance of these results warrants further investigation, as NaHS above 100  $\mu$ M probably does not bespeak physiological relevance.

*c. Decrease in cAMP level.* H<sub>2</sub>S-induced vasoconstriction was not completely abolished in the presence of NOS inhibitor, L-NAME, or removal of endothelium, suggesting the involvement of an NO-independent mechanism. One possibility is the downregulation of the cAMP level in VSMCs (141). A decrease in cAMP brought about by H<sub>2</sub>S induces vasoconstriction by upregulating the activation of MLC kinase, the enzyme responsible for triggering the interaction between actin and myosin.

Lim *et al.* reported that NaHS (at concentrations between 10 and 100  $\mu$ M) could inhibit relaxations induced by isoprenaline, salbutamol (two  $\beta$ -adrenoceptor agonists), and forskolin (an AC activator), implying that H<sub>2</sub>S at lower concentrations can counteract the relaxant effects brought about by vasodilators *via* vasoconstrictive effects (141). More importantly, NaHS (5–100  $\mu$ M) administration significantly inhibited forskolin-induced cAMP accumulation in rat aortic SMC (141). Since the low concentration of NaHS utilized is near physiological, this could be a potential underlying signaling mechanism *in vivo*.

*d. Reactive oxygen species.* O<sub>2</sub><sup>−</sup> is implicated in the etiology of a variety of vascular diseases, such as atherosclerosis (294), diabetic angiopathy (84), and hypertension (13). It generally induces vasoconstriction, probably due to the quenching of NO. The incubation of human VSMCs with NaHS was found to concentration dependently inhibit O<sub>2</sub><sup>−</sup> formation, nicotinamide adenine dinucleotide phosphate oxidase expression, and Rac<sub>1</sub> activity (175). Importantly, O<sub>2</sub><sup>−</sup> was found to mediate H<sub>2</sub>S-induced vascular responses. Tiron (an SOD mimetic) significantly enhanced NaHS-induced vasorelaxation, but greatly inhibited NaHS-induced vasoconstriction (318). The transport of O<sub>2</sub><sup>−</sup> across the cellular plasma membrane *via* anion exchanger (CBE) was found to be upregulated in the presence of H<sub>2</sub>S (127, 318), and the rise in

extracellular  $O_2^-$  depletes NO, resulting in a reduction in relaxation response and an enhancement of constrictive response (318). ONOO<sup>-</sup> derived from  $O_2^-$  and NO interaction might be removed by  $H_2S$  acting as an ONOO<sup>-</sup> scavenger as reported in human neuroblastoma SH-SY5Y cells (272). As mentioned in the previous Acidification section,  $H_2S$ -induced stimulation of CBE at a concentration  $>100 \mu M$  results in intracellular acidosis and a relaxant response, masking this vasoconstrictive effect. Nevertheless, the inhibition of CBE resulted in a stronger degree of  $H_2S$ -induced vasorelaxation (148), evident of the NO depletion phenomenon.

## VI. Other Roles of $H_2S$ in Blood Vessels

### A. Angiogenesis

Angiogenesis is a physiological process responsible for the growth of new blood vessels from existing ones. It is a complex biological process characterized by extracellular matrix remodeling and alterations in the behavior of ECs. Cell proliferation, adhesion, and migration, and assembly into capillary structures (development of vessel lumen) are all key events in blood vessel formation. The dysregulation of angiogenesis may result in tumor growth, psoriasis, arthritis, neurodegeneration, wound healing defects, and hair loss (190).

1. Effect of  $H_2S$  on angiogenesis. In blood vessels, ECs are both targets and sources of  $H_2S$  (288). Current evidence suggests that  $H_2S$  promotes angiogenesis and cell growth (Fig. 11), or apoptosis and cell death according to its con-

centration (28, 241, 256). For example, the expression of survivin (an inhibitor of apoptosis) was significantly upregulated at low concentrations of NaHS (1–10  $\mu M$ ), while its expression at a high concentration of 200  $\mu M$  NaHS was significantly attenuated (28), suggestive of its physiological importance.

a. *In vitro EC growth and migration.* In *in vitro* experiments with different types of cells such as RF/6A ECs (28) and HUVECs (190),  $H_2S$  has been shown as enhancing growth and proliferation. In addition, either exogenous or endogenous  $H_2S$  enhances cell migration in ECs using both the transwell migration assay and the *in vitro* scratch wound healing assay (28, 190). EC adhesion was observed only at low concentrations of NaHS (10 and 20  $\mu M$ ) but not at concentrations above 50  $\mu M$ , indicating that  $H_2S$  is proangiogenic only at lower concentrations (28).

The initial phase of angiogenesis involves the organization of ECs into a three-dimensional tube-like structure. The treatment of RF/6A ECs with NaHS (10–20  $\mu M$ ) or  $H_2S$  solution (10  $\mu M$ ) increased the tube length and branching points of a microvessel formed on Matrigel after 16 h of culture (28). Similarly, Papapetropoulos *et al.* also observed a 34% increase in the formation of a capillary-like structure of ECs cultured on reduced-growth factor Matrigel in response to  $H_2S$  (60  $\mu M$ ) stimulation (190).

Liu *et al.* found that cobalt chloride ( $CoCl_2$ ) (to mimic hypoxia) treatment of cultured VSMCs increased hypoxia inducible factor (HIF)-1 $\alpha$  and vascular endothelial growth

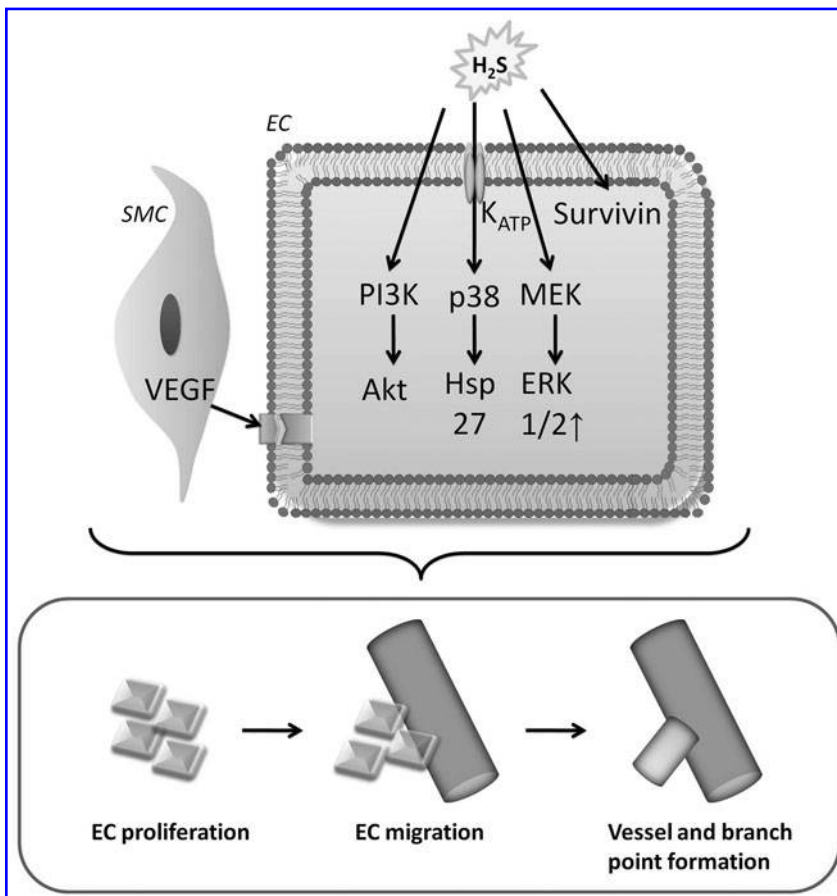


FIG. 11. Schematic illustration of possible signal transduction pathways that may underly  $H_2S$  induced endothelial cell (EC) proliferation, migration, and vessel/branch point formation.

factor (VEGF) mRNAs and proteins expression. Furthermore, the co-administration of NaHS (300  $\mu$ M) with CoCl<sub>2</sub> further enhanced the expressions of HIF-1 $\alpha$  and VEGF (146). In separate experiments, cultured ECs exposed to conditioned media from CoCl<sub>2</sub>-treated VSMC cultures showed decreased EC cell numbers than cultured ECs exposed to media from untreated VSMCs. However, ECs that had been exposed to media from VSMCs co-treated with NaHS (300  $\mu$ M) and CoCl<sub>2</sub> showed increased proliferation and migration (146). Hence, under hypoxic conditions, H<sub>2</sub>S-induced angiogenesis is probably HIF-1 $\alpha$ /VEGF dependent. The physiological relevance of this study, however, requires further in-depth study, because 300  $\mu$ M NaHS is nonphysiological.

*b. In vivo vascular network formation.* In an *in vivo* model utilizing Matrigel plug in mice, histological analysis revealed that injections of NaHS (10–50  $\mu$ mol/kg/day, i.p.) for 7 days significantly enhanced EC infiltration into the Matrigel embedded under the abdominal skin. A significantly increased level of hemoglobin content further demonstrated a higher degree of neovascularization. However, the proangiogenic effect of H<sub>2</sub>S *in vivo* was lost when the dose was 200  $\mu$ mol/kg/day NaHS (28).

The administration of H<sub>2</sub>S (0.24–240 pmol/egg) to fertilized White Leghorn chicken eggs dose dependently increased the total length of the vascular network in the chick chorio-allantoic membrane (190). Conversely, the administration of CSE inhibitors (PAG or BCA) dose dependently reduced the network length and vessel branching, effects that could be reversed by exogenously applied H<sub>2</sub>S (190). These results suggest that endogenous H<sub>2</sub>S is positively involved in the basal vascular network formation *in vivo*.

H<sub>2</sub>S also promotes vascular network formation in pathological situations. A hindlimb ischemic model was established in rats that had been subjected to unilateral femoral artery ligation. Chronic administration with NaHS (50  $\mu$ mol/kg/day) for 4 weeks promoted collateral vessel growth in ischemic hindlimbs, along with increased regional blood flow and increased capillary density, both indicative of microvessel growth. Furthermore, NaHS was also shown to enhance the growth of ECs and myocytes in ischemic muscles (255). Nevertheless, it is noteworthy that NaHS at a higher dose (200  $\mu$ mol/kg/day) failed to produce any significant proangiogenic effect (255). Taken together, these studies suggest that H<sub>2</sub>S may promote vascular network formation *in vivo* at a near physiological concentration.

*c. In vivo wound healing.* The effect of H<sub>2</sub>S on angiogenesis was also tested with burn wound assay (30% of the total body surface area) in rats. Daily topical administration of H<sub>2</sub>S (300  $\mu$ g/kg) enhanced wound healing along with a significant augmentation in re-epithelialization. When comparing CSE-knockout mice against wild-type controls, the sizes of the wound areas were consistently larger in CSE-knockout mice, suggesting that endogenous H<sub>2</sub>S generation contributes toward wound healing (190).

## 2. Mechanisms of the proangiogenic effects of H<sub>2</sub>S

*a. Effect of H<sub>2</sub>S on proangiogenic factors.* NaHS (1–200  $\mu$ M) has no effects on the levels of VEGF, basic fibroblast growth factor (bFGF), and angiopoietin-1 in the culture medium of RF/6A ECs (28). Subsequently, Papapetropoulos *et al.* also

showed that endogenous H<sub>2</sub>S synthesis or action does not affect bFGF-induced migration (190). However, Zhu *et al.* reported that NaHS (20–50  $\mu$ mol/kg/day) administration induced VEGF biosynthesis and protein expression in skeletal muscle cells, suggesting that H<sub>2</sub>S may exert its proangiogenic effect on neighboring ECs in a paracrine fashion (255). VEGF receptor 2 (VEGFR2) is the main receptor type involved in mediating the proangiogenic effect of VEGF, and these receptors are mainly localized in the vascular ECs (255). The downregulation of VEGFR2 during ischemia was reversed by NaHS (50  $\mu$ mol/kg/day) treatment, with specific phosphorylation at Tyr 996 of the receptor (255). This further strengthens the idea that H<sub>2</sub>S-induced VEGF in SMCs may act on ECs to promote angiogenesis.

### *b. Signaling mechanisms*

(1). *MAPK/ERK kinase/ERK pathway.* Cai *et al.* first noted that NaHS (10  $\mu$ M) increased Akt phosphorylation by about twofold, but not ERK and p38 activation (28). However, it was later discovered that H<sub>2</sub>S (60  $\mu$ M) elevates the phosphorylation of ERK1/2, p38 and Akt with differing kinetics: ERK1/2 activation was rapid and sustained; p38 activation was rapid and transient; and Akt activation was slow but sustained (190). In addition, MAPK/ERK kinase (MEK), a tyrosine/threonine kinase that phosphorylates and activates ERK, inhibitors (PD098059 or U0126) attenuated the H<sub>2</sub>S-elicited EC migration (190), suggesting that MEK lies upstream of ERK in mediating the angiogenic effect of H<sub>2</sub>S at a low concentration.

(2). *K<sub>ATP</sub>/p38/hsp27 pathway.* As just mentioned, ECs show a rapid and transient activation of p38 when exposed to a low concentration of H<sub>2</sub>S (60  $\mu$ M). p38 inhibitors (SB203580 or SB239063) were found to completely abolish the H<sub>2</sub>S-induced migration of EC (190), revealing an essential role of p38 in mediating H<sub>2</sub>S-induced angiogenic effects.

Hsp27, a modifier of actin cytoskeleton, is also implicated in the regulation of cell migration. Knockdown of hsp27 using the siRNA method significantly reduced H<sub>2</sub>S-induced migration of EC (190). Moreover, inhibition of p38 with SB203580 and SB239063 reduced hsp27 phosphorylation (190), suggesting that the p38 pathway is upstream of hsp27 in the H<sub>2</sub>S signaling cascade.

H<sub>2</sub>S-elicited EC migration was attenuated by a nonselective K<sub>ATP</sub> channel blocker (glibenclamide) or a selective mitochondrial-K<sub>ATP</sub> channel inhibitor (5-HD). Furthermore, glibenclamide attenuated H<sub>2</sub>S-induced p38 and hsp27 phosphorylation, while the K<sub>ATP</sub> opener (SG209) alone was capable of enhancing EC migration in a concentration-dependent manner, along with increased p38 phosphorylation (190). Taken together, these data suggest that the actions of H<sub>2</sub>S on K<sub>ATP</sub> channels are upstream of p38/hsp27.

(3). *PI3K/Akt pathway.* Cai *et al.* discovered that pretreatment with LY294002 (an inhibitor of PI3K) or transfection of dominant negative (kinase-inactive mutant Myr-Akt-K179M) Akt could greatly attenuate H<sub>2</sub>S (10–20  $\mu$ M)-induced wound-healing acceleration along with increments in tube length and branch points (28). The application of PI3K inhibitors (wortmannin or LY 294002) completely abolished NaHS (10–200  $\mu$ M)-induced increase in Akt phosphorylation in RF/6A ECs, suggesting that PI3K is upstream of Akt in the

underlying H<sub>2</sub>S signaling mechanism. In a hindlimb ischemia model, rats treated with NaHS (20–200  $\mu$ mol/kg/day) injections also showed significantly increased Akt phosphorylation compared with control animals (255).

In direct opposition, Papapetropoulos *et al.* reported that the angiogenic effect of H<sub>2</sub>S (60  $\mu$ M) is independent of the PI3K/Akt pathway, as the PI3K inhibitor LY-2924002 failed to affect the migratory rate of ECs in response to H<sub>2</sub>S (190). Therefore, the involvement of the PI3K/Akt pathway in the angiogenic effect of H<sub>2</sub>S is yet to be confirmed.

### B. H<sub>2</sub>S as an oxygen sensor

In mammals, hypoxia has been known to relax the systemic vessels while constricting the pulmonary vessels (270). Olson and colleagues found many similarities between H<sub>2</sub>S and hypoxia-induced pulmonary responses (182–184). Oxygen is critical for the catabolism of H<sub>2</sub>S *via* mitochondrial oxidation, the main route of H<sub>2</sub>S disposal. During hypoxia, when intracellular oxygen is low, the catabolism of H<sub>2</sub>S was found to be greatly reduced (56), resulting in an increased concentration of H<sub>2</sub>S. Conversely, during normoxia, much of the intracellular H<sub>2</sub>S would be oxidized by available oxygen, resulting in a low intracellular level of H<sub>2</sub>S. In addition to H<sub>2</sub>S catabolism, the oxygen level was also found to affect H<sub>2</sub>S production. At a low concentration of O<sub>2</sub> (<5  $\mu$ M), H<sub>2</sub>S production was detectable in various rat tissues that diminished in an air-equilibrated buffer with 200  $\mu$ M O<sub>2</sub> (56).

Therefore, the tissue H<sub>2</sub>S concentration is inversely correlated with the oxygen concentration (184), making it a likely candidate for an oxygen sensing/signal transduction cascade involved in hypoxic response. The balance between endogenous H<sub>2</sub>S biosynthesis and its catabolism by available oxygen is considered the “oxygen sensor.”

### C. Atherosclerosis

Atherosclerosis, also known as arteriosclerotic vascular disease, is a chronic and complex condition that involves arterial wall thickening as a result of fat accumulation and plaque formation, mainly in large and medium-sized arteries. While the exact underlying mechanisms are not clearly understood, atherosclerotic plaque formation is thought to be mediated by a number of factors such as vascular inflammation, endothelial damage, smooth muscle proliferation and migration, macrophage and foam cell accumulation, and lipid deposition (68, 156). The consequential narrowing and stiffening of blood vessels impedes blood flow and increases plaque thrombogenicity. Current findings suggest that H<sub>2</sub>S, while it exerts proliferative roles in ECs that enhance angiogenesis, induces apoptosis in SMCs that protects against atherosclerosis (Figs. 12 and 13).

1. Potential therapeutic effects of H<sub>2</sub>S on atherosclerosis. CSE expression and activity and/or H<sub>2</sub>S level were significantly reduced during the development of atherosclerosis in either vascular beds or plasma using a balloon injury model (163) and a vascular calcification model (281). However, in apolipoprotein E knockout (apoE<sup>-/-</sup>) mice, CSE mRNA in aorta was found to be elevated, although plasma H<sub>2</sub>S and aortic H<sub>2</sub>S synthesis were lowered compared with wild-type mice (262). The discrepancy might be attributed to the existence of a positive compensatory feedback mechanism

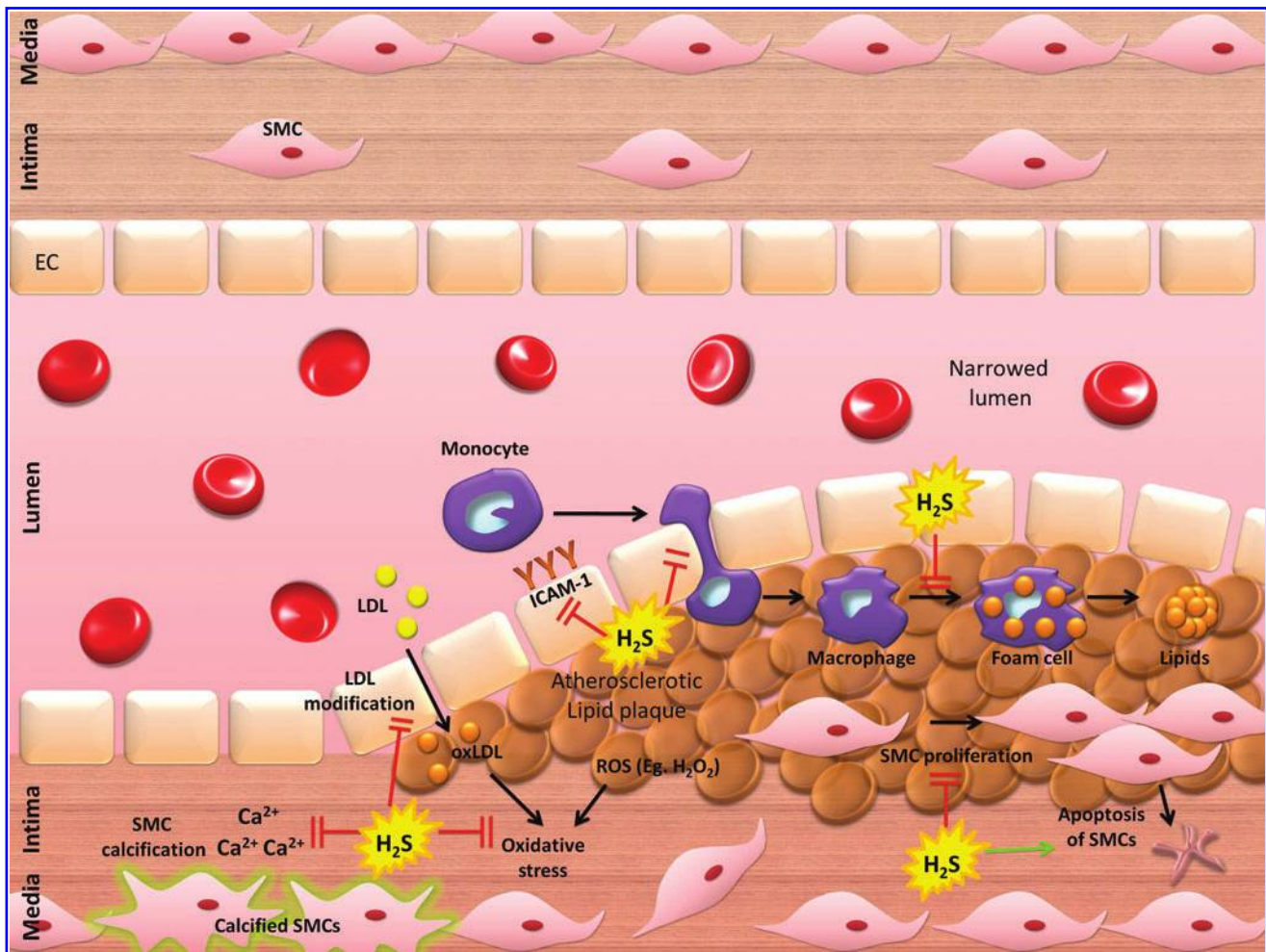
under *in vivo* conditions to salvage the low H<sub>2</sub>S level by CSE activity upregulation. In apoE<sup>-/-</sup> mice, PAG administration induced marked reductions in plasma H<sub>2</sub>S level and aortic H<sub>2</sub>S synthesis, and increases in CSE mRNA expression and the size of atherosclerotic plaques in comparison to untreated apoE<sup>-/-</sup> mice (262).

Exogenously administered NaHS suppressed the development of neointima hyperplasia (163), decreased vascular calcium content, calcium overload, and alkaline phosphatase activity in calcified vessels (281); reduced atherosclerotic plaque size; and improved aortic ultrastructure (262).

2. Mechanisms underlying the effects of H<sub>2</sub>S on atherosclerosis. The inhibitory effect of H<sub>2</sub>S on atherosclerosis is a novel discovery reported only within recent years; hence, the underlying mechanisms are not completely understood. A myriad of factors, including inhibition of SMC proliferation, anti-inflammatory responses, and attenuation of low-density lipoprotein (LDL) modification, are known as preventing the development of atherosclerosis.

a. Proapoptotic/antiproliferative effects of H<sub>2</sub>S on SMCs. The proliferation of VSMCs plays an important role in the structural remodeling of blood vessels. Many *in vitro* studies have been performed to investigate the involvement of the proapoptotic effect of H<sub>2</sub>S in the treatment of atherosclerosis. Cultured human aorta SMCs (HASMCs) treated with exogenous NaHS (200–500  $\mu$ M) showed a concentration-dependent increase in the number of apoptotic cells revealed by Hoechst 33258 staining and TUNEL assay, and further confirmed by internucleosomal DNA fragmentation (286). The physiological relevance of this finding is questionable, as H<sub>2</sub>S concentration utilized was beyond physiological. Du *et al.* also reported an antiproliferative effect of H<sub>2</sub>S on VSMCs by measuring the incorporation of [<sup>3</sup>H]-thymidine into cells (59). S-diclofenac (10–100  $\mu$ M), a slow releasing H<sub>2</sub>S donor, also attenuated cell proliferation in both primary (A-10) SMCs and immortalized (SV-40 transformed) SMCs (10).

Interestingly, when endogenous synthesis of H<sub>2</sub>S was decreased by inhibiting CSE with PAG, the increased cell apoptosis induced by exogenously administered H<sub>2</sub>S (100  $\mu$ M) was increased. However, PAG treatment alone failed to exert any proapoptotic effect (286). These authors explained this observation by proposing that endogenous H<sub>2</sub>S acts in suppressing or desensitizing cells to apoptotic signaling. Therefore, PAG unveiled an increased response of HASMCs to an apoptosis-inducing concentration of H<sub>2</sub>S (286). These researchers later established a CSE overexpression model by infecting HASMCs with a recombinant defective adenovirus containing the CSE gene (Ad-CSE) (289). Ad-CSE transfection resulted in a 12.3-fold increase in CSE activity and a 6.2-fold increase in endogenous H<sub>2</sub>S production rate. CSE overexpression significantly reduced cell proliferation, shown by a cell growth curve and a cell viability assay. In line with this, CSE overexpression promoted cell apoptosis, as seen in Hoechst 33258 staining and TUNEL assay, together with the activation of caspase-3. In order to confirm that the proapoptotic effect of CSE overexpression is attributable to H<sub>2</sub>S production but not products of other CSE-catalyzed reactions, CSE gene activity was knocked down by the RNA interference approach, and H<sub>2</sub>S was then exogenously administered. As expected, H<sub>2</sub>S (100  $\mu$ M) promoted the apoptosis of



**FIG. 12. H<sub>2</sub>S inhibits atherosclerosis.** H<sub>2</sub>S suppresses (indicated by red line indicators) intercellular adhesion molecule-1 (ICAM-1) expression and prevents monocyte infiltration into atherosclerotic plaques that form macrophages. Furthermore, when the macrophages are exposed to oxidized low-density lipoprotein (oxLDL), foam cells are formed, which make up the atherosclerotic core. H<sub>2</sub>S prevents the formation of atherosclerotic plaques by preventing the modification of LDL along with the inhibition of foam cell formation. Moreover, H<sub>2</sub>S acts on SMCs that prevent SMC proliferation and calcification, while SMC apoptosis is enhanced (indicated by green arrows). H<sub>2</sub>S is also responsible for scavenging ROS and reducing the oxidative stress of atherosclerotic tissues (To see this illustration in color, the reader is referred to the web version of this article at [www.liebertonline.com/ars](http://www.liebertonline.com/ars)).

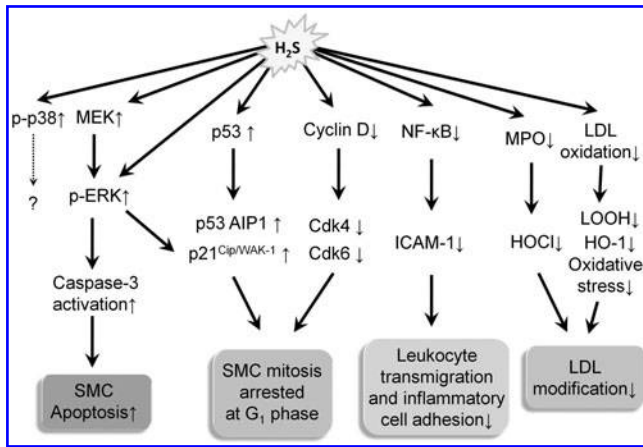
HASMCs, while ammonium and pyruvate (two other products of a CSE-catalyzed reaction) exerted no induction effect on cell apoptosis (289).

The effect of endogenous H<sub>2</sub>S on apoptosis in VSMCs was investigated in CSE-knockout mice (287). Consistent with observations in the CSE overexpression model, CSE-knockout mice exhibited SMC overproliferation in the media of the aorta. These data collectively suggest that the endogenous CSE/H<sub>2</sub>S system plays a role in suppressing SMC proliferation while enhancing apoptosis.

(1). **Signaling mechanisms.** Exogenously administered H<sub>2</sub>S (20–500  $\mu$ M) activated ERK and p38 MAPK in a concentration-dependent manner (286). In line with this, H<sub>2</sub>S produced endogenously *via* CSE overexpression with Ad-CSE transfection was also found to upregulate the activation of ERK and p38 MAPK (289). In CSE-knockout mice, a reduced level of ERK1/2 activation was observed as a result of de-

creased endogenous H<sub>2</sub>S production (287). In contrast to these findings, Du *et al.* failed to observe MAPK activation when VSMCs were treated with NaHS at 50–100  $\mu$ M in the absence of fetal bovine serum (FBS) or endothelin-1 (59). In fact, in the presence of FBS or endothelin-1, NaHS (50–500 or 50–100  $\mu$ M, respectively) significantly reduced the activity of ERK1/2 in a concentration-dependent manner. Nevertheless, these results suggest that the H<sub>2</sub>S-induced inhibitory effect on VSMC proliferation is, in part, mediated *via* the MAPK pathway (59).

The blockade of MEK was capable of attenuating a H<sub>2</sub>S (500  $\mu$ M)-elicited increase in ERK activation, caspase-3 activation, and the induction of apoptosis (286), suggesting that H<sub>2</sub>S-induced apoptosis in VSMCs is mediated *via* the MEK/ERK/Caspase-3 pathway. On the other hand, the administration of SB203580, a powerful inhibitor of p38 phosphorylation, failed to inhibit the effect of H<sub>2</sub>S (500  $\mu$ M) on caspase-3 activation and cell apoptosis. Moreover, the inhibition of caspase-3 has no influence on the H<sub>2</sub>S-induced p38 activation (286). Thus, unlike



**FIG. 13. Signaling mechanisms underlying H<sub>2</sub>S inhibition on atherosclerosis.** H<sub>2</sub>S could elevate phosphorylation (prefix p-) of both extracellular signal-regulated kinases (ERK) and p38, but only the ERK pathway was capable of caspase-3 activation and induction of the proapoptotic effect on SMCs. H<sub>2</sub>S may regulate cell-cycle progression *via* the inhibition of cyclin D1 expression and the upregulation of p53, p53 apoptosis-inducing protein (AIP) 1, and p21 Cip/WAK-1. H<sub>2</sub>S inhibition of leukocyte transmigration and inflammatory cell adhesion is mediated by its inhibitory effect on nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) and subsequent downregulation of ICAM-1. H<sub>2</sub>S also prevents modifications of LDL by the inhibition of LDL oxidation, lowering heme oxygenase 1 (HO-1) expression, lipid hydroperoxide (LOOH) levels and reducing oxidative stress. The downregulation of the MPO/H<sub>2</sub>O<sub>2</sub>/Cl<sup>-</sup> system by H<sub>2</sub>S may also contribute toward the prevention of atherosclerotic plaque developments. MPO, myeloperoxidase.

MEK/ERK, the p38 pathway is probably not involved in H<sub>2</sub>S-induced proapoptotic responses.

The involvement of these pathways warrants further research efforts using lower H<sub>2</sub>S concentrations (below 100 μM), as concentrations beyond this range are probably not physiological.

**(2). Cell-cycle regulators.** A family of protein kinases, cyclin-dependent kinases (cdk), may associate with cyclins to form cyclin-Cdk complexes and trigger cell-cycle events. H<sub>2</sub>S appears to interfere with cell-cycle regulation, as both NaHS (50 μM) and S-diclofenac (50 μM) were found to decrease the percentage of cells in the G<sub>1</sub> phase after 24 and 48 h of treatment. Further studies showed that S-diclofenac altered the cell cycle and prevented the mitosis of asynchronized cells (G<sub>1</sub> phase). Its effect was much less in synchronized cells (G<sub>0</sub> phase) (10).

Both endogenously produced H<sub>2</sub>S (CSE overexpression) or exogenously applied H<sub>2</sub>S (100 μM) suppressed the expression of cyclin D1 (an activator of cdk4 and cdk6) while elevating the expression of p21<sup>Cip/WAK-1</sup> (a cdk inhibitory protein) (289). An increase in p21 protein level was also induced by S-diclofenac, accompanied by a near fourfold increase in p53. Proapoptotic proteins such as p53 apoptosis-inducing protein (AIP) 1 and Bax were also time-dependently elevated by S-diclofenac (10). Intriguingly, it was previously reported that treatment of HASMCs with H<sub>2</sub>S (20–500 μM) failed to alter the protein expression of Bax (286). Such a discrepancy warrants further studies. Nevertheless, the lack of Bcl-2 involvement in

mediating H<sub>2</sub>S-induced proapoptosis was consistent in both studies (10, 286).

Using both isolated SMCs and whole vascular tissues, Yang *et al.* demonstrated that the reduced level of endogenous H<sub>2</sub>S in CSE-knockout mice correlates with lowered p21<sup>Cip/WAF-1</sup> expression but enhanced cyclin D1 expression when compared with wild-type mice (287). The exogenous administration of H<sub>2</sub>S could increase the expression of p21<sup>Cip/WAF-1</sup> while attenuating the expression of cyclin D1 in both CSE-knockout and wild-type mice, but with a much greater effect in the former. In addition, the CSE-knockout mice expressed a higher level of CDK4, which could be attenuated by exogenously applied H<sub>2</sub>S. Exogenous H<sub>2</sub>S administration elevated the expression of p15<sup>INK4B</sup> and p27<sup>Kip1</sup> in the CSE-knockout mice, but not in wild-type mice. Surprisingly, SMCs of CSE wild-type, but not of CSE knockout, expressed an increased level of cyclin D3 in response to exogenous H<sub>2</sub>S. The expression of p16<sup>INK4A</sup> increased by a similar extent in both CSE-knockout and wild-type mice. The gene expression using microarray analysis demonstrated that the CSE-knockout mice express a higher level of calcitonin receptor-like, integrin β<sub>1</sub>, heparin-binding epidermal growth factor-like growth factor in aortic tissues than that in their wild-type counterparts (287).

**b. Anti-inflammation.** ICAM-1 facilitates leukocyte transmigration and inflammation cell adhesion to the endothelium. The administration of NaHS suppressed ICAM expression in both the plasma and aorta of apoE<sup>-/-</sup> mice, whereas PAG enhanced the upregulation of ICAM expression. The inhibitory effect of NaHS (1–100 μM) on the ICAM-1 protein level was further reaffirmed in TNF-α-elicited HUVECs. These data suggest that H<sub>2</sub>S may exert its protective effect against atherosclerosis formation *via* the downregulation of ICAM-1 under both *in vivo* and *in vitro* conditions (262).

NF-κB is a transcription factor involved in the stimulation of ICAM-1 expression in ECs. NaHS (100 μM) significantly inhibited NF-κB activation, indicated by an attenuated breakdown of the NF-κB inhibitor (IκBα) and nuclear translocation of NF-κB (262).

The formation of foam cells contributes toward the formation of atherosclerosis. Macrophages are converted into foam cells after being exposed to oxidized LDL (oxLDL). H<sub>2</sub>S prevented oxLDL-induced foam cell formation in a concentration-dependent manner (262, 311) along with blunted oxLDL binding and uptake in macrophages (311). Studies unveiling the underlying mechanisms revealed that NaHS (50–200 μM) suppresses oxLDL-induced elevated expressions of scavenger receptor CD36, scavenger receptor A, and acyl-coenzyme A: cholesterol acyltransferase-1 mediated by the K<sub>ATP</sub>/ERK1/2 signaling pathway (311).

**c. Inhibition of LDL modification.** The oxidative modification of LDL is implicated in the early development of arterogenesis. Hypochlorite (HOCl), the product of the activated myeloperoxidase/H<sub>2</sub>O<sub>2</sub>/chloride (MPO/H<sub>2</sub>O<sub>2</sub>/Cl<sup>-</sup>) system, has been implicated as a trigger for LDL modification, and HOCl-modified LDL has been found in atherosclerotic plaques (125). The elevation of Apo B modifications (assessed by relative electrophoretic mobility and carbonyl formation) in LDL could be attenuated by either the exogenous administration of NaHS (750 μM–3 mM) or the endogenous precursors of H<sub>2</sub>S (cysteine, homocysteine, or methionine) (125). The inactivation

of HOCl-induced LDL modification could be attributed to the scavenging property of H<sub>2</sub>S, which was demonstrated using the 3,3',5,5'-tetramethylbenzidine assay to quench equimolar concentrations of HOCl. Moreover, H<sub>2</sub>S was further proved to inactivate MPO enzyme activity and decompose H<sub>2</sub>O<sub>2</sub>, suggesting that the antiatherosclerotic property of H<sub>2</sub>S is mediated by downregulation of the MPO/H<sub>2</sub>O<sub>2</sub>/Cl<sup>-</sup> system that limits LDL modifications. The high H<sub>2</sub>S concentrations used in these studies were far beyond the physiological and may be of little value to our understanding of H<sub>2</sub>S biology *in vivo*.

In another study, the LDL oxidation model was established by hemin exposure as inducing EC damage either directly or *via* the induction of oxidative alterations in lipids and apolipoproteins to form cytotoxic oxidized products (101). The administration of NaHS (5–20  $\mu$ M) during hemin-induced LDL modification resulted in the concentration-dependent inhibition of LDL oxidation, represented by the retarded formation of lipid peroxidation products such as conjugated dienes, lipid hydroperoxide (LOOH), and thiobarbituric acid-reactive substrates (TBARS) (101). In both HUVECs and atherosclerotic plaques, NaHS (25–200  $\mu$ M) treatment, subsequent to the induction of LDL oxidation, lowered the LOOH level in a concentration-dependent manner, while not affecting conjugated dienes and TBARS. This phenomenon may account for the cytoprotection seen in ECs subjected to H<sub>2</sub>S treatment, as elevated LOOH content is implicated in cytotoxicity (177). Moreover, the oxidized LDL was found to upregulate the expression of HO-1,

and H<sub>2</sub>S (25–200  $\mu$ M) could concentration dependently reduce HO-1 mRNA and protein induction (101). H<sub>2</sub>S may also exert its cytoprotective effects on ECs *via* its antioxidant property, as it was found to reduce oxidative stress elicited by H<sub>2</sub>O<sub>2</sub> or oxidized LDL (101).

## VII. Hypertension

Hypertension is the most common CVD and a leading risk factor for stroke or heart problems. In the United States, for example, hypertension has a prevalence rate of about 40% (204). Generally, H<sub>2</sub>S is a vasodilator (in the short term, it relaxes vascular smooth muscles) and is able to produce an antihypertensive effect. Studies demonstrating the antihypertensive effects of H<sub>2</sub>S are summarized in Table 4.

### A. Effect of H<sub>2</sub>S on BP of normotensive animals

The blockade of endogenous H<sub>2</sub>S production by hydroxylamine hydrochloride (0.5 mg/kg *i.p.*), a nonspecific inhibitor of both CSE and CBS, for over 4 weeks failed to influence systolic blood pressure (SBP) in rats (154). In contrast, Yan *et al.* found that the administration of PAG (37.5 mg/kg), an inhibitor of CSE, to rats for 5 weeks significantly elevated BP and increased the medial cross-sectional area of the thoracic aorta wall, indicating significant aortic structural remodeling (284). The discrepancy may arise from the different drugs used in the two studies, as well as from the different dosages and duration of treatment. It should be noted that existing

TABLE 4. ANTIHYPERTENSIVE EFFECT OF HYDROGEN SULFIDE IN DIFFERENT HYPERTENSIVE ANIMAL MODELS

Animal model	Endogenous H <sub>2</sub> S level	Treatment	Effect of exogenous H <sub>2</sub> S	Mechanisms	Ref.
Genetic model-SHR	Plasma H <sub>2</sub> S ( $\downarrow$ ), aortic CSE mRNA ( $\downarrow$ ) aortic CSE activity ( $\downarrow$ )	NaHS 56 $\mu$ mol/kg or PAG (37.5 mg/kg) <i>i.p.</i> for 5 weeks	NaHS treatment: Antihypertension, Plasma H <sub>2</sub> S ( $\uparrow$ ), aortic CSE mRNA ( $\uparrow$ ) aortic CSE activity ( $\uparrow$ ) PAG treatment: hypertension development and vascular remodeling	Lessened aorta remodeling	(284, 310)
SHR	NA	NaHS (10, 30, 90 $\mu$ mol/kg/day for 3 m)	Antihypertension and antihypertrophy	Antioxidant	(219)
SHR and L-NAME-induced hypertensive rats	NA	GYG (133 $\mu$ mol/kg/day for 14 days, <i>iv</i> or <i>ip</i> )	Antihypertension	K <sub>ATP</sub> channels	(138)
2K1C renin-dependent hypertensive rats	NA	NaHS (0.56, 1.68, 5.6 mg/kg/day for 4 m)	Antihypertension	Renin-inhibition	(154)
1K1C renin-independent hypertensive rats	NA	NaHS (0.56, 1.68, 5.6 mg/kg/day for 4 m)	No effect	NA	(154)
Aorta-inferior cava vein shunting pulmonary hypertension	Plasma and lung H <sub>2</sub> S ( $\downarrow$ ), CSE mRNA ( $\downarrow$ )	NaHS (56 $\mu$ mol/kg/day) for 11 weeks	Antipulmonary hypertension	NA	(283)
Hypoxia Pulmonary hypertension Rats	Plasma H <sub>2</sub> S ( $\downarrow$ ), aortic CSE mRNA ( $\downarrow$ ) aortic CSE activity ( $\downarrow$ )	NaHS 14 $\mu$ mol/kg	Antipulmonary hypertension	Lessened vascular remodeling	(48)

1K1C, one-kidney-one-clip; 2K1C, two-kidney-one-clip; L-NAME, N (G)-nitro-L-arginine methyl ester; SHR, spontaneously hypertensive rats; NA, not applicable.

H<sub>2</sub>S synthesizing enzyme inhibitors have low potency and specificity for their intended use, and the data obtained should be interpreted with caution.

A better alternative than using CSE inhibitors in animal experiments would be the use of genetically modified animals. CSE-knockout mice were reported to exhibit pronounced BP elevations compared with their wild-type counterparts (288), suggesting that the physiological production of H<sub>2</sub>S by CSE may be critical for the regulation of basal BP. However, the possibility that hypertension in CSE-knockout animals developed as a result of hyperhomocysteinemia cannot be totally excluded. Although the researchers explained that male and female knockout mice have similar BP despite vastly different homocysteine levels, BP control mechanisms differ between genders, and factors such as sex hormones could have counteracted the effect of homocysteines. In a further attempt to exclude the contribution of hyperhomocysteinemia, Yang *et al.* carried out a separate set of experiments. L-methionine was administered to wild-type mice *via* drinking water for 6 weeks to induce hyperhomocysteinemia, and no hypertension was observed in these animals (288). Unfortunately, plasma homocysteine levels in L-methionine-treated animals increased by merely 1.5 times, a huge disparity compared with the 18-fold difference between CSE-knockout and wild-type mice. In contrast to the findings of Yang *et al.*, the CSE-knockout mice developed by Ishii *et al.* did not exhibit hypertension (99), whereas L-methionine-induced hyperhomocysteinemia resulted in elevated BP (233). The great disparity in observations and the complexity of the science involved definitely warrant further research efforts that study these effects in greater detail.

The acute administration of H<sub>2</sub>S causes transient dose-related falls in MAP without significant alterations in heart rate (3, 138, 288, 309). For example, Li and colleagues demonstrated that an i.v. bolus injection of NaHS (2.5 to 20  $\mu$ M) caused an immediate fall in BP in anesthetized rats, and this transient effect lasted for only 10 to 30 s (138). However, the chronic administration of NaHS (5.6 mg/kg/day, i.p.) for 4 weeks failed to significantly affect SBP in conscious rats, measured using the tail-cuff method (154). Similarly, exoge-

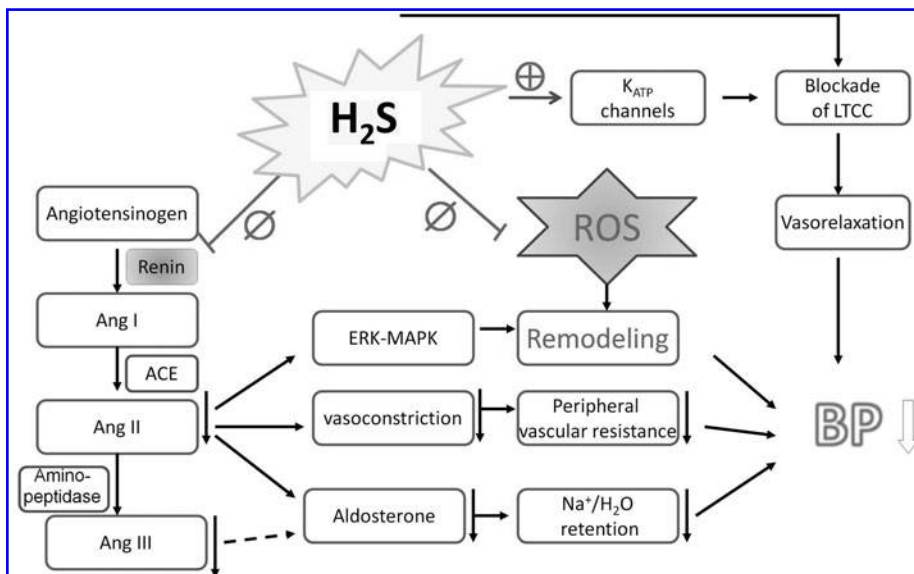
nous NaHS (56  $\mu$ mol/kg, i.p.) administered for 5 weeks did not affect the SBP and aortic structure in normotensive rats (284). Therefore, it appears that H<sub>2</sub>S does not significantly affect the BP in normotensive rats.

### B. Effect of H<sub>2</sub>S on BP in hypertensive animals

Several groups investigated the effect of H<sub>2</sub>S on hypertension in different hypertensive models. Treatment with H<sub>2</sub>S can significantly lower the BP of hypertensive animals (Fig. 14). We will first introduce the antihypertensive effects of H<sub>2</sub>S in these models followed by discussing the underlying mechanisms.

1. **H<sub>2</sub>S in SHR.** Yan *et al.* found that the plasma level of H<sub>2</sub>S in SHR was significantly lower than that of the WKY controls (284). The expression of CSE mRNA was also found to be lower than that of control rats. These findings suggest that the hypertension in SHR involves a reduction in the production and function of H<sub>2</sub>S (284). This is supported by the observation that chronic daily administration of NaHS (56  $\mu$ mol/kg/day, i.p.) for 5 weeks significantly reduced the SBP by 25 mm Hg and inhibited aortic structural remodeling in the SHR (284). Consistently, Shi *et al.* reported significantly reduced SBP, diastolic blood pressure (DBP), and MAP of SHR to similar extents after NaHS (30 or 90  $\mu$ mol/kg/day, i.p.) for 3 months. A lower dose (10  $\mu$ mol/kg/day i.p.) also reduced DBP and MAP, but the decrease in SBP was not statistically significant (219). In addition, chronic treatment with GYY4137 (a slow-releasing H<sub>2</sub>S donor, 133  $\mu$ mol/kg, i.p.) also significantly reduced the SBP in SHR from day 2 after treatment (138). Therefore, H<sub>2</sub>S has a profound antihypertensive effect in SHR.

2. **H<sub>2</sub>S in renovascular hypertension.** The two-kidney-one-clip (2K1C, a clip constricting one renal artery) model is a good animal model of renovascular hypertension, which is a common type of secondary hypertension and the most prevalent form of curable hypertension (201). The removal of the unclipped kidney gives rise to the one-kidney-one-clip (1K1C) model. The mechanism underlying



**FIG. 14. Effect of H<sub>2</sub>S on blood pressure (BP) of hypertensive animals.** H<sub>2</sub>S lowers the BP of hypertensive animals mainly *via* the inhibition of renin, as H<sub>2</sub>S could significantly attenuate the rise in the BP of renin-dependent renal hypertension but not that of renin-independent mechanisms. Other contributing factors of H<sub>2</sub>S may include the scavenging of ROS and the inhibition of the K<sub>ATP</sub> channels.

the hypertensive effect of 2K1C and 1K1C models differs. Hypertension in the 2K1C model results from an increased release of renin and aldosterone (23, 118). In the 1K1C model, on the other hand, the renin level increased only transiently, and is considered a trigger for the development of hypertension, which is then maintained by Na<sup>+</sup>/H<sub>2</sub>O retention. (70, 71).

Lu *et al.* found that the administration of NaHS (5.6 mg/kg/day, i.p.) successfully attenuated the development of hypertension in the renin-dependent 2K1C model (154) (Fig. 15). Furthermore, NaHS treatment also reversed the high BP after hypertension had developed in the 2K1C rats. On the contrary, NaHS administration failed to affect the BP in the renin-independent 1K1C model (154). These data suggest that the antihypertensive effect of H<sub>2</sub>S is mediated *via* a renin-dependent mechanism.

**3. H<sub>2</sub>S in pulmonary hypertension.** Pulmonary hypertension (PH) is a common complication of congenital heart disease with a left-to-right shunt characterized by high pulmonary blood flow (283) in which blood from the arteries is mixed with that of the veins. Xiaohui *et al.* found that the H<sub>2</sub>S level in lung tissue was decreased, while the mean pulmonary arterial pressure (mPAP) increased significantly 11 weeks after aorta-veno cava shunting, which was utilized as a rat

model for PH. This suggests that a reduced level of H<sub>2</sub>S may contribute to the detrimental effect of shunting (283).

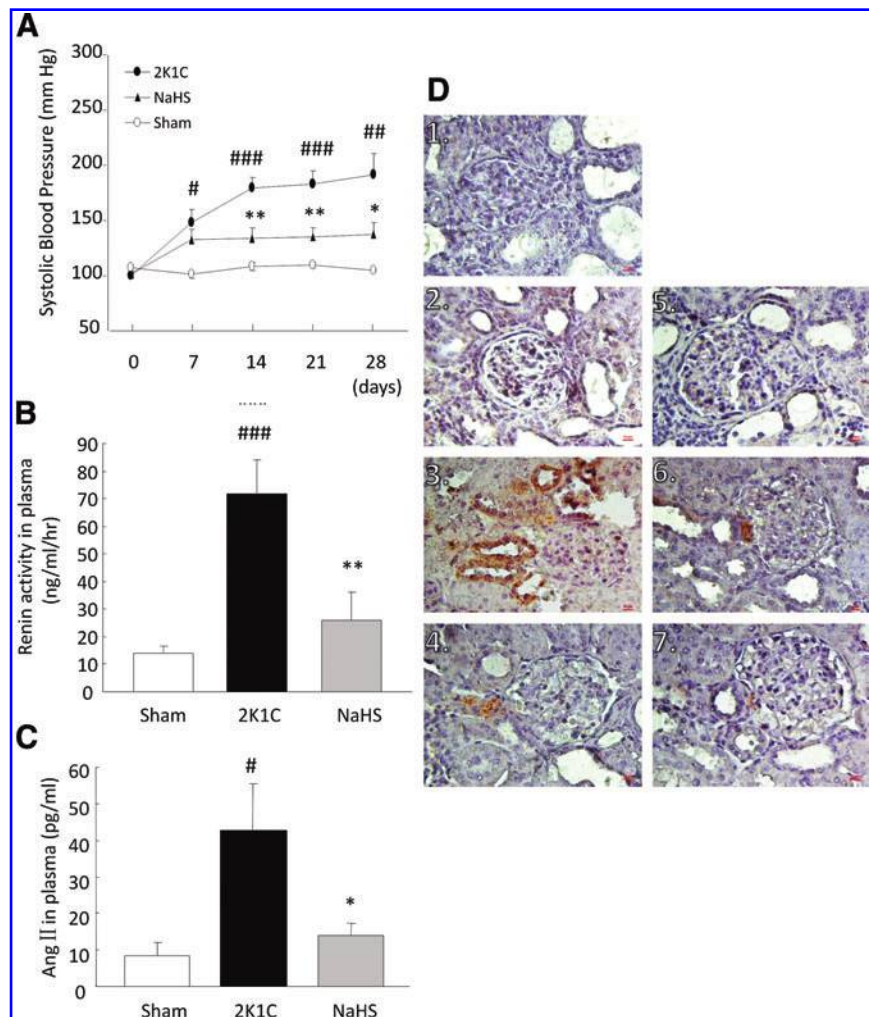
Hypoxic pulmonary hypertension (HPH) is another important pathophysiological process that commonly occurs in several cardiac and pulmonary diseases. It is characterized by high pulmonary arterial pressure and high pulmonary vascular resistance, often resulting from a combination of hypoxic vasoconstriction and vascular structural remodeling (303). It was found that the CSE mRNA level, H<sub>2</sub>S synthesis, and plasma level of H<sub>2</sub>S were all significantly decreased in the rat HPH model. Conversely, the exogenous administration of NaHS completely abolished hypoxia-induced mPAP elevation, and greatly attenuated right ventricular hypertrophy (303). These findings suggest that the impaired H<sub>2</sub>S production might contribute to the pathogenesis of HPH, and the exogenous application of H<sub>2</sub>S may have therapeutic value in the treatment of HPH.

#### 4. Mechanisms underlying antihypertensive effects of H<sub>2</sub>S

*a. Inhibition of the renin-angiotensin system.* Renin-angiotensin system (RAS) plays an important role in regulating blood volume and systemic vascular resistance, which influence BP. Renin catalyzes the first and rate-limiting step in the

**FIG. 15. Effect of H<sub>2</sub>S on renin-dependent renal hypertension.** (A). Time course for the development of renovascular hypertension in the presence and absence of NaHS treatment (5.6 mg/kg/day) in two-kidneys-one-clip (2K1C) rats over 4 weeks. NaHS treatment significantly reversed the BP elevations as compared with the 2K1C untreated group (B and C). Elevations in renin (B) and angiotensin II (C) levels were significantly attenuated by NaHS treatment. Data are expressed as mean  $\pm$  SEM, #*p* < 0.05, ###*p* < 0.01 and ####*p* < 0.001 versus sham rats. \**p* < 0.05 and \*\**p* < 0.01 versus 2K1C rats (D). NaHS treatment decreased renin expression in the clipped kidneys. (1) Negative control stained by preimmune mouse IgG. Renin expression increased drastically in (3) clipped kidneys compared with those in (2) the sham group. (4) NaHS treatment markedly attenuated the upregulation of renin expression in the clipped kidneys (5 through 7). There was no significant change of renin expression in the unclipped kidneys of 2K1C rats along (5) sham, (6) vehicle, (7) and NaHS treatment groups.

The photos were taken at  $\times 40$  magnification [Adapted with permission from *Journal of the American Society of Nephrology* (Ref. 154)]. (To see this illustration in color, the reader is referred to the web version of this article at [www.liebertonline.com/ars](http://www.liebertonline.com/ars)). NaHS, sodium hydrosulfide (H<sub>2</sub>S donor).



production of Ang II. Lu *et al.* found that H<sub>2</sub>S decreased plasma renin activity (PRA) in renin-dependent 2K1C rats, but had no significant effect on PRA in normotensive or renin-independent 1K1C rats (154). H<sub>2</sub>S suppressed renin degranulation and release was further shown to be mediated by the attenuation of AC activity and lowered cAMP production (155). Although NaHS (1 mM) also exhibited an inhibitory effect on PDE activity, the high concentration of H<sub>2</sub>S needed has limited physiological relevance and implications. The authors also reported that neither acute (up to 1 mM, 30 min) nor chronic (5.6 mg/kg/day for 4 weeks) treatment with H<sub>2</sub>S significantly affected the angiotensin converting enzyme (ACE) activity in normal (basal ACE activity) or hypertensive (elevated ACE activity) rat aortic tissues (154). Laggner *et al.* however, found that H<sub>2</sub>S (60  $\mu$ M) inhibited ACE activity, but not its mRNA expression, on the surface of cultured HUVEC monolayers, *ex vivo* in umbilical veins and in HUVEC protein extracts (124). The inconsistent H<sub>2</sub>S effect on ACE could be attributed to the different type of blood vessel used. Nevertheless, H<sub>2</sub>S inhibition of renin or ACE would lead to significantly attenuated Ang II. For example, H<sub>2</sub>S inhibited the fivefold elevation of Ang II level in the plasma of 2K1C rats through the inhibition of renin (154).

Zhao *et al.* also observed that H<sub>2</sub>S can decrease the binding affinity of Ang II type 1 (AT1) receptors, thereby attenuating Ang II-induced AT1 activation. This would lead to a reduction in downstream signaling, including MAPK and ERK 1/2 activation (310). Collectively, these findings indicate that the inhibition of various RAS components plays a substantial role in the underlying H<sub>2</sub>S-induced antihypertensive effects.

*b. Inhibition of vascular remodeling.* Vascular structural remodeling is one important factor in the development of hypertension. In addition, elevated BP itself promotes blood vessel remodeling. Therefore, structural remodeling of small arterioles is a target for the treatment of hypertension. Vascular remodeling is linked to RAS, as the interruption of the RAS with ACE inhibitors or AT1 receptor antagonists corrects vascular structure (209), and ERK1/2 signaling stimulated by Ang II is often associated with VSMC hypertrophy (120). H<sub>2</sub>S also reversed the elevated hydroxyproline and collagen type I levels (310). Another mechanism for the preventive/therapeutic effects of NaHS on vascular structural remodeling may be its antioxidant effects. H<sub>2</sub>S may reduce the tissue ROS level, which is important in vascular remodeling.

*c. Activation of K<sub>ATP</sub> channels.* The administration of GYY4137, a water-soluble compound that slowly releases H<sub>2</sub>S, can cause an increase in the plasma H<sub>2</sub>S concentration for up to 180 min (138). This slow and sustained increase of the H<sub>2</sub>S level may produce an antihypertensive effect *via* the activation of K<sub>ATP</sub> channels, which hyperpolarizes and, therefore, relaxes vascular smooth muscle. However, it was observed that BP only returned to normal 7 days after the last injection, implying that other long-term regulatory mechanisms probably exist.

### VIII. Concluding Remarks

Current evidence indicates that H<sub>2</sub>S plays important roles in mediating cardiovascular functions under both physio-

logical and pathological conditions. H<sub>2</sub>S inhibits heart contractile functions *via* its inhibitory effects on the  $\beta$ -adrenergic system and intracellular calcium handling. These observations are further supported with discoveries that H<sub>2</sub>S exerts inhibitory effects on LTCC and chloride channels, while it stimulates the K<sub>ATP</sub> channels. In the vascular system, *in vitro* tissue contractility studies demonstrated that NaHS induces vasoconstriction at lower concentrations (10–100  $\mu$ M) while exerting vasodilatory responses at concentrations above about 100  $\mu$ M. Though uncertainties still persist in the determination of endogenous H<sub>2</sub>S level in plasma and tissues, the latest consensus favors the notion that free H<sub>2</sub>S exists in the nanomolar range. However, tissues may contain stored sulfide, probably near a low micromolar range, and this sulfide can be released under specific conditions. It is still thought by many that endogenous H<sub>2</sub>S produces vasodilation under physiological conditions. Thus, local micro environments may contain much higher concentrations of free H<sub>2</sub>S than those found in tissue samples.

It is important to note that the endogenous production of H<sub>2</sub>S appears to be significantly reduced in many disease states, including myocardial ischemia; MI-induced or AVF-induced CHF; spontaneous, pulmonary, or hyperhomocysteinemia-induced hypertension; vascular calcification; or apoE<sup>-/-</sup> atherosclerotic models. Whether reduction of H<sub>2</sub>S level is the cause or consequence of these pathological states is not well understood. However, these findings are clear evidence that support the involvement of endogenous H<sub>2</sub>S in maintaining basal physiological functions.

Another exciting aspect of H<sub>2</sub>S research will be the crosstalk among H<sub>2</sub>S and other gaseous transmitters—NO and CO. Efforts have been made to understand the interaction between NO and H<sub>2</sub>S. Though great disparities and complexities exist in research outcomes, these interactions have important potential implications for our understanding of the cardiovascular system and are of potential therapeutic value.

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### Abbreviations Used

[Ca<sup>2+</sup>]<sub>i</sub> = intracellular calcium  
 [cAMP]<sub>i</sub> = intracellular cAMP  
 [K<sup>+</sup>]<sub>i</sub> = intracellular potassium  
 [Na<sup>+</sup>]<sub>i</sub> = intracellular sodium+dl  
 dt = maximal velocity of cell shortening  
 1K1C = one-kidney-one-clip  
 2K1C = two-kidney-one-clip  
 3-MST = mercaptopyruvate sulphurtransferase  
 4-AP = 4-aminopyridine  
 5-HD = 5-hydroxydecanoate  
 AA = arachidonic acid  
 AC = adenylyl cyclase  
 ACE = angiotensin converting enzyme  
 Ach = acetylcholine  
 ADAM-12 = disintegrin and metalloproteinase domain-containing protein 12  
 Ad-CSE = adenovirus containing CSE gene  
 ADR = adriamycin  
 ADRF = adipocyte-derived relaxing factor  
 AE = anion exchanger  
 AIF = apoptosis-inducing factor  
 AIP = apoptosis-inducing protein  
 Ang II = angiotensin II  
 AOAA = amino-oxyacetate  
 APD = action potential duration  
 apoE = apolipoprotein E  
 apoE<sup>-/-</sup> = apolipoprotein E knockout  
 AT1 = Ang II type 1  
 ATP = adenosine triphosphate  
 AVF = arteriovenous fistula  
 BCA = β-cyano-L-alanine  
 Bcl-2 = B-cell lymphoma 2  
 bFGF = basic fibroblast growth factor  
 BP = blood pressure  
 CAD = coronary artery disease  
 cADPR = cyclic ADP ribose  
 cAMP = cyclic adenosine monophosphate  
 CAT = cysteine aminotransferase  
 CBE = Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchangers  
 CBS = cystathionine β-synthase  
 cdk = cyclin-dependent kinase  
 cGMP = cyclic guanosine monophosphate  
 CHF = chronic heart failure  
 CLT = chloramine T  
 CO = carbon monoxide  
 CoCl<sub>2</sub> = cobalt chloride  
 COX = cyclooxygenase  
 CPB = cardiopulmonary bypass  
 CSE = cystathionine γ-lyase  
 CVD = cardiovascular diseases  
 DBP = diastolic blood pressure  
 DNS-Az = dansyl azide  
 DTT = dithiothreitol  
 ECs = endothelial cells  
 EDHF = endothelium-derived hyperpolarizing factor  
 eNOS = endothelial NO synthase  
 ERK = extracellular signal-regulated kinase  
 FBS = fetal bovine serum  
 GC = gas chromatography

**Abbreviations Used (Cont.)**

GSH = glutathione  
 GSK-3 $\beta$  = glycogen synthase kinase-3  
 HA = hydroxylamine  
 HASMCs = human aorta SMCs  
 HF = heart failure  
 HHcy = hyperhomocysteinemia  
 HIF = hypoxia inducible factor  
 HNO = nitroxyl anion  
 HO-1 = heme oxygenase-1  
 HOCl = hypochlorite  
 HPH = hypoxic pulmonary hypertension  
 H<sub>2</sub>S = hydrogen sulfide  
 HSP = heat shock protein  
 HUVECs = human umbilical vein endothelial cells  
   i.p. = intraperitoneal  
   i.v. = intravenous  
 I/R = ischemia/reperfusion  
 ICAM-1 = intercellular adhesion molecule-1  
 IL = interleukin  
 IPreC = ischemic preconditioning  
 ISO = isoproterenol  
 K<sub>ATP</sub> = ATP-sensitive potassium  
   K<sub>v</sub> = voltage-dependent K<sup>+</sup>  
 LAD = left anterior descending  
 LCA = left coronary artery  
 LDH = lactate dehydrogenase  
 LDL = low density lipoprotein  
 L-NAME = N (G)-nitro-L-arginine methyl ester  
 LOOH = lipid hydroperoxide  
 LPO = lipid hydroperoxidation  
 LTCC = L-type Ca<sup>2+</sup> channels  
 LV = left ventricle  
 LVDP = left ventricular developed pressure  
 LVdp/dt<sub>max</sub> = maximal left ventricular pressure development  
 LVEDP = left ventricular end diastolic pressure  
 MAP = mean arterial pressure  
 MAPK = mitogen-activated protein kinase  
 MBB = monobromobimane  
 MEK = MAPK/ERK kinase  
 MI = myocardial infarction  
 MLC = myosin light chain  
 MMP = matrix metalloproteinases  
 mPAP = mean pulmonary arterial pressure  
 MPO = myeloperoxidase  
 mPTP = mitochondrial permeability transition pore  
 NADPH = nicotinamide adenine dinucleotide phosphate  
 NaHS = sodium hydrosulfide (H<sub>2</sub>S donor)

NCX = Na<sup>+</sup>/Ca<sup>2+</sup> exchanger  
 NEM = N-ethylmaleimide  
 NF- $\kappa$ B = nuclear factor kappa-light-chain-enhancer of activated B cells  
 NHE = Na<sup>+</sup>/H<sup>+</sup> exchanger  
 NO = nitric oxide  
 NOS = NO synthase  
 NRF-1 = nuclear respiratory factor 1  
 Nrf2 = nuclear factor-erythroid-derived 2 related factor 2  
   O<sub>2</sub><sup>-</sup> = superoxide anion  
 ONOO<sup>-</sup> = peroxynitrite  
 oxLDL = oxidized low-density lipoprotein  
 PAG = propargylglycine  
 PDE = phosphodiesterase  
 PH = pulmonary hypertension  
 pHi = intracellular pH  
 PI3K = phosphoinositide 3-kinase  
 PKA = protein kinase A  
 PKC = protein kinase C  
 PRA = plasma renin activity  
 ROS = reactive oxygen species  
 rvSUR1 = rat vascular sulfonyleurea receptor 1  
   RyR = ryanodine receptor  
   SBP = systolic blood pressure  
 SERCA = sarcoplasmic/endoplasmic reticulum Ca<sup>2+</sup> ATPase  
   SF = sulfidefluor  
   SFP = selective fluorescent probe  
   SHR = spontaneously hypertensive rats  
 siRNA = small interfering RNA  
 SMC = smooth muscle cell  
 SNAP = S-nitroso-N-acetylpenicillamine  
 SNP = sodium nitroprusside  
 SOD = superoxide dismutase  
 SPostC = H<sub>2</sub>S postconditioning  
 SPreC = H<sub>2</sub>S preconditioning  
 STAT = signal transducer and activator of transcription  
 TBARS = thiobarbituric acid-reactive substrates  
 TIMP = tissue inhibitor of matrix metalloproteinases  
   TNF $\alpha$  = tumor necrosis factor- $\alpha$   
   Trx1 = thioredoxin-1  
 TUNEL = terminal deoxynucleotidyl transferase dUTP nick end labeling  
   VEGF = vascular endothelial growth factor  
 VEGFR2 = VEGF receptor 2  
 VSMC = vascular smooth muscle cell  
 WKY = Wistar-Kyoto rats  
 XE991 = 10,10-bis(4-pyridinylmethyl)-9(10H)-anthracenone

**This article has been cited by:**

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